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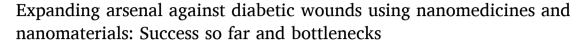
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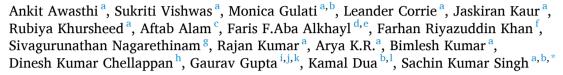
Journal of Drug Delivery Science and Technology

journal homepage: www.elsevier.com/locate/jddst



Review article





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ARTICLE INFO

Keywords: Diabetic wound Growth factors Nucleic acids Conventional treatments NDDS Nanomaterials

ABSTRACT

Diabetic wound (DW) is disease that occurs as a secondary complication of diabetes. The use of current therapies being employed to treat DW is associated with a number of limitations. However, the advancement in novel drug delivery system (NDDS) and nanomaterials in recent years has provided new options in the field of tissue rejuvenation. This review discusses the pathogenesis of DW, role of GFs and nucleic acids in DW healing, conventional treatments and need for NDDS. Various NDDS and nanomaterials reported for the treatment of DWs include liposomes, niosomes, transfersomes, exosomes, nanoemulsion, nano hydrogel, SLNs, NLCs, metallic NPs, polymeric NPs, dendrimers, wafers, hydrogel nanotubes, scaffolds, nanofibers and nanocomposites. Delving deeper, the clinical trials of NDDS based formulations and patents on treatment strategies of DW are also enlisted. The article has shown the potential of the aforementioned NDDS in treating DW, however, their clinical translation is hindered by certain issues such as their physicomechanical and storage instability, toxicity, poor drug loading and yield, poor site specificity and expensive manufacturing procedures. It is important to work on these bottlenecks to achieve better in vivo performance of formulation. This can be possible by the joint efforts of formulation scientists, biomaterial engineers, clinicians as well as regulatory officials. They have to understand the key areas that require more attention and that should be addressed for their better clinical translation.

1. Introduction

Diabetes mellitus (DM) is a multifactorial endocrine disorder characterized by glucose intolerance and hyperglycaemia. Lack of insulin or its inefficient action or both lead to development of DM [1]. Globally, DM is reported to be afflicting more than 422 million individuals [2]. With progress in time, DM leads to various complications such as

diabetic nephropathy, diabetic retinopathy and diabetic wounds (DWs) [3]. Global prevalence of these complications is reported as; DWs 42.2% [4], diabetic nephropathy 21.8% [5], diabetic retinopathy 3·4–12·3% while the rest includes other complications related to the cardiovascular system and neurological complications [6]. DW generally manifests as an open sore wound that can occur in any part of the body but it is most common in the foot and is known as a diabetic foot ulcer (DFU). Among



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the DWs occurring in various parts of the body, the global prevalence of DFU is 85% while the rest 15% manifest in the form of gangrene. The major attributing factors for DW/DFU are neuropathy, hypoxia, impeded angiogenesis and cellular damage due to release of reactive oxygen species (ROS) [7,8]. These, in turn, lead to vasculopathy, immunopathy and neuropathy that result in delayed wound healing sequences of defensive/inflammatory, proliferative and maturation phase. This delay in the healing process increases the chance of infection, which eventually may lead to amputation. Global percentage of amputation due to DW/DFU is reported to be 7-20% [9,10]. As the prevalence rate of DW/DFU is increasing rapidly, it is important to explore various treatment strategies for the condition. In this review, we critically discuss the pathophysiology of DW. A comprehensive overview of various treatment strategies currently in use against DW and their limitations is also provided. Finally, we highlight the potential of novel drug delivery systems in increasing the therapeutic activity of existing treatment options for combating DWs.

2. Pathogenesis of diabetic wound and diabetic foot ulcer

During hyperglycemia, the levels of miRNA-146a and miR-132 decrease while those of miR-155 increase. Suppression of miRNA-146a upregulates the proinflammatory mediators such as interleukin-1 receptor (IL-1R) associated kinase (IRAK1) and tumour necrosis factor (TNF) receptor-associated factor 6 (TRAF6). These extend the inflammatory phase and impede the wound healing process [11]. Reduction in the level of *miR-132* expression also delays the wound healing process by increasing the levels of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), nucleotide-binding oligomerization domain (NOD)-like receptor, toll-like receptors (TLR) and TNF signaling pathway. Increase in the levels of these proteins prolongs the inflammatory phase by releasing various inflammatory mediators from macrophages, monocytes and delays the wound healing process. Furthermore, miR-132 also targets the heparin-binding EGF-like growth factor (Hb-EGF) and facilitates the transition from the inflammatory phase to the proliferative phase [12]. On the other hand, overexpression of miR-155 leads to an increase in myeloperoxidase (MPO)-positive cells and decrease in angiogenic markers indicative of ECM build-ups such as TGF- β 1, collagen 1 and alpha-smooth muscle actin (α -SMA) [13]. It was found that an increase in the levels of miR-191 and miR-200b is positively associated with higher levels of inflammation-associated markers such as C-reactive protein (CRP) which result in reduced tube formation capacity, migration, and zona occludens-1 expression in human dermal endothelial cells and impede wound healing process [14]. Increase in the levels of miR-26a also influences the DW healing process by targeting SMAD-1 gene [Mothers Against Decapentaplegic Homolog1 (Drosophila)] and impairs wound edge angiogenesis and granulation tissue thickness. The other nucleic acids implicated in DW healing are miR-15b, miR-200 and miR-205-5p. Increase in levels of these nucleic acids results in the deactivation of vascular endothelial growth factor (VEGF) pathways and impairs the wound healing process [15]. Overall, all these factors lead to the formation of DW.

Hyperglycemia is known to result in idiopathic complications viz. neuropathy, immunopathy, and vasculopathy. Neuropathy affects autonomic nerves, motor nerves, and sensory nerves. In motor neuropathy, weakness and wasting of intrinsic foot muscles take place that finally leads to ulceration. In sensory neuropathy, there is a loss of pain leading to unnoticed trauma, which, in turn, may lead to ulcer formation. In autonomic neuropathy, sweating is decreased resulting in dry and brittle skin. This leads to secondary infections and finally causes ulceration. Vasculopathy is a general term used to describe any disease affecting blood vessels. It is categorized into two types i.e., macroangiopathy and microangiopathy. Macroangiopathy occurs due to the deposition of blood clots and fats in the blood vessels. Obstruction in the blood flow leads to tissue necrosis and finally, ulceration takes place. In case of microangiopathy, more glycoproteins are formed on the surface

of the basement membrane due to which vessel walls grow abnormally thicker and weaker leading to disruption of vessels. It causes leakage of blood and proteins and also slows down the flow of blood to different parts of the body. This is another cause of ulcer formation. In immunopathy, polymorpho-nuclear leukocyte migration, phagocytosis, chemotaxis and intracellular killing rate get decreased. Decreased chemotaxis of growth factors (GFs) and cytokines, coupled with an excess of metalloproteinases, impedes normal wound healing by creating a prolonged inflammatory state (Fig. 1a and b) [16,17].

3. Treatment strategies against the diabetic wound

Various molecules that have been successfully employed for the treatment of diabetic wounds are described below:

3.1. Growth factors (GFs)

On the onset of DW, platelets start moving towards the wound site to control the loss of blood and release matrix metalloproteinases (MMPs) as well as platelet-derived growth factor (PDGF). These help in the chemotaxis of macrophages and neutrophils towards the site of injury. Neutrophils help in the infiltration of the wound and secrete MMP-8, MMP-9, and fibronectin. In the next step, monocytes migrate towards the wound site and bind to the fibronectin. This results in macrophage differentiation, which secretes other GFs such as VEGF, transforming growth factor beta-1 (TGF- β -1), PDGF and fibroblast growth factor-1 (FGF-1). Release of TGF- β -1 stimulates the production of collagen as well as extracellular matrix (ECM) components that include fibronectin and hyaluronic acid. ECM provides tensile strength to the injured skin and aids in migration of keratinocytes towards the wound site. This helps in the healing of the wound [18,19].

Also, the inflammation and hypoxic environment in the DW cause activation of Angiotensin II (Ang II) which, in turn, results in the release of ROS, MMP-9 and MMP-2. All these, in turn, lead to cleavage of laminin-5. The resulting fragment binds to EGF receptors to promote fibroblast migration. The released FGF from macrophages binds to the FGF receptors of endothelial cells and heparan sulfate proteoglycan (HSP), resulting in the migration of keratinocytes. This promotes epithelization at the site of injury. Collagen binds to discoidin domain receptors (DDR) and integrin on fibroblasts, which stimulate the production of MMP-2, ECM remodelling, cell differentiation, and migration of fibroblasts [20,21]. This leads to healing of a wound.

3.2. Nucleic acid

A number of nucleic acids have shown remarkable potential in the field of tissue rejuvenation due to their effects on the biological behaviours of cells. They have been reported to exert their action on parameters like -inflammation, proliferation, migration and oxidation resistance [22]. Tetrahedral framework nucleic acids (tfNAs) which are DNA nanomaterial which synthesized from single-stranded deoxyribose nucleic acid (DNA) molecules are known to possess good stability, water solubility and biocompatibility [23]. These, tFNAs help in DW healing by activating Akt/Nrf2/HO-1 signaling pathway, increase nuclear factor erythroid 2-related factor 2 (Nrf2) levels and heme oxygenase-1 (HO-1) levels. Upregulation of these pathways results in angiogenesis, collagen deposition, oxidation resistance, neovascularization, epithelization and collagen alignment which, in turn, accelerates the DW healing [24]. Ganglioside-monosialic acid 3 (GM3) siRNA based spherical nucleic acids help in DW healing by GM3 synthase lockdown. It is pertinent to add here that GM3 inhibits cell growth and the function of GF receptors. They also increase the phosphorylation of IGF-1 receptor and epidermal growth factor and allow migration of keratinocytes at the site of injury [25]. TNF- α is a mediator of inflammation and its low blood levels promote normal wound healing. It is responsible for the migration and proliferation of fibroblasts and wound remodelling when present in a

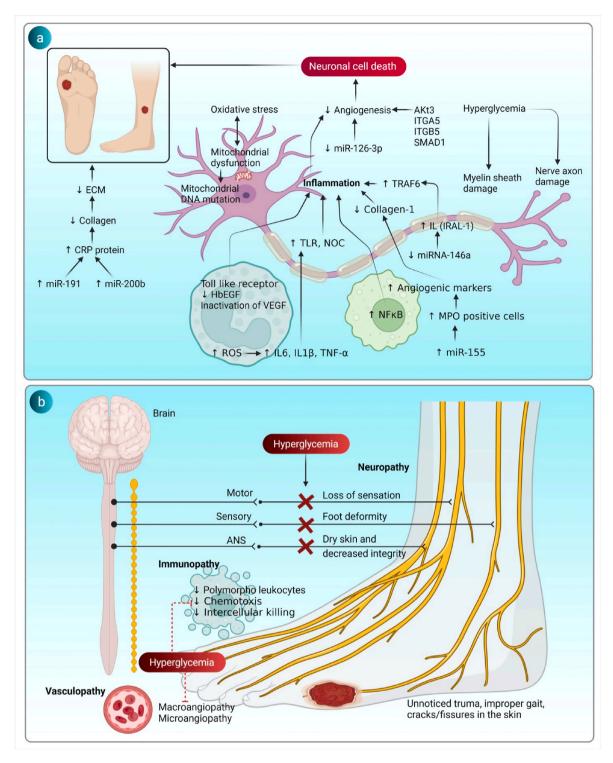


Fig. 1. Pathogenesis of a. DW; b. DFU.

small concentration. While the body is afflicted with DW, the levels of TNF- α get increased, which impedes wound healing by increasing cell apoptosis, matrix degradation and oxidative stress. TNF- α siRNA targets the TNF- α and lowers its level and promotes DW healing [26,27]. In DWs, levels of ROS are higher than normal, which results in oxidative stress. Due to oxidative stress, modification of Kelch-like ECH-associated protein 1 (Keap 1) takes place at some regions and prevents degradation of Nrf2. As a result of this, Nrf2 enters the nucleus and dissociates from the repressor site of Keap 1. Then Nrf2 binds to the antioxidant response element (ARE) in the promotor region and interrupts genes and enzymes

that are responsible for proteasome integrity, oxidation resistance, cellular protection and protein stability. Keap 1 *siRNA* lowers the levels of Keap 1, promotes degradation of NrF2 and decreases the cellular oxidative stress which helps in wound healing [28]. In DWs, there is also an activation of nuclear factor-kappa B, which results in the overproduction of pro-inflammatory cytokines such as interleukin IL-6, and IL-8 which impede the wound healing process by prolonging the inflammatory phase [29]. The *miR-146a* acts as a molecular brake in the inflammatory response. It inhibits nuclear factor kappa B by suppressing TNF receptor-associated kinase 6 (TRAF6) and interleukin-I

receptor-associated kinase 1 (IRAK 1) and downregulates overproduction of pro-inflammatory cytokines such as IL-6 and IL-8 and thus, promotes DW healing [30].

During normal conditions, the GFs and nucleic acids are available in our body to heal the wounds through the aforementioned mechanisms (Fig. 2.). However, during diabetes, these get decreased to levels that are insufficient to mediate wound healing. This causes a delay in wound healing [20,21]. To increase their levels, various GFs and nucleic acids may be administered externally. Some of the GFs that are widely used to treat DW include PDGF, TGF- β , recombinant human platelet-derived growth factor (rhPDGF) and placenta growth factor (PlGF) [30–34]. Nucleic acids used so far to treat DW are small interfering ribonucleic acid (siRNA), tFNAs, long non-coding RNA (lncRNA), sphingosine phosphate, and HoxD3 plasmid DNA [35–37]. A list of commonly used GFs and nucleic acids in DWs is given in Table 1 [36,37].

3.3. Synthetic drugs

Hyperglycemia is the one of the most important factors that contribute towards the susceptibility of DW patients for bacterial infections. Impairment of the T lymphocyte response, neutrophil function and humoral immunity is the starting point of this phenomenon. This results in reduction of inflammatory cytokines and polymorphonuclear leukocytes, which promote invasion of bacteria at the site of injury [38]. The most common bacteria found in DW patients are *Pseudomonas aeruginosa*, *Escherichia coli, Staphylococcus aureus* and *Proteus mirabilis* [39]. To contain the invasion of microorganisms at the DWs site, antibiotics

are used [40]. Commonly used antibiotics used for this purpose include vancomycin, ceftazidime, piperacillin/tazobactam, amoxicillin-clavulanate, clindamycin, ampicillin-sulbactam and ciprofloxacin [41]. Diabetic patients with clean ulcers but showing positive ulcer swabs are recommended for an early antibiotic treatment. As DW is susceptible to repeat infection and untreated infection can lead to amputation, this cycle needs be broken only with antibiotics [42]. Antibiotics do not heal the wound but decrease the microbial load on DW by different mechanisms which are discussed in the subsequent sections.

Vancomycin exerts its antibacterial effect by acting on the bacterial cell wall and inhibiting the peptidoglycan polymerization. This inhibition leads to leakage in the bacterial cell wall and causes bacterial cell death [43]. In one of the studies, Davani et al., revealed that vancomycin nanofibers when used against DW microorganisms (Methicillin resistant Staphylococcus aureus (MRSA), S. aureus, Escherichia coli and P. aeruginosa. They exhibited antimicrobial action against S. aureus and MRSA with zone of inhibition of 2.9 and 2.5 cm and gram-negative bacteria species of E. coli and P. aeruginosa with zone of inhibition of 1.9 and 2.8 cm, respectively [44]. It is available in the market under the name Vancocin (ViroPharma). It is used in parenteral and oral form to treat DW. Its recommended dose to treat DW is 10–15 mg/kg (up to 1g) in divided doses [45]. Ceftazidime shows its antibacterial activity against penicillin-binding protein-3 (PBP-3), which results in filamentation and cell lysis of P. aeruginosa and E. coli. It also helps in the inhibition of PBP-1 α and PBP 1 β that induces cell elongation [46,47]. It is available in the market under the name of Fortaz (GlaxoSmithKline) and administered parenterally to treat DW. Its dose to treat DW is 500

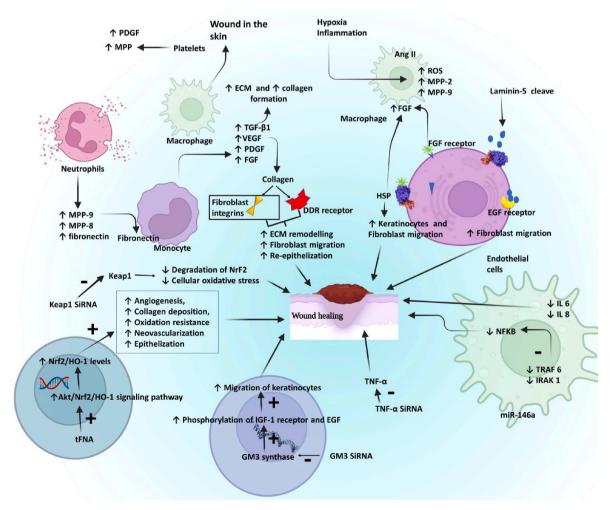


Fig. 2. Role of growth factors and nucleic acids in diabetic wound healing.

Table 1 Various treatments available for diabetic wounds.

Sr. No.	Therapeutics	Formulation	Animal model used	Key findings	Referen
Fs					
	TGF-β and Insulin-like Growth Factor-I	Gel	STZ induced male SD rats	50% greater wound tensile strength as compared to collagen vehicle alone	[30]
•	Human epidermal growth	Cream	-	2.25 folds increase in wound healing as compared to control groups	[31]
	Placenta growth factor	-	STZ induced C57Bl/6 male mice	1.68-folds increase in granulation tissue area as compared to saline- treated groups	
•	Recombinant human platelet-derived growth factor	Gel	Male Wistar diabetic rats	1.14-folds increase in re-epithelization as compared to vehicle-treated groups	
	PDGF	Gel	STZ induced male Wistar rats	1.41-folds increase in cell proliferation as compared to a vehicle control group	[32]
uclei	ic acids				
	HoxD3 plasmid DNA	Methylcellulose pellets	Genetically diabetic db/db mice	↑collagen deposition and angiogenesis	[37]
	Sphingosine phosphate	Injection	db/db diabetic mouse	3.75-folds increase in angiogenesis as compared to control groups	[36]
•	IncRNA	-	ICR male Swiss mice	Activate hypoxia-inducible factor-1a signaling pathway and increased fibroblast migration and collagen deposition at the site of injury	[35]
•	tFNAs	-	Alloxan induced male Wistar diabetic rats	2.19-folds increase in epidermal thickness as compared to a diabetic control group	[22]
vnthe	etic drugs				
	Acidified nitrite Azelnidipine	Cream Solution	Lepr db diabetic mice Diabetic rats	1.11 -folds increase in wound closure as compared to control groups \uparrow collagen fibers, fibroblast density and angiogenesis as compared to	[83] [59]
	Simvastatin	Ointment	db/db diabetic mice	control groups 1.28-folds increase in wound contraction as compared to control	[84]
	Tocopherol	Cream	STZ induced male SD rats	* *	
•	Hydrogen sulfide	Ointment	STZ induced male SD rats	•	
	Dropropolol	Cream	Eomala diabatia misa	diabetic control groups	[87]
	Propranolol Naltrexone	Cream	Female diabetic mice STZ induced male SD rats	4-folds decrease in wound area as compared to control groups 1.93-folds increase in percentage VEGF + vessels as compared to Regranex treated groups	
	Grotto cream + Fucidin cream	Cream	STZ induced male Wistar rats	1.07-folds increase in epithelization as compared to grotto cream treated groups	[89]
 Ierba	ıl drugs				
	Panchavalkala	Cream		↑ antimicrobial and anti-inflammatory action	[90]
	Acheflan	Cream	STZ induced SD diabetic rats	† angiogenesis, collagen deposition and hydroxyproline content. Exhibited 1.10-folds increase in hydroxyproline content as compared to control groups	[91]
	Aloe vera	Ointment	STZ induced male Wistar rats	1.14-folds increase in wound closure as compared to marketed formulation	[92]
	Hypericum perforatum	Gel	STZ induced Female Wistar rats	†collagen deposition, antibacterial activity, antioxidant activity and re- epithelization	[93]
•	Chrozophora tinctoria	Leaves extract	STZ induced Albino Wistar rats	1.34-folds increase in collagen content as compared to control groups	[94]
	Rosa Damascena Mill	Lotion	Diabetic rats	1.11-folds increase in wound closure as compared to control groups	[95]
	Euphorbia hirta linn	Ointment	Alloxan induced female Swiss albino diabetic rats	1.7-folds increase in wound contraction as compared to control groups	[96]
	Curcumin	Cream	Inbred diabetic male C57BL/6 mice	↑ neoangiogenesis and exhibited about 1.56-folds increase in angiogenesis as compared to placebo-treated groups	[97]
em c	cells			5 5 1 1 1 br.	
	Bone marrow-derived MSC Autologous Keratinocytes	Gel Cell suspensions	db/db diabetic mice STZ induced female	\uparrow cytokines production and stimulation of endogeneous resident cells 1.6-folds increase in epithelization rate as compared to normal saline-	[98] [99]
	and fibroblasts		diabetic pigs	treated groups	
	ADMSC	Intra-dermally	STZ induced male SD rats	1.04-folds increase in collagen fibers	[100]
	Embryonic stem cells	Injection	STZ induced male SD rats	3.3-folds decrease in wound size as compared to control group	[101]
	Human Wharton's jelly stem cells	Jelly	SCID mice	1.18-folds increase in wound closure as compared to control groups	[102]
	Allogenous skin fibroblasts	Cell suspensions	Alloxan induced diabetic	1.16-folds increase in wound contraction as compared to control group	[103]

Abbreviations ADMSC; Adipose derives mesenchymal stem cells, *lncRNA*; *long noncoding RNA* TGF-β; Transforming growth factor-beta, *tFNAs*; Tetrahedral framework nucleic acids.

mg to 1 g by intravenous (i.v) or intramuscular (i.m) route every 8 h [48]. Piperacillin, acts on bacterial cell wall and binds to specific PBPs. It also inhibits the third and last stage of bacterial cell wall synthesis. Cell lysis is then mediated by bacterial cell wall autolytic enzymes such as autolysins [49]. Anneke et al., conducted a clinical trial on use of piperacillin/tazobactam treatment in 25 DW patients with *P. aeurogonisa, E. coli* and *S. aureus* load at the site of injury. The

piperacillin/tazobactam treatment showed antibacterial action against these microorganisms with an average zone of inhibition of 18 mm and 22 patients showed no sign of bacterial load at the wound site. It is available in the market under the name of Tazocin® and Zosyn® and is administered both orally as well as i.v. A combination of piperacillin 4g and tazobactam 0.5g is i.v. administered 3 times a day to treat DW [50, 51]. Amoxicillin exerts its antimicrobial effect by acting on bacterial cell

wall and results in bacterial lysis. It is used in superficial and neuroischemic ulcers to combat staphylococcal and streptococcal infections [52]. It is available in the market under the name Augmentin® and is given through oral route. Its dose to treat DW is 500-875 mg every 8-12 h peroral [42]. Clindamycin binds to the 50s ribosomal subunit, which results in inhibition of protein synthesis. It is given in the oral form to treat DW. It is available in the market under the name Cleocin®. Its oral dose to treat DW is 300 mg Q6H [53]. Ampicillin sulbactam binds reversibly to the 30s ribosomal subunit which results in the inhibition of protein synthesis and shows a bacteriostatic effect. It is given either orally i.v. It is available in the market under the name Unasyn®. Its dose to treat DW is 3g every 6 h through iv route [52]. Ciprofloxacin acts on bacterial DNA-gyrase and DNA topoisomerase, and as a result of this, replication of DNA does not take place. It is given orally as well as in i.v. form. It is available in the market under the name Cipro®. Its dose to treat DW is 750 mg BD (oral) [54]. Doxycycline binds reversibly to the 30s ribosomal subunit which results in the inhibition of protein synthesis and shows a bacteriostatic effect. It penetrates the cell wall of bacteria and binds to PBPs and results in bacterial cell death. The bactericidal effects result in inhibition of cellular growth and division as well as loss of cell wall integrity, eventually causing cell wall lysis [52]. It is available in the market under the name WELL-DOX 100. Its dose to treat DW is 100 mg BD (oral) [54]. . However, overuse of antibiotics has negative effects for the patient and may promote the problem of antibiotic resistance. In diabetic foot infections antibiotics are to treat infection, not to heal wounds [55,56].

In recent years, some of the existing drugs have been repurposed for treating DW owing to some of their mechanisms of action that are involved in treating the disease. These drugs include HMG CoA reductase inhibitors e.g. mevastatin, simvastatin [57] and atorvastatin [58], biguanides e.g. metformin and calcium channel blockers (CCBs) e.g. azelnidipine [59]. HMG CoA-reductase inhibitors such as meyastatin, simvastatin and atorvastatin help in inhibition of cholesterol synthesis. This leads to the inhibition of farnesyl pyrophosphate (FPP) and cortisol which, in turn, causes increase in epithelization and migration of keratinocytes at the site of injury. Furthermore, statins also exhibit anti-inflammatory, antioxidant and antibacterial actions and promote wound healing process. The biguanides such as metformin help in wound healing by showing inhibition of mTOR and NOD-like receptor protein 3 (NLRP3) which further activate the AMPK pathway. This leads to increase in the polarization of macrophages towards the phenotype M2 and accelerate wound healing process [60]. In addition, (CCBs) such as, azelnidipine help in wound healing by showing antioxidant action as well as enhancing endothelial NO synthase (eNOS)-catalyzed NO production which improves vascular endothelial function. Furthermore, CCBs also decrease the expression of nicotinamide-adenine-dinucleotide phosphate (NADPH) oxidase responsible for production of superoxide (O_2^{-1}) [59]. These drugs are administered in the form of tablets, capsules, gels, ointments and solutions to treat DWs. Advantages of these conventional drugs include a quick onset of action, well-known safety and clinical profiles, cost-effectiveness and clinical applicability. A list of some synthetic drugs that have been explored to treat DW are presented in Table 1.

3.4. Herbal drugs

Many herbal drugs possess anti-inflammatory as well as antioxidant potential. These can quench the free radicals and reduce oxidative stress. Inflammation and oxidative stress are the root causes of DWs. These can be explored for their potential in the treatment of the DWs owing to their anti-inflammatory and antioxidant potential [61]. Some of the plants in their crude form or in the form of extracts are already reported for their use in DWs since ancient times. These include *Trigonella foenum-graecum*, *Vitis vinifera*, *Rubus fruticosus*, *Prosopis farcta*, *Cinnamomum zeylanicum*, *Ferula assafoetida*, *Melilotus officinalis*, *Hypericum perforatum* [62], *Cordia verbenacea* [63], *Aloe vera* [64], *Chrozophora tinctoria* [65], *Rosa*

damascena Mill [66] and Euphorbia hirta linn [67], Radix rehmanniae [68], Carica papaya [69], Punica granatum [70], Curcuma longa [71], Allium cepa L. [72], Phoenix dactylifera [73], Vitis vinifera [74], Citrus aurantium L [75], Martynia annua [76] and Annona squamosa [77]. The isolated phytoconstituents that are extensively used to treat DW include flavonoids such as curcumin [78], quercetin [79], hesperidin [80] and luteolin [76], glycosides such as aloenin [81], naringin [74] and carbohydrates such as beta-glucan [82]. A list of herbal drugs that have been utilized to treat DW is given in Table 1.

4. Need for novel drug delivery systems and their mechanism of drug release

Various treatment strategies mentioned in the previous sections of the manuscript to treat DWs exhibit certain limitations related to their delivery. For example, GFs get degraded by the enzyme protease present at the wound site upon topical application at the wound. Large molecular size, short half-life and toxicity at higher systemic doses showed that the conventional treatment of GFs in their free form is not suitable to deliver them successfully at the wound site. Similarly, nucleic acids such as siRNA, upon topical application at the wound get degraded by the enzyme nuclease present at the wound site. Also, as siRNAs are negatively charged, they do not penetrate the negatively charged cells due to electrostatic constraints. Herbal drugs given through the conventional route also possess some inherent challenges similar to those discussed in above sections for synthetic drugs such as poor stability, solubility, permeability as well as target specificity. To overcome these limitations, various research endeavours have been made for encapsulating/complexing these both natural as well as synthetic drugs used for wound healing, using novel drug delivery system to deliver them through topical route. Advantages of using NDDS include closer interactions with the biological target, increased penetration at the wound site and sustained delivery of encapsulated drugs [104,105]. Some of the NDDS extensively explored to treat DW include polymeric nanoparticles, inorganic nanoparticles [106], nanohydrogels [107], liposomes [108], nanoemulsions [109], lyophilized wafers [110], solid lipid nanocarriers (SLNs) [111] and nanostructured lipid carriers (NLCs) [112]. Another significant advantage of non-invasive topical delivery of these NDDS is the increase in patient compliance due to convenience. They can enhance the permeability of drug across the skin via transcellular, paracellular and endocytosis, provide target specificity and release the drug at the wound site. In transcellular route drug directly passage from the lipid layers whereas, in paracellular route drug reaches at the target site by passing through the tight junctions present in the skin. In endocytosis drug reaches the wound site by entering through the pores present on skin. However, these nanocarriers show short retention time at wound site, leading to limited success of the aforementioned nanocarriers in treating DW [107-112].

To achieve sustained release and better localization at the site, these NDDS have generally been prepared by incorporation into nanomaterials such as nanofibers [113], scaffolds [114] and nanocomposites [115]. These nanomaterials are versatile in nature, possess controlled size and tunable physicochemical properties. They also offer a larger surface area to volume ratio to allow cell adhesion of the drug for the desired time period [116]. The various nanocarriers and nanomaterials used to treat DW and their mechanism of drug release at the wound site are shown in Fig. 3.

Despite having many advantages of NDDS and nanomaterials they are associated with some limitations which are given in Table 2.

4.1. Vesicular drug delivery systems (VDDS)

VDDS are defined as highly ordered assemblies comprising of concentric lipid bilayers with encapsulation of active moieties in their lipid or aqueous layer (95). They are generally formed by self-assembling of the amphiphilic building blocks on coming in contact

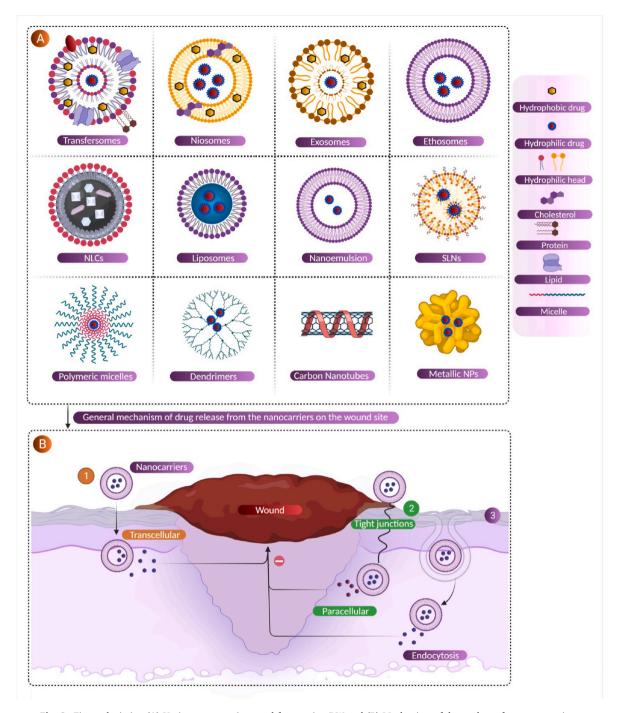


Fig. 3. Figure depicting (A) Various nanocarriers used for treating DW and (B) Mechanism of drug release from nanocarriers.

with an aqueous medium [117]. The VDDS reported so far to treat DW include liposomes, exosomes, niosomes and transfersomes. These are discussed below:

Liposomes are spherical vesicles composed of one or more partially substituted phospholipid bilayers along with cholesterol (CHL). They consist of hydrophilic aqueous core and hydrophobic phospholipid bilayer due to which they can encapsulate both hydrophilic and lipophilic drugs. This composition of liposomes facilitates their use in encapsulating a range of drug payloads, due to which their application in drug delivery systems is increasing rapidly. In addition to this, liposomes have gained importance as potential drug delivery systems owing to their stable lipid bilayer membrane, minimized enzymatic degradation, biocompatibility and cell-specific targeting [118,119]. These have

been also explored as carriers to treat DW by surface functionalization of therapeutic moieties such as collagen mimetic peptide (CMP) (97) and RGPK (peptide) (100). In one of the research works Olekson et al., have investigated the effect of stromal-derived factor-1 (SDF-1) liposomes against DW in streptozocin (STZ) induced BKS.Cg-Dock7m1/1Lepr (db)/J genetically modified diabetic mice. In this study, the percentage wound closure of SDF-1 liposomes was evaluated and results were compared with free SDF-1. The results revealed that the SDF-1 liposomes treated group showed 15% more wound contraction within 21 days as compared to free SDF-1. SDF-1 liposome treated group demonstrated an increase in diameter of CD31⁺ luminal structure as well as resident cell proliferation [120]. Similarly, Choi et al. investigated the effect of cationic elastic liposomes (CELs)-low molecular weight protamine

Table 2Advantages and disadvantages of NDDS and nanomaterials.

Delivery system	Advantages	Limitations	References	
Liposomes	Increased the therapeutic index and efficacy of the drug entrapped Increased the drug stability by entrapping in its core Non-toxic, biodegradable and flexible to tailor Helpful in reducing toxicity of therapeutics Target specific drug delivery	 It consists of phospholipids which undergo hydrolysis and oxidation The cost of production is high The issue of fusion and leakage of encapsulated drug/molecule 	[108]	
Nanoemulsion	Nanoemulsion is a best alternate for vesicles and liposomes It is helpful in improving bioavailability of drugs Non-irritant and non- toxic It improves physical stability of drugs It increases solubility of lipophilic drugs It is used in formulating many formulations like foams, spray and creams	 Require high amount of surfactants and co-surfactants Associated with surfactant induced toxicity risk Stability related concerns pertaining to change in temperature and pH of the system 	[109]	
Nanohydrogels	absorption Targeted drug delivery Controlled release of bioactive substances High drug encapsulation capacity Less toxicity Size tunable property Ease of preparation	 High cost Difficult to handle Low mechanical strength 	[107]	
Solid lipid nanocarriers (SLNs)	Provide targeted and controlled release property Capacity of carrying both hydrophilic and lipophilic drugs in its core Bio-compatible and bio-degredable in nature Cost effective or economic	 .Poor drug loading capacity The dispersions consist of high water content (70–90%) Due to polymeric transitions, explusion of drug takes place during storage 	[111]	
Nano lipid carriers (NLCs)	The ability to enhance storage stability Increases the solubility of drugs in their lipid matrix Improved permeability and bioavailability Impart higher adhesive property by reducing water loss from skin Tissue targeted delivery	Chances of skin irritation specially when fatty acids used in formulating NLCs	[112]	
Scaffolds	 Good biocompatibility and biodegradability Good bioresorption Overcome the limitation of hydrophobicity of several therapeutics Helps in tissue regeneration Non- toxic in nature 	 Difficult to process while preparing Highly brittle Hard to maintain stability for a longer period of time 	[114]	

Various NDDS explored for treatment of DWs are described below.

(LMWP)-GFs-hyaluronic acid (HA) against DW in STZ induced female C57/BL6 diabetic mice in which they found that on 11th day, CELs-LMPW-GFs-HA treated groups exhibited 2.84-folds and 2.4-folds decrease in wound area in comparison to diabetic control and HA treated groups. In addition, CELs-GFs-HA treated groups exhibited about 1.2-folds increase in re-epithelization thickness as compared to vehicle (CELs) treated groups indicating the DW healing potential of formulation [108]. In another study, Rabbani et al., studied the effect of Keap1 siRNA liposomes against DW in male Lepr db/db diabetic mice where they observed significant enhancement in wound healing potential (1.47-folds) as compared to siNS treated group [121]. In another study, Chhibber et al. reported the effect of bacteriophage entrapped liposomes (LCP) against DW in an alloxan-induced BALB/c diabetic mice model where they found six days faster wound healing rate in comparison to free cocktail phage (FCP) treated groups due to enhanced collagen deposition and re-epithelization at the site of injury indicating the wound healing potential of the formulation [122]. In one of the study, effect of collagen mimetic peptide (CMP)-vancomycin (Van) loaded liposomes-based co-gel was reported against methicillin-resistant S. aureus (MRSA) wounds in BALB/c mice model in which they found 7-folds faster antibacterial activity as compared to CMP-Van-Lipo treatments alone [123]. Similarly, in a study on the effect of cinnamon extract liposomes against DW in alloxan-induced monohydrate induced albino rats, the topical administration of cinnamon extract liposomes exhibited about 1.10-folds and 1.11-folds decrease in wound diameter as compared to chitosan and imipenem treated groups respectively [124]. In another study, Bhattacharyya et al., studied the effect of co-administration of RGDK (small peptide)-lipopeptide-1 and

recombinant human platelet-derived growth factor-B (rhPDGF-B) lipoplex against DW in STZ induced SD diabetic rats. The results revealed that RGDK lipopeptide-1 and rhPDGF lipoplex co-administration accelerated DW healing within 10 days by 1.8-folds and 2.3-folds as compared to RGDK lipopeptide-1 alone and RGDK lipopeptide-2 co-administered with rhPDGF lipoplex, respectively The accelerated wound healing was attributed to enhanced keratinization, epithelization, angiogenesis and fibrocollagenation activity of RGDK lipopeptide-1 co-administered with rhPDGF lipoplex at the site of injury [125].

To overcome the problem of poor elasticity associated with liposomes which leads to their poor permeation, a newer class of VDDS i.e., "transferosomes" has been introduced with slight modifications in their chemical composition. Transfersomes are defined as highly deformable and elastic vesicular structures comprising of phospholipids, surface and edge activators, alcohol and buffering agents [126]. Edge activators are bilayer softening component in which amphiphilic drug is incorporated in order to increase lipid bilayer flexibility and permeability. These include Span 65, Tween 80, Tween 60 and sodium deoxycholate. They are known to alter the interfacial tension within the system. Penetration ability of transfersomes makes them an ideal carrier for target specificity at the local region of the DW. However, the major drawback of this VDDS is their low stability due to predisposition towards oxidative degredation and their high cost [127]. The study on transfersomes based delivery system for DW is very limited. Only one study is available on transfersomes for the treatment of DW which is discussed below.

Gizawy et al., studied the effect of deferoxamine (DFO) transfersomes based gel against DW in STZ induced adult male Wistar

diabetic rats where they found 1.36- folds and 3.09-folds increase in VEGF levels as compared to DFO solution and plain Na-CMC gel respectively. Furthermore, DFO transfersomes based gel showed 1.42-folds and 2.08-folds decrease in TGF- β levels as compared to that of DFO solution and plain Na-CMC gel respectively. The collagen study revealed that DFO transfersomes based gel showed 1.5-folds and 1.9-folds increase in collagen deposition as compared to that of DFO solution and plain Na-CMC gel respectively [128].

Niosomes are modified form of liposomes consisting of non-ionic surfactants such as polyoxyethylene fatty acid esters and CHL. Niosomes may be unilamellar or multilamellar depending upon the preparation technique. Presence of non-ionic surfactant in niosomes helps in the alteration of the structural arrangements of stratum corneum (SC) leading to enhanced permeation across the membrane. The better chemical stability and deeper permeability of these nanocarriers have led to their use in the treatment of DW which is discussed below.

El-Ridy et al., studied the effect of metformin loaded niosomes against DW in STZ induced adult male Wistar rat diabetic model. Topical application of metformin niosomes accelerated DW healing within 2 weeks by stimulating serum TGF- β -1, collagen deposition and antidiabetic action at the site of injury as compared to that on treatment with oral metformin. In addition to above observation, reduction in blood glucose level of rats treated with metformin niosomes and oral metformin were also observed [129].

Exosomes are endosomal derived nanovesicles that help in the intercellular transportation of materials. Therapeutics such as proteins, nucleic acids and drugs can be incorporated into these nanovesicles and can be targeted to the specific site [130]. Exosomes are associated with various advantages such as amenability to gene and drug delivery, convenient storage and stability in the body. Furthermore, exosomes help in the proliferation, migration and angiogenesis process in the wound environment [131]. Looking at these benefits, a number of drugs in the form of exosomes have been used for the treatment of DW are discussed below.

In one of the study, Yu et al., explored the effect of atorvastatin (ATV) pretreated MSC exosomes against DW in STZ induced male SD rats in which they found 1.31-folds and 1.66-folds increase in blood vessels area as compared to control and exosomes alone treated group respectively. In addition, ATV pretreated MSC exosomes exhibited about 1.06folds and 1.12-folds increase in percentage wound closure within 14 days as compared to control and exosome alone treated group respectively [132]. In another study, co-administration of Curcuma zedoaria polysaccharide (ZWP) and platelet-rich plasma exosomes (PRP-Exos) assembled chitosan/silk hydrogel sponge (ZWP-PRP-Exos sponge) was studied on DW in STZ induced female SD diabetic rats in which they found that on 15th day, ZWP/PRP-Exos sponge treated groups demonstrated 1.02-folds, 1.34-folds and 2.05-folds increase in wound closure rate as compared to diabetic control, diabetic group/100 µg PRP-Exos and diabetic/ZWP treated groups respectively [133]. Similarly, Qijun et al., explored the effect of miR-21-5p exosomes against DW healing in STZ induced male SD rats for a period of 15 days. miR-21-5p exosomes accelerated DW healing by showing a 1.9-folds increase in re-epithelization as compared to diabetic control groups [134]. In another study, effect of long noncoding RNA (IncRNA)-H19 exosomes were tested against DW in STZ induced male C57BL/6J mice revealed that the IncRNA H19 exosomes accelerated DW healing within 13 days by showing decreased apoptosis and inflammation of fibroblasts by inhibiting phosphatase and tensin homolog (PTEN) protein. Additionally, IncRNA H19 exosomes exhibited about 1.33-folds increase in wound closure as compared to negative control groups [135]. In one of the study, effect of miRNA-221-3p exosomes were evaluated against DW in STZ induced male C57BL/6 mice in which they 1.6-fold increase in wound contraction as compared to diabetic control groups indicating wound healing potential of the formulation [136].

The VDDS used so far to treat DW healing are summarized in Table 3.

4.2. Nanoemulsion

Nanoemulsion (NE) is a thermodynamically stable isotropic mixture of oil, emulsifier, and water. The oil phase used in NE is generally made up of free fatty acids, monoacylglycerol, diacylglycerol, and triacylglycerol [151], while the aqueous phase consists of co-solvents and polar solvents [152]. The most commonly used stabilizers are emulsifiers (surfactants) which help in reducing the interfacial tension between the two immiscible liquids (oil and water) and provide stability to NE [153]. The surfactants used in NE may be non-ionic and ionic. Non-ionic surfactants oppose the aggregation of droplets by showing hydration, steric hindrances, and thermal fluctuation interactions. In contrast, ionic surfactants prevent agglomeration by electrostatic repulsion [154]. Some of the recent studies in which NE are used to treat DW healing are discussed in subsequent sections.

In one of the study, Kaur et al., reported the effect of sea buckthorn (Hippophae rhamnoides L.) seed oil NE gel against DW in STZ induced male Wistar diabetic rat model in which they found 1.49-folds increase in hydroxyproline content as compared to sea buckthorn oil indicated the wound healing potential of the formulation [155]. In another study, effect of astaxanthin and alpha-tocopherol conjugated κ-carrageenan NE (AS-TP@KCNE) was studied against DW in STZ induced adult ICR (Institute of cancer research) female mice diabetic model, it was found that on the 15th day the percentage of wound contraction of prepared formulation was found to be 1.3-folds higher than that of diabetic control group respectively [156]. Similarly, Jee et al., studied the effect of LMWP-GFs/quercetin/oxygen-perfluorooctyl bromide-(LMWP-GFs/QCN/OXY-PFOB-NE-GEL) against DW in STZ induced female C57BL/6 diabetic mice model in which they found 1.42-folds, 1.13-folds and 1.08-folds increase in wound contraction as compared to QCN-NE-GEL, LMWP-GFs-GEL and OXY-PFOB-NE-GEL respectively [157]. In one of study, the effect of topical application of boronophenylalanine (BFA)-Zn-containing NE was studied against DW in STZ induced male SD diabetic rat model where they observed significant enhancement in DW healing potential by 2-folds, 1.3-folds and 1.1-folds as compared to placebo NE, Zn NE and diabetic control groups respectively [158]. Similarly, Chua et al., studied the effect of palm glyceryl monocaprylate (GMCY) NE against S. aureus infected wounds in a male adult SD rat model. Topical application of GMCY NE exhibited DW healing within 14 days by promoting antibacterial action against E. coli and S. aureus. On the 14th day, the percentage of wound closure was found to be 80% and 75% for GMCY NE and placebo NE respectively [159]. In another study, the effect of sesame oil NE (SONE) containing levofloxacin (SONEL) was reported against DW in STZ induced male albino Wistar rat diabetic model where they exhibited 2-folds, 2.3-folds, 1.97-folds, 1.06-folds and 2.57-folds increase in blood vessels and 2.33-folds, 4.44-folds, 2-folds and 5.1-folds decrease in inflammatory cells as compared to SONE, SSD, D-NC and non-diabetic control group (ND-NC) respectively [160].

4.3. Nanohydrogels

Nanohydrogels are combinations of nanoparticulate system and hydrogels. They can be prepared by using polymers and crosslinking agents. Common polymers used in the preparation of nanohydrogels are N-vinyl-2-pyrrolidone (NVP), methoxy ethoxy ethyl acid (AA), ethylene glycol (EG), polyethylene glycol methacrylate (PEGMA), polyethylene glycol dimethacrylate, N-(2-hydroxypropyl) methacrylamide (HPMA), ethylene glycol dimethacrylate (EGDMA), hydroxy ethoxy ethylmethacrylate (HEEMA), hydroxyethyl methacrylate (HEEMA), hydroxyethyl methacrylate (HEEMA), hydroxydiethoxyethylmethacrylate (HDEEMA) [161,162]. Commonly used crosslinking agents in the preparation of nanohydrogels include $N_0\text{-Methylene-bis-acrylamide}$ (N, $N_0\text{-MBAAm}$) (BIS), epichlorohydrin (ECH) and divinyl sulfone (DVS) etc [163,164]. Some of the recent studies in which nanohydrogels are used to treat DW healing are discussed in subsequent sections.

Table 3
List of various vesicular drug delivery systems used so for to treat diabetic wounds.

Sr. No.	VDDS	Therapeutic moiety	Method of Preparation	Animal model	Research highlights	Reference
1.	Liposome	Haaemoglobin	Ultracentrifugation followed by precipitation	dB/dB mice	 ↑ 4.7-folds wound contraction at 7^h day ↓ cell apoptosis and inflammatory cascade and matrix metalloproteinase-9 (MMP-9) ↑ granulation formation 	[137]
2.	Liposome	α-gal	Thin lipid film hydration	Diabetic mice	↑ 3.03-folds wound contraction within 12 days ↑ vascularization, epithelization and	[138]
3.	Exosome	Human umbilical cord blood-derived EPCs	Ultracentrifugation	SD rats	granulation tissue deposition • ↑ 1.6-folds wound contraction within 14 days • ↑ collagen deposition, re-epithelialization	[139]
4.	Exosome	Menstrual blood-derived MSCs	Ultracentrifugation	Male C57BL/ 6 mice	and less scar formation $ \bullet \uparrow 1.82 \hbox{-folds wound contraction within } 12 \\ days$	[140]
5.	Exosome	Macrophage-derived Ultracentrifugation Exosomes		SD rats	↑ migration of epithelial cells ↑ 1.17-folds wound contraction within 14 days ↑ reepithelization, collagen deposition and	[141]
6.	Exosome	Oxidative hyaluronic acid	Differential centrifugation	Diabetic mice	angiogenesis • ↑ 1.37-folds wound contraction within 14 days • • ↑ reepithelization, collagen deposition,	[142]
7.	Exosome	Polyurethane + Calcium peroxide	Ultrafiltration	Diabetic rats	antimicrobial action and Angiogenesis 1.06-folds wound contraction within 14 days	[143]
8.	Exosome	Engineered Human adipose stem-cell	Ultracentrifugation	SD rats	↑ collagen deposition and angiogenesis ↑ 1.39-folds wound contraction within 15 days ↑ re-epithelialization, angiogenesis and	[134]
9.	Exosome	Human Nrf2-active multipotent stromal cells	Differential ultracentrifugation	Adult Lepr ^{db/}	collagen remodelling action • DW healing within 14 days • ↓ epithelial gap • ↑ granulation and density of CD31 ⁺	[144]
10.	Exosome	mmu_circ_0000250-modified adiposederived stem cells	Centrifugation followed by Ultrafiltration	Male C57BL mice	vessels • DW healing within 14 days • microvascular development	[145]
11.	Exosome	hAECs	Ultracentrifugation	Male db/db mice	 ↓ skin tissue apoptosis ↑ 2.43-folds wound contraction within 14 days ↑ collagen deposition 	[146]
12.	Exosome	Dia-Exos AntagomiR-15a-3p	Ultracentrifugation	C57BL/6J mice	 ↑ in capillary density Anti-miR-15a-3p activity ↑ blood perfusion, collagen synthesis, angiogenesis and fibroblast proliferation 	[147]
13.	Exosome (MSCs- Exo oe-H19)	MSCs	Ultracentrifugation	Male C57BL/ 6J mice	 † 1.3-folds wound contraction within 13 days † collagen I, VEGF, transforming growth factor-beta (TGF β-1) 	[148]
14.	Exosome	Melatonin	Ultracentrifugation	SD rats	↓ cell apoptosis ↑ 1.08-folds wound contraction within 14 days ↑ anti-inflammatory action, angiogenesis	[149]
15.	Exosome	PUAO-CPO	Ultrafiltration	Diabetic rats	and collagen deposition • † 1.06-folds wound contraction within 14 days	[150]
16.	Exosome	HUC MSCs exosomes encapsulated polyvinyl alcohol/silver nano hydrogel	Ultracentrifugation	SD rats	 † collagen deposition and angiogenesis † 1.6-folds migration area within 18 days † formation of epithelium, collagen bundle fibers and hair follicles 	[107]

Abbreviations: EPC; endothelial progenitor cells, hAECs; Human amniotic epithelial cells, HUC; Human umbilical cord, MSCs; Mesenchymal stem cells, PUAO-CPO; Polyurethane-Calcium peroxide.

A study was carried out to investigated effect of fibroblast growth factor-2 (FGF-2)/chitosan hydrogel against DW in impaired db/db diabetic mice. On the 10th day, FGF-2/chitosan hydrogel exhibited 1.4-folds wound closure as compared to chitosan treated groups due to neovascularization and angiogenesis at the site of injury [165]. In another study, effect of transforming growth factor- $\beta 1$ (TGF- $\beta 1$) hydrogel was checked against DW in female Lepr db diabetic mice. The results showed that TGF- $\beta 1$ hydrogel was found to exhibit about 1.28-folds increase in proliferating cells at the site of injury as compared

to placebo hydrogel treated groups [166]. In one of the studies, potential of recombinant human epidermal growth factor (rhEGF) hydrogel against DW in male SD diabetic rats. Not much advantage could be achieved in this case when compared with rhEGF solution treated as the increase in wound contraction was found to be only by a factor of 1.05-folds after 14 days [167]. In another study, *Sida cordifolia* Linn. (SDL) in the form of its hydrogel was evaluated against STZ induced male albino diabetic rats. SDL hydrogel was found to show 1.35-folds and 4.4-folds increase in wound closure and hydroxyproline content in

a period of 12 days when compared to the diabetic control group respectively [168]. In another study, a hydrogel of sodium carboxymethyl chitosan-recombinant human epidermal growth factor conjugate (NaCMCh-rhEGF) was found to accelerate the DW healing by a 1.58-folds increase in wound contraction and 1.05-folds increase in epithelization as compared to its placebo hydrogel counterpart [169]. Similarly, Thangavel et al., investigated the effect of L-Glutamic acid and chitosan loaded hydrogel (LG + CS hydrogel) against DW in STZ induced male Albino diabetic rat model where they showed 4 days and 6 days faster wound healing rate in comparison to CS hydrogel and diabetic control group. This effect was due to enhanced angiogenesis and collagen deposition at the site of injury [164]. In a similar study, hydrogel of *Moringa oleifera* (MO) leaf extract was tested for its healing

properties against DW in STZ induced male Wistar diabetic rat model. Topical application of MO leaf extract loaded hydrogel showed 1.96-folds and 1.22- folds faster DW healing as compared to diabetic control and standard treated groups respectively [170]. In another study, DW healing effect of deferoxamine (DFO) hydrogel was evaluated on db/db diabetic mice where they exhibited 1.13-folds and 1.3-folds increase in epidermal length and 4.25-folds and 2-folds increase in antibacterial activity against *S. aureus* as compared to diabetic control and placebo hydrogel treated groups respectively indicating wound healing potential of the formulation [171]. In another study, Patil et al., investigated the effect of fluorinated methacrylamide chitosan (MACF) hydrogel dressings against DW in male adult db/db diabetic mice model where they observed significant increase in wound closure (1.75-folds

Table 4Preclinical studies of nanohydrogels loaded with various drugs for treating diabetic wounds.

Sr. No.	Drug	Method of preparation	Animal model	Research highlights References	
1.	PVA	Ultraviolet light-initiated polymerization	Female db/db mice	• ↑ 2-folds scar thickness within 22 days ↑ angiogenesis and migration of endothelial	[177]
2.	Chitosan	Ultraviolet light-initiated polymerization	db/db mice	cells • ↑ 5-folds wound contraction rate within 16 days	[178]
3.	НА	Michael-type addition	Female db/db mice	 † collagen deposition 2.2-folds decrease in wound area within 14 days 	[179]
4.	IL-8 and MIP-3 α loaded gelatin hydrogel	Cross-linking	Male ICR mice	angiogenesis and granulation tissues formation 5 days faster DW healing as compared to other groups	[180]
5	Glutamine-histidine-arginine-glutamic acid- aspartic acid-glycine-serine	Cross-linking	Male db/db mice	 ↑ neovascularization and collagen deposition Complete DW healing within 21 days ↑ re-epithelialization and angiogenesis 	[181]
6.	Neomycin sulfate	Freezing-thawing	Male SD rats	 ↑ 1.07-folds wound contraction within 6 days ↑ angiogenesis and collagen deposition 	[182]
7.	AC	Crosslinking	BKS db mice	 † 1.06-folds wound contraction within 20 days 	[183]
8.	Insulin/L929	Cross-linking	SD rats	• † 1.05-folds wound contraction within 18 days	[184]
9.	Sacran	Solvent evaporation	Male albino mice	 ↑ collagen deposition ↑ 1.26-folds wound contraction within 12 days 	[185]
10.	HP-PEGs, HA-SH and ADSCs	Thiol-ene click reaction	Male SD rats	 ↑ angiogenesis and collagen deposition ↑ 1.87-folds wound closure within 21 days ↑ angiogenesis and re-epithelialization 	[186]
11.	DFO	Solution polymerization	SD rats	 ↑ 1.5-folds wound contraction within 14 days ↑ angiogenesis and antibacterial action 	[171]
12.	HEP and BEM	Solution polymerization	Female Wistar rats	† 1.02-folds wound contraction within 21 days † angiogenesis and neovascularization	[187]
13.	ILM and EGF	Solution polymerization	SD rats	• ↑ 1.11-folds wound contraction within 12 days	[188]
14.	aECM and SC	Solution polymerization	Male SD rats	 † collagen deposition 4 days faster DW healing as compared to other groups 	[189]
15.	Berberine	Solution polymerization	Female SD rats	 † extracellular matrix † 2-folds wound closure within 13 days † angiogenesis and anti-inflammatory action 	[190]
16.	PAP	Solution polymerization	Male Wistar rats	 ↑ 1.26-folds wound closure within 15 days ↑ angiogenesis and re-epithelialization, 	[175]
17.	PVA-CEC-AGA/Ag	Solution polymerization	Adult male SD rats	collagen deposition • ↑ 1.55-folds wound closure within 20 days	[191]
18.	Ceffe-γ-PGA hydrogel	Polymerization	Male BKS-Leprem2Cd479/	 ↑ angiogenesis and collagen deposition • ↑ 1.19-folds wound closure within 17 days 	[192]
19.	Human Umbical derived mesenchymal stem cells exosomes	Solution polymerization	Nju mice SD rats	↑ capillary density and cell proliferation Complete DW healing in 18 days ↑ formation of epithelium, collagen bundle	[193]
20. FA	Α	EDC/NHS cross-linking method	Male C57BL/6 mice	fibers and hair follicles ↑ 1.22-folds wound closure within 14 days ↑ angiogenesis and collagen deposition	[194]

Abbreviations AC; Atellocollagen, aECM; Acellular extracellular matrix, ADSCs; Adipose-derived stem cells, BEM; Bemiparin, DFO; Desferrioxamine, EGF; Epidermal growth factor, EDC; Ethylenediamine carbamide, FA; Fayalite, HA; Hyaluronic Acid, HEP; Chitosan functionalized with heparin, HP-PEGs; Hyperbranched multi-acrylated poly (ethylene glycol) macromers, HA-SH; thiolated hyaluronic acid ILM; Insulin-loaded micelles, NHS; N hydroxy succinimide, PVA-CEC-AGA/Ag; poly-vinyl alcohol-N-carboxyethyl chitosan agarose/Ag, PAP; Periplanar Americana herbal residue, SC; Sacchachitin.

and 2-folds) and angiogenesis (1.71-folds and 2.11-folds) as compared to Aquaderm (marketed formulation) and negative control groups indicating wound healing potential of the developed formulation [172]. Similarly, activity of a hydrogel prepared from edaravone nanoparticles (EDA-NP) was checked against DW in STZ induced male diabetic C57BL/6J mice. The EDA-NP hydrogel (L) was found to exhibit about 1.92-folds, 1.35-folds and 1.01-folds increase in wound healing rate in 15 days as compared to that in diabetic control, EDA-NP (H) and normal control-treated groups respectively. The enhanced activity of EDA-NP hydrogel was attributed to its increased antioxidant and anti-inflammatory effect at the site of injury [173]. In another study, in a 60 days clinical study on 56 patients, Gallelli et al., investigated the effect of quercetin (Qu) and oleic acid (OA) embedded nano hydrogel against DFU in diabetic patients having lower limb skin wound. Only 16 patients (57.1%) treated with 2% w/w hydrogel of Hyaluronic acid showed complete re-epithelization while 26 patients (92.8%) treated with 0.2% hydrogel containing Qu and OA showed complete re-epithelization at the site of injury. Moreover, topical application of HA resulted in some side effects while nano-hydrogel embedded with Qu and OA was found to be devoid of any side effect and thus was considered to be safe to use [174]. In one of the study, Wang et al., compared the wound closure effect of Periplaneta americana (PA) hydrogel against DW in diabetic rats with that by Kangfuxin Liquid (KFX), carbomer (CBM)/carboxymethyl cellulose (CMC) gel where they observed 1.03-folds, 1.36-folds and 1.53-folds increase in wound closure as compared to those by KFX liquid, CBM/CBC gel and diabetic control groups respectively indicating wound healing potential of the formulation [175]. In a study on the effect of alginate (Alg)-polydeoxyribonucleotide (PDRN) hydrogel against DW in C57BLKS/J-db/db male diabetic mice, it was found to exhibit 2.05-folds 1.9-folds and 2.18-folds increase and in collagen density and 2 folds, 2.1-folds and 2.8-folds decrease in wound area as compared to Alg, PDRN injection and saline-treated groups respectively [176].

The various nanohydrogels explored so far to treat DW are given in Table 4.

4.4. Solid lipid nanocarriers (SLNs)

SLNs are submicron colloidal carriers with size ranging from 50 to 1000 nm. These are generally composed of physiological lipids dispersed in water or aqueous surfactant solution [195]. The examples of lipids used in SLNs are epic cerana, stearic acid, hexadecyl hexadecanoate, caprylic/capric triglyceride, glyceryl monostearate, compritol, docosanoic acid and solid paraffin. The surfactants used in SLNs include egg and soya lecithin, polyoxyethylene, poloxamine, Tween 80, and poloxamer [196]. The advantages of SLNs are their large surface area, small size, and high drug loading capacity, all of which help in improving the therapeutic effects of the drugs contained therein [197, 198]. The SLNs explored so far for the treatment of DW are discussed in the subsequent sections.

Recombinant human epidermal growth factor (rhEGF) loaded lipid nanoparticles (SLNs) at a dose of 10 µg were used against chronic wounds in male db/db mice model where they exhibited 1.09-folds, 1.2folds, 1.3-folds, 1.29-folds and 1.8-folds decrease in inflammation as compared to the treatments in the given doses of rhEGF-SLNs, MSrhEGF, free rhEGF, empty SLNs and untreated control groups, respectively [199]. In another study, Rajesh et al., have investigated the antibacterial effect of the methanolic extract of Abutilon indicum (MEAI) SLNs against microorganisms that causes DFU. The antibacterial effects evaluated in terms of zone of inhibition (mm) against E. coli, Bacillus subtilis and P. aeruginosa. The results revealed that MEAI SLNs exhibited maximum zone of inhibition for *E. coli* (22 ± 0.57 mm for MEAI vs 24.33 \pm 0.33 mm for MEAI SLNs), *B. subtilis* (15.33 \pm 0.88 mm for MEAI vs 18.66 ± 0.66 mm for MEAI SLNs) and P. aeruginosa (20.66 ± 0.88 mm for MEAI vs 23.66 \pm 0.33 mm for MEAI SLNs). [200]. Similarly, wound healing effect of SLNs based rutoside ointment (NRO) in STZ induced

male Albino Wistar diabetic rat model. Application of NRO accelerated DW healing by promoting hydroxyproline content and antioxidant action at the wound site. Percentage wound size on the 16th day of treatment was found to be 11.35%, 69.5% and 99.9% for NRO, rutoside ointment and ointment base treated groups respectively. In one of the study, application of silence tumour necrosis factor α loaded lipid nanoparticles (siTNFα-loaded LNPs) against DW in STZ induced C57BL/6 diabetic mice model was found to accelerate DW healing by promoting anti-inflammatory action and reducing tumour necrotic factor-alpha (TNF- α) microsomal ribonucleic acid (mRNA) expression in the DW by 40-55%. Percentage of wound contraction was found to be 100% and 70% for siTNF α -loaded LNPs and control groups respectively on the 13th day of treatment [201]. Arantes et al., studied the effect of retinoic acid-SLNs (RA-SLNs) containing chitosan films against DW in STZ induced C57BL/6 male diabetic mice. On the 14th day of study, the percentage of wound healing was found to be 90% and 70% for RA-SLNs surrounded chitosan films, and placebo SLNs surrounded chitosan films respectively which was attributed to enhanced collagen deposition and decreased leukocyte infiltration rate as compared to that by placebo SLNs surrounded by chitosan films [202]. In a similar study on valsartan (Va)-SLNs gel against DFU in STZ induced male SD diabetic rats, the topical application of Va-SLNs gel was found to accelerate DW healing by promoting collagen deposition, angiogenesis and re-epithelization at the site of injury as compared to placebo SLNs gel.

4.5. Nanostructured lipid carriers (NLCs)

NLCs are the combinations of solid lipids, liquid lipids and emulsifiers. The ratio of these components is varied to obtain different forms of NLCs such as imperfect type, amorphous type and multiple types [203]. NLCs are reported to minimize the limitations of SLNs such as limited drug loading, entrapment efficiency and drug release during storage [204,205]. Reports on NLCs of drugs to be used for DWs are relatively recent. In 2017, Simvastatin (SV)-stem cells-based NLCs scaffolds were explored for treatment of DW in db/db mice model wherein they exhibited 1.19-folds, 2.56-folds, 1.02-folds and 1.97-folds decrease in wound area as compared to SV, placebo NLCs scaffolds, placebo NLCs \pm stem cell scaffolds and SV NLCs respectively [206]. In a subsequent clinical trial on 27 patients with DFU phenytoin-NLCs (PHT-NLCs) hydrogel was investigated. The formulation was found to promote collagen deposition, angiogenesis and fibroblast proliferation at the ulcer site. In the eight day study, the percentage of wound healing was found to be 95.82 \pm 2.22%, 47.10 \pm 4.23% and 34.91 \pm 28.33% for PHT-NLC-hydrogel, PHT and blank-hydrogel respectively [207]. In another study, Natrajan et al., studied the effect of pioglitazone NLCs loaded collagen/chitosan composite scaffolds (Pio-NLC-COL-CS scaffolds) against DW in STZ induced male Albino Wistar diabetic rat model where they observed 1.45-folds and 1.20-folds increase in wound contraction as compared to control and blank NLC-COL-CS scaffolds treated groups respectively indicating the wound healing potential of the formulation [208]. In another study, taking a cue from the use of aromatic oils in the treatment of DWs, Ghodrati et al., investigated the effect of peppermint essential oil-NLCs (PEO-NLCs) against the infective wound in the BALB/c mice model. On 14th day, PEO-NLCs showed maximum vascularization (2.66 \pm 0.40 for PEO-NLCs vs 2.16 \pm 0.40 for PEO), fibroblast infiltration (2.83 \pm 0.40 for PEO NLCs vs 2.33 \pm 0.50 for PEO) and epithelization (2.83 \pm 0.20 for PEO NLCs vs 2.60 \pm 0.30) as compared to PEO treated groups [209]. Similarly, Khezri et al., have studied the effect of rosemary essential oil NLCs (REO-NLCs) against infected BALB/c mice wound model. On the 12th day, REO-NLCs and REO treated groups showed excellent wound healing while 5 mm² and 8 mm² wound area were still left in mupirocin and control groups respectively. The results of antibacterial activity indicated that REO-NLCs showed 1-folds, 3-folds and 4-folds decrease in total bacterial count as compared to REO' mupirocin and control groups respectively [210]. In a study on the effect of poly (lactic-co-glycolic acid)

(PLGA)-Aloe vera (AV) based NLCs against DW in male db/db mice model, it was found to exhibit 1.03-folds and 1.53-folds increase in wound contraction as compared to PLGA-AV and control group (untreated group) respectively. The re-epithelization study revealed that PLGA-AV NLCs exhibited 2.2-folds increase in re-epithelization rate as compared to untreated control group [211]. In one of the study, wound healing effect of epidermal growth factor (EGF)-curcumin (Cur) based NLCs against DW in STZ induced male SD diabetic rats where they showed increase in wound contraction by 1.13-folds, 1.26-folds and 1.30-folds in comparison to EGF, Cur and blank NLCs treated groups respectively [212]. In another study, silicon elastomer-20 (S)-protopanaxadiol-loaded NLCs (PPD-NS) based gel were evaluated against DFU in db/db diabetic mice model where it was observed that PPD-NS based gel exhibited 1.13-folds, 1.39-folds and 1.04-folds decrease in wound closure as compared to control, PPD-S and PPD-N treated groups respectively. The angiogenesis study revealed that PPD-NS based gel exhibited 1.06-folds, 1.37-folds and 2-folds increase in blood vessels as compared to PPD-N, PPD-S and PBS treated groups respectively [213]. In one of the study, wound healing effect of mentha pulegium essential oil (MPO) based NLCs were evaluated against DW in BALB/c mice model. The results revealed that the topical application of MPO based NLCs accelerated DW healing within 15 days by showing 1.6-fold decrease in wound area in comparison to diabetic control group

4.6. Nanoparticles

NPs are colloidal solid particles having the size in the nanometer range [215]. Generally, NPs can be zero-dimensional (0-D), one-dimensional (1-D), two-dimensional (2-D) and three-dimensional (3-D) in nature [216]. The basic composition of NPs includes three layers viz. surface layer (polymers, surfactants, small molecules, and metal ions), shell layer, and core [217]. The various NPs such as gold NPs, silver NPs, copper NPs, polymeric NPs, cerium oxide NPs, zinc oxide NPs used to treat DW are discussed in the subsequent sections.

4.6.1. Gold NPs (AuNPs)

AuNPs are extensively used in tissue engineering due to their biocompatibility. Moreover, unlike silver, AuNPs does not impart antimicrobial activity. Thus, AuNPs always used along with other biomolecules for their utility in biomedical applications including DW. For instance, crosslinking of AuNPs with collagen, polysaccharides and peptides are used in integrating GFs without altering the structural integrity of attached biomolecules. These modification in AuNPs results in better biocompatibility and biodegradability [218]. Moreover, in one of the studies, it has been found that when AuNPs were applied to the wound site they exhibit increase in re-epithelization, collagen deposition, granulation tissue formation and ECM deposition. Looking at these benefits various AuNPs explored so far to treat DW are discussed in the subsequent sections [219].

Chen et al., studied the effect of coadministration of AuNPs, epigallocatechin gallate (EGCG) and α -lipoic acid (ALA) (AuEA) against DW in STZ induced male BALB/c diabetic mice. On the 7th day, AuEA exhibited about 2-folds decrease in wound area as compared to vehicletreated groups indicated wound healing potential of the formulation [220]. Similarly, Jing et al., compared the wound healing effect of nerolidol functionalized AuNPs (N-AuNPs) against DW in male Albino diabetic rats with that by antimicrobial peptide (LL37) AuNPs and diabetic control groups. Application of N-AuNPs accelerated DW contraction by 1.2-folds and 1.03-folds as compared to control and LL37 AuNPs treated groups [221]. In another study, the wound healing effect of LL37/pDNAs AuNPs were evaluated against DW in STZ induced male C57BL/6 diabetic mice. The results revealed that LL37/pDNAs AuNPs treated groups exhibited 2.25-folds, 2.57-folds and 1.8-folds increase in wound closure within 10 days as compared to control, pDNAs and LL37 treated groups respectively [222]. In another study, Ponnanikajamideen et al., compared the wound healing effect of Chamaecostus cuspidatus (CC) AuNPs against DW in STZ induced adult male Wistar diabetic mouse model with that by CC (plant extract) treated groups. The results revealed that CC AuNPs treated groups exhibited about 1.03-folds increase in wound contraction as compared to plant extract treated group [223]. In one of the study, effect of dual-functional AuNPs i.e., 11-mercaptoundecyl-N, N, N-trimethylammonium bromide (11-MTA)/-VEGF-A165 AuNPs were evaluated against DW in db/db diabetic mice. On the 12th day, dual-functional AuNPs exhibited 5-folds and 5.5-folds decrease in wound size as compared to 11-MTA and untreated groups respectively [224]. In another study, wound closure and re-epithelization potential of keratinocytes growth factor (KGF) AuNPs against DW in STZ induced female SD diabetic rats. The results were compared with control, KGF and AuNPs alone. The results revealed that KGF AuNPs treated groups exhibited about 1.58-folds, 1.26-folds and 1.46-folds increase in percentage wound healing and about 3.75-folds, 1.5-folds and 3.4-folds increase in re-epithelization as compared to control, KGF and AuNPs alone treated groups respectively [225]. Similarly, wound closure activity of calreticulin conjugated AuNPs were evaluated against DW in STZ induced male BALB/C diabetic mice, it was found to exhibit 1.39-folds increase in percentage wound closure within 10 days as compared to Au NPs alone treated groups [226]. In another study, effect of DsiRNA-loaded AuNPs based thermoresponsive gel were evaluated against DW in In vitro cultured human dermal fibroblasts cells. The results revealed that DsiRNA-loaded AuNPs exhibited 98% of cell viability and promoted an increase in cell migration and proliferation rate that helped in wound healing [227].

4.6.2. Silver NPs (AgNPs)

Silver is a bactericidal agent and is frequently used to treat wounds, burns and various ulcers. For example, silver in the form of nitrate is used to treat non-healing chronic wounds. Furthermore, silver is also coated on dressings to distribute drug efficiently, which plays a key role in healing wounds. Moreover, the application of AgNPs based dressings on the wound site is devoid of toxicity which makes them more advantageous in terms of dressings. In addition, the co-administration of AgNPs with collagen enhances antibacterial activity and make them as a suitable biomaterial for wound dressings. In some of the studies it has been reported that AgNPs reduces oxidative stress and inflammatory cytokines as well, which aids in wound healing. Various uses of AgNPs in treating DW is highlighted below.

Tehrani et al., have studied the antibacterial effect of AgNPs coated leather and leather without AgNPs coating on different strains of grampositive and gram-negative bacteriae isolated from foot ulcers of 95 diabetic patients. The results revealed a significant decrease in the number of counts of bacteriae treated with AgNPs coated leather as compared to the non AgNPs coated leather indicating the antibacterial potential of AgNPs [228]. In one of the study, wound healing effect of Azadirachta indica (AI) AgNPs were evaluated against DW in STZ induced male Swiss Albino diabetic mice. On the 21st day AI AgNPs exhibited increase in wound contraction by 1.13-folds and 1.78-folds in comparison to SSD and control groups respectively [229].

Hernandez1 et al., conducted a clinical trial in which the effect of AgNPs was investigated against DFU. In this study, wound healing rate (days) of AgNPs were evaluated in two patients with Wagner third grade foot ulcer (W3GFU) and Wagner second grade foot ulcer (W2GFU) respectively. To treat these patients, AgNPs were applied topically at the site of the injury. The results revealed that topical application of AgNPs showed complete wound healing in W3GFU patient within 21 days by promoting re-epithelization and anti-inflammatory action. While in the case of a patient having W2GFU AgNPs took 26 days to completely heal the wound. The outcomes of the study inferred that AgNPs exhibited about 1.23-folds faster wound healing rate in the patient having W3GFU as compared to a patient having W2GFU [230]. In one of the study, wound healing activity of bamboo cellulose nanocrystals (BCNCs) co-administered AgNPs were evaluated against DW in STZ induced

Swiss albino diabetic mice. Application of BCNCs co-administered AgNPs at the wound site exhibited increase in percentage wound closure by 1.1-folds and 1.06-folds as compared to AgNPs and BCNCs treated groups respectively [231]. In another study, Shi et al., compared the wound healing effect of AgNPs composed of thiolated chitosan (C), N-acetyl-L-cysteine (N) and maleic acid-grafted dextran (DM) (AgNPs-CNDM) against DW in STZ induced male SD diabetic rats where they observed on 10th day, AgNPs-CNDM exhibited about 5-folds, 2-folds, 4.8-folds decrease in wound area as compared to control, AgNPs and CNDM treated groups [232]. In another study, activity of AgNPs impregnated chitosan (CS)- poly ethylene glycol (PEG) hydrogel was evaluated against DW in alloxan monohydrate induced diabetic rabbits. The results showed that AgNPs impregnated CS-PEG hydrogel exhibited about 1.58-folds and 1.18-folds increase in percentage wound contraction as compared to AgNPs alone and CS treated groups respectively [233]. Similarly, effect of co-administration of hypochlorous acid (HClO), chitin-nanofiber sheet (CNFS) and AgNPs were evaluated against DW in db/db diabetic mice. The results revealed that co-administration of HClO), chitin-nanofiber sheet (CNFS) and Ag NPs treated groups exhibited about 1.66-folds and 1.37-folds increase in angiogenesis and length of granulation tissue as compared to combination of HClO and CNFS treated groups respectively. This indicated the potential of AgNPs in the therapy [234]. In one of the study, Kumar et al., have investigated the wound healing effect of Aloe arborescens AgNPs against diabetic wounded fibroblast cells (WS1) using the scratch assay. The results revealed an accelerated wound healing within 48 h when Aloe arborescens AgNPs were used against WS1 human skin fibroblast [235]. In another study, antibacterial effect of Aerva lanata (AL) AgNPs were tested against microorganisms isolated from the foot of the DFU patients. The microorganisms isolated from DFU patient foot were E. coli, (PA), Bacillus Subtilis (BS) and S. aureus. These isolated microorganisms were tested against AL AgNPs. The results revealed that AL AgNPs showed antibacterial action against these DFU bacterial isolates and showed minimum bactericidal concentration (MBC) and minimum inhibitory concentration (MIC) of $10-20 \mu g/mL$ and 5-15μg/mL respectively. Besides these, AL AgNPs also showed an increase in accumulation of intracellular reactive oxygen species (ROS), augmented membrane leakage, modified membrane integrity, and drastically ruptured membrane which helps in wound healing [236]. Similarly, activity of co-administration of synthetic chitosan-based composite hydrogel (SNP_FCHG) and EGF AgNPs were evaluated against DW in STZ induced male SD diabetic rats, it was found to exhibit about 1.18-folds and 1.35-folds increase in percentage wound closure as compared to HeraDerm (commercial) and gauze treated groups. Additionally, SNP_ECHG co-administered EGF AgNPs exhibited about 1.24-folds and 1.55-folds increase in collagen content as compared to HeraDerm (commercial) and gauze treated groups [237].

4.6.3. Copper NPs (CuNPs)

Copper NPs are gaining tremendous attention in the treatment of chronic wounds. They are essential for modulation of various GFs and cytokines as well participate in all phases of wound healing [238]. Furthermore, they can promote angiogenesis by affecting the hypoxia inducible factor-1a (HIF-1a) and regulates VEGF secretion [239]. Moreover, CuNPs also exhibit antimicrobial effect which play key role in wound healing. In addition, several studies revealed that CuNPs also promote cell migration and cell proliferation which is helpful in accelerating wound healing process. Utilizing these remarkable potential various CuNPs are used to treat DW which are discussed in the subsequent sections [240].

In another study, the effect of CuNPs was examined on a 58-year-old patient with T2DM, micro and macrovascular complications, critical limb ischemia, diabetic retinopathy, peripheral artery disease, chronic renal disease of stage II and ulcer in the right heel as well as necrotic injuries in the 2nd, 3rd, and 4th toes. Due to these complications, the patient was programmed for a supracondylar amputation. To avoid

amputation, this patient was included in the clinical trial and given the treatment of CuNPs. When the CuNPs were applied at the site of injury, it was observed that within three months, the ulcer gets healed. The patient was saved from supracondylar amputation. This effect was due to antibacterial action, epithelization and stimulation of autolytic debridement at the site of injury [241]. Similarly, wound healing activity of folic acid-based CuNPs (F-HKUST-1) were tested against DW in STZ induced diabetic mice with that by HKUST-1, PBS and folic acid-treated groups. Topical application of F-HKUST-1 at the wound site exhibited about 1.05-folds, 1.11-folds and 1.26-folds increase in wound contraction as compared to HKUST-1, PBS and folic acid respectively [242]. In a study on the effect of yeast extract (YE)-immobilized-copper (Cu) nanoparticle (NP)-dispersed carbon nanofibers (CNFs) was checked against DW in STZ induced female Wistar diabetic rats. On the 14th day, YE-immobilized CuNPs dispersed CNFs demonstrated 1.09-folds and 1.28-folds increase in wound contraction as compared to CNFs and control groups respectively [243]. In one of the study, antibacterial effect of herbal extract-based CuNPs ointment were tested against microorganisms isolated from the foot of DFU patient. The microorganisms isolated from the foot of the DFU patient were E. coli and S. aureus. The results revealed that CuNPs ointment showed antibacterial action against S. aureus and E. coli with a zone of inhibition of 14 mm (S. aureus) and 15 mm (E. coli), respectively [244]. In another study, Zhang et al., compared the wound closure and angiogenesis activity of copper-based metal-organic framework (HKUST-1) against DW in C57BL/6J diabetic mice where they exhibited about 2-folds decrease in percentage wound area as compared to control groups. Additionally, HKUST-1 treated groups showed increase in length of blood vessels by 2.1-folds in comparison to control groups [245]. Similarly, Liu et al., investigated the effect of ultrasmall CuNPs (Cu5.4O USNPs) against DW in a male BALB/c diabetic mouse model. On the 15th day, Cu5.4O USNPs exhibited about 2.6-fold increase in wound contraction as compared to control (PBS) treated groups respectively [246].

4.6.4. Cerium oxide NPs (CONPs)

CONPs are gaining remarkable attention in the field of tissue engineering. Because they promote growth of vasoendothelial cells (VECs), keratinocytes and fibroblasts, which are essential for wound healing. Moreover, CONPs exhibit antioxidant effect by scavenging reactive oxygen species (ROS). In addition, CONPs also promote angiogenesis and anti-inflammatory action [269]. CONPs also helps in the modulation of intracellular oxygen environment [247]. Looking at these potentials various CONPs used to treat DW are discussed in the subsequent sections.

In a study on the effect of CONPs conjugated small non-coding RNA (MicroRNA-146a) against DW in STZ induced diabetic porcine model. Application of CONPs conjugated MicroRNA-146a accelerated DW healing within 14 days by 1.51-folds as compared to diabetic control groups [248]. Similarly, Augustine et al., compared the wound healing effect of CONPs loaded electrospun Polv hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) membranes against DW in male SD rats where they found that on the 30th day, CONPs loaded PHBV treated groups exhibited about 1.14-folds increase in percentage wound contraction as compared to PHBV [249]. In another study, effect of CONPs based gel were checked against neuropathic DFU. A 62-year-old patient having a history of T2DM was selected for the study. The patient was suffering from T2DM for the last 20 years and had multifactorial diabetic complications such as retinopathy, neuropathy and microangiopathy of the lower limb. The patient also had hypertension and dyslipidemia. To treat diabetic complications, the patient had received insulin therapy and for hypertension perindopril. Atorvastatin was administered to manage dyslipidemia. Despite these ongoing treatment strategies, the foot ulcer got developed in the plantar region of the foot. For the treatment of foot ulcer, Trikacide 1500 mg orally per day was given along with gauze with the hypertonic solution (0.45% NaCl), and bandages were applied at the ulcer area for two months.

However, these treatments were failed to heal the wound. Finally, CONPs based gel was applied topically at the site of the foot ulcer. The treatment with CONPs based gel accelerated wound healing within 2 months by showing anti-inflammatory, antioxidant, bacteriostatic, and tissue regenerative activity at the site of injury [250]. In one of the study, activity of zwitterionic cryogel miRNA146a-conjugated CONPs were examined against DW in female db/db diabetic mice. The results were compared with the control zwitterionic cryogel. Topical application of zwitterionic cryogel miRNA146a-conjugated CONPs accelerated DW within 14 days by 1.43- as compared to control zwitterionic cryogel treated groups [251]. Similarly, in a study on the effect of nano silk (NS) co-administered CONPs conjugated miRNA146a against DW in female diabetic mice. The results revealed that NS co-administered CONPs conjugated miRNA146a treated groups exhibited 2.3-folds faster-wound healing as compared to diabetic control groups [252].

4.6.5. Zinc oxide NPs (ZnO NPs)

Zinc is an important micronutrient which plays key role in human physiology. It acts as a cofactor for various metalloenzymes which are necessary for immunomodulation, cell growth, proliferation and cell membrane repair. The deficiency of Zn is responsible for skin lesions, decrease in immunity, growth retardation and impede wound healing process. Moreover, Zn helps in the regulation of ECM and also exhibit antioxidant effect [253]. The various ZnO NPs explored so far to treat DW are discussed in the subsequent sections.

Antibacterial effect of Aristolochia indica (A. indica) loaded ZnO NPs were tested against micro-organisms isolated from the foot of the DFU patients. The microorganism isolated from the foot of the patients were coagulase-negative staphylococci (CNS) (0.44%), E. coli (45%), E. faecalis (1.16%), Acinetobacter baumannii (A. baumannii) (10%), S. aureus (15%) and Klebsiella pneumoniae (K. pneumonia) (8.4%). The results revealed a significant decrease in the number of counts of bacteriae i.e., 99.9% killing rate within 6 h when treated with A. indica loaded ZnO NPs as compared to vancomycin indicating the antibacterial potential of ZnO NPs [254]. In another study, wound healing effect of mucin-ZnO NPs based hydrogel were checked against DW in alloxan monohydrate induced male Wistar diabetic rats where they exhibited 1.21-folds, 1.17-folds, 1.08-folds and 1.03-folds increase in wound contraction in comparison to blank hydrogel, gentamicin, mucin and ZnO NPs treated groups respectively [255]. In one of the study, antimicrobial effect of radish root extract-based ZnO NPs against DFU. In this study, radish root extract-based ZnO NPs were tested against microorganisms isolated from the foot of the DFU patients where they exhibited 2.5-folds, 1.5-folds, 2.1-folds and 2-folds increase in zone of inhibition against E. coli, E. faecalis, P. aeruginosa and MDR-A. baumannii as compared to radish root extract alone respectively [256].

4.6.6. Poly (lactic-co-glycolic acid) NPs (PLGA-NPs)

PLGA is a copolymer and is extensively used in wound healing due to their biocompatibility and biodegradability. Moreover, PLGA helps in angiogenesis by releasing exogeneous lactate and accelerates wound healing process [257]. Several studies also revealed that when PLGA NPs is used along with other biomolecules it allows cell proliferation, migration as well as exhibit anti-inflammatory and antioxidant effect. In addition, promotes neovascularization, keratinocytes and fibroblasts migration, re-epithelization and collagen deposition at the site of injury. Looking at these benefits various PLGA NPs explored so far to treat DW are discussed below.

Activity of scaffolds/growth factor (GF) loaded PLGA NPs were evaluated against DW in male db/db diabetic mice. The results were compared with the control group and scaffolds/unloaded NPs. On the 15th day, scaffolds/GF-loaded PLGA NPs exhibited 1.41-folds and 4.25-folds increase in wound closure as compared to scaffolds/unloaded NPs and control scaffolds respectively [258]. In another study, effect of PLGA combined VEGF NPs were evaluated against DW healing in leptin receptor-deficient diabetic db/db mice. The results revealed that PLGA

combined VEGF NPs exhibited 1.05-folds, 1.53-folds and 1.48-folds increase in wound contraction as compared to PLGA-NPs, control and VEGF alone treated groups respectively indicating wound healing potential of the formulation [259]. Similarly, wound heling effect of ferulic acid PLGA-NPs (FA PLGA-NPs) were evaluated against DW in male Wistar diabetic rats. Topical application of FA PLGA-NPs accelerated DW healing within 14 days by 3.1-folds, 1.6-folds, 3.1-folds and 2.4-folds as compared to the control group, oral ferulic acid nanosuspension, mupirocin ointment and oral metformin-treated groups respectively [260]. In a study on the effect of insulin loaded PLGA NPs based hydrogel for DW healing in male SD diabetic rats. The results revealed that the percentage of wound injury index got reduced by 20-fold with insulin loaded PLGA NPs as compared to diabetic control groups. This wound healing effect was attributed to angiogenesis, collagen deposition and anti-inflammatory action at the site of injury [261]. Similarly, Hasan et al., compared the wound healing effect of polyethylenimine/diazeniumdiolate (PEI/NONOate)-doped nanoparticles (PLGA-PEI/NO NPs) against methicillin-resistant S. aureus (MRSA) biofilm-infected wounds in male BALB/c diabetic mice. On the 12th day, the percentage of wound closure of mice treated with PLGA-PEI/NO NPs was found to be 95.5%. However, control and PLGA-PEI/NO NPs treated groups showed negative effect [262]. In another study, sesamol-PLGA (SM-PLGA) nanosuspension were evaluated against DW in STZ induced male Wistar diabetic rats, it was found to exhibit about 1.33-folds, 1.25-folds and 1.11-folds increase in percentage wound closure as compared to DFU control, SM-50 and SM-100 treated groups respectively. Additionally, SM-PLGA nanosuspension exhibited about 1.3-folds and 1.5-folds decrease in TNF- α levels as compared to SM-100 and SM-50 treated groups respectively [263]. In one of the study, rate of granulation and angiogenesis activity of Archidendron pauciflorum (AP) PLGA NPs ointment were evaluated against DW in STZ induced male Swiss-Webster diabetic mice. The results revealed that AP PLGA NPs ointment exhibited about 1.25-folds and 1.05-folds increase in the average area of granulation tissues and blood vessels in comparison to AP alone treated group [264]. Similarly, activity of PLGA/GE electrospun nanofibrous mats loaded liraglutide were evaluated against DW in STZ induced male SD diabetic rats. The results showed that PLGA/GE electrospun nanofibrous mats loaded liraglutide showed 1.33-folds, 2-folds and 1.42-folds increase in percentage wound closure, blood vessels and cell migration rate as compared to control groups respectively [265].

4.7. Wafers

Wafers is a novel class of delivery system that have been applied directly on the suppurating wounds. These wafers adhere instantly at wound the surface and absorb exudate. Wafers are formed by using polymers such as sodium alginate, carrageenan, xanthan gum, methylcellulose and chitosan [266]. The Wafers used so far to treat DW have been discussed in the subsequent sections.

Antimicrobial effect of ciprofloxacin-loaded calcium alginate wafers was tested against microorganisms isolated from the foot of the DFU patients. The isolated microorganisms were found to be *P. aeruginosa*, *S. aureus* and *E. coli*. The results showed that ciprofloxacin-loaded calcium alginate wafers exhibited 45-folds, 1.09-folds and 45-folds increase in zone of inhibition against *E. coli*, *S. aureus* and *P. aeruginosa* as compared to commercial Algisite Ag® treated groups [267]. Similarly, Atia et al., compared the wound healing effect of diosmin nanocrystals loaded wafers against DW in STZ induced SD diabetic rats where they exhibited 1.25-folds, 1.15-folds and 1.50-folds decrease in ulcer area as compared to placebo wafers, diosmin powder gel and diabetic control groups respectively [268].

4.8. Dendrimers

Dendrimers are highly branched star-shaped macromolecules with

dimensions of nanometer-scale. These consist of three components a central core, an interior dendritic structure (the branches) and an exterior surface with functional groups [269]. They serve as a novel carrier due to their various advantages such as a capacity for improving solubility, absorption, bioavailability and target specificity [270]. Looking at these benefits various dendrimers explored so far to treat DW are discussed below.

In a study on the effect of polyamidoamine-starburst (PAM-RG4)/ minicircle (MC) VEGF165 *deoxyribose nucleic acid (DNA*) complex dendrimer (PAM-RG4/MC VEGF165 *DNA* complex dendrimer) against DW in STZ induced male C57BL/6J diabetic mice. The results revealed that PAM-RG4/MC VEGF165 *DNA* complex dendrimer treated groups accelerated DW healing within 12 days and exhibited about 1.1-fold decrease in wound area as compared to diabetic control-treated groups [271]. In another study, wound healing potential of peptide dendrimers (AK₂₁) were evaluated against DW in STZ induced male Kunming diabetic mice. Wound healing studies revealed that the AK₂₁ treated groups showed decrease in wound area by 10.8-folds than that of diabetic control groups due to increased re-epithelization at the site of injury [272].

4.9. Hydrogel nanotubes

The hydrogel nanotubes are formed by crosslinking of nanotubes with hydrogel in order to increase the three-and-four-dimensional printability, tunability, stiffness and electrical conductivity. Moreover, hydrogel nanotubes are gaining attention in the field of tissue engineering due to its biocompatibility and wound healing properties [273].

Activity of HNTs were evaluated against DW in male Wistar diabetic rats. Topical application of HNTs accelerated DW healing within 16 days by 1.33-folds as compared to diabetic control-treated groups. In addition, HNTs treated groups exhibited 1.3-folds, 1.8- folds and 1.6-folds decrease in IL-6 levels, TNF- α levels and IL-10 levels as compared to diabetic control groups respectively [274].

5. Biomaterials

5.1. Scaffolds

Scaffolds are three-dimensional polymeric structures consisting of biocreamics, hybrid materials and polymers which helps in tissue rejuvenation [275,276]. Based on tissue response, biocreamics are divided into three categories \emph{viz} . bioactive (bioactive glass), nearly inert (zirconia and alumina), and resorbable ceramics (β - and α tricalcium phosphate, TCP) [277]. Besides these, polymers used in the scaffolds are of two types, either synthetic or natural origin. The polymers that come under the synthetic category are polyphosphazene, poly (propylene) fumarate (PPF), polycaprolactone, polyanhydride, polyether ether ketone (PEEK), polyglycolic acid (PGA) and polylactic acid (PLA) [278]. The natural polymers used in scaffolds are chitosan, fibrin, collagen, and hyaluronic acid [279]. Some of the studies wherein, scaffolds have been used to treat DW are discussed below.

Effect of collagen-based VEGF scaffolds were evaluated against DW in STZ induced diabetic rats. Application of collagen-based VEGF scaffolds accelerated DW healing by 1.23-folds as compared to VEGF alone treated groups [280]. In another study, Lee et al., studied compared the wound healing effect of glucophage loaded collagen/PLGA scaffolds membranes against DW in STZ induced male SD diabetic rats with that by gauze sponge. The results showed that glucophage loaded PLGA scaffolds membranes exhibited about decreases in wound area by 3.5-folds within 14 days in comparison to gauze sponge treated groups [281]. In another study activity of prolyl hydroxylase domain protein 2 (PHD2) siRNA scaffolds were evaluated against DW in STZ induced SD diabetic rats. The results showed that PHD2 siRNA scaffolds exhibited about 2-folds increase in relative VEGF protein content as compared to SCR-siNPs [282]. Similarly, activity of perclan-VEGF chitosan scaffolds

were tested against DW in male Lewis diabetic rats, it was found to exhibit about 3.3-folds and 2.5-folds increase in reepithelization and angiogenesis as compared to the diabetic control group respectively [283]. In a study on the effect of co-administration of Eudragit RL, gentamicin and rhEGF scaffolds against DW in STZ induced female C57BL/6 diabetic mice where they exhibited decrease in open wound area by 1.63-folds within 12 days as compared to Eudragit RL alone treated groups [284]. Similarly, activity of laminin heparin peptide bonded GF scaffolds was tested against DW in STZ induced db/db diabetic mice. The results revealed that laminin heparin peptide bonded GF scaffolds exhibited about 2-folds and 1.8-folds increase in percentage wound closure and granulation tissue thickness within 10 days as compared to fibrin treated groups [285]. In another study, effect of co-administration of konjac glucomannan, keratin and Avena sativa extract (KGM/KER/OAT) hydrogel scaffolds were evaluated against DW in STZ induced healthy male Wistar diabetic rat model. The results revealed that KGM/KER/OAT hydrogel scaffolds showed complete wound healing within 16 days, whereas KGM/KER scaffolds and control group took 20 and 24 days for complete wound healing respectively. In addition, co-administration of KGM, KER and OAT hydrogel scaffolds showed 2.08-folds increase in antioxidant activity as compared to diabetic control group [286]. Similarly, activity of psyllium seed husk polysaccharide (PSH), KER and 1% morin (MOR) co-loaded scaffolds were evaluated against DW in STZ induced male Wistar diabetic rat model. Topical application of PSH, KER and 1% MOR co-loaded scaffolds accelerated DW healing within 16 days by showing 2.4-folds, 1.07-folds and 2.15-folds increase in wound closure as compared to diabetic control, PSH, KER and 0.5% MOR co-loaded scaffolds and combination of PSH and KER scaffolds treated groups respectively [287]. Similarly, wound healing effect of acellular dermal matrix PEGylated graphene oxide-mediated quercetin (ADM-GO-PEG/Que) modified collagen hybrid scaffolds were evaluated against DW in STZ induced ICR diabetic mice where they showed 1.38-folds, 1.22-folds and 1.03-folds increase in wound closure as compared to diabetic control, ADM, and GO-PEG/Que scaffolds treated groups respectively [288]. Similarly, activity of peritoin (PN) and connective tissue growth factor (CCN2) scaffolds were evaluated against DW in db/db diabetic mice. The results revealed that CCN2 scaffolds exhibited about 1.75-folds, 3.5-folds and 4.66-folds increase in vessel density as compared to PN, collagen and control-treated groups respectively [289]. In another study effect of gelatin-curcumin (GC)/lithospermi radix extract (L)/chitosan (C) nanofibrous scaffolds against DW in STZ induced male SD diabetic rat model, it was found to exhibit about 1.01-folds, 1.04-folds, 1.01-folds, 1.07-folds and 1.02-folds increase in wound recovery rate as compared to GC/L, GC, GC/C, Comfeel® and gauze (control) treated groups respectively [290]. In one of the study, Han et al., compared the wound healing effect of Asiatic acid (AA) scaffolds against DW in STZ induced male C57BL/6J diabetic mice with that by diabetic control groups. The results revealed that the AA scaffolds treated groups increase in the wound closure and collagen content by 2-folds and 1.58-folds respectively in comparison to diabetic control groups [291, 292]. Similarly, angiogenesis activity of platelet-derived growth factor-BB (PDGF-BB) co-loaded scaffolds were evaluated against DW in female C57BL/6JNju DIO type II diabetic mice. It was found to exhibit about 2-folds increase in angiogenesis rate as compared to placebo scaffolds [293]. In a study on the effect of rGO-ADM-mesenchymal stem cells (MSCs) scaffolds were tested against DW in STZ induced male ICR diabetic mice. Topical application of rGO-ADM-MSCs scaffolds accelerated DW healing within 14 days by showing 1.17-folds, 1.13-folds and 1.03-folds increase in wound closure as compared to control, ADM-MSCs scaffolds and ADM-GO scaffolds treated groups respectively [294]. Similarly, Jiang et al., compared the wound healing effect of silicon-doped amorphous calcium phosphate scaffolds against DW in STZ induced diabetic mice with that by diabetic control groups. It was found from the study that silicon-doped amorphous calcium phosphate scaffolds exhibited about 1.26-folds increase in wound closure within 15

days as compared to control groups [295]. In another study, effect of co-administration of the collagen-binding domain (CBD)/vasoendothelial growth factor (V)/stromal-derived growth factor-α (S) scaffolds against DW in STZ induced male SD diabetic rats. The DW healing studies revealed that CBD/V/S scaffolds showed 1.11-folds and 1.23-folds increase in wound healing as compared to control and placebo scaffolds treated groups respectively [296]. In one of the study, activity of polylactic acid (PLA)/chitosan (CS) nano scaffolds loaded cod liver oil (30%) were evaluated against DW in STZ induced diabetic rats. Topical application of PLA/CS nano scaffolds loaded cod liver oil (30%) exhibited increased in wound closure by 2.25-folds and 1.57 -folds in comparison to control and PLA/CS nano scaffolds alone treated groups respectively [297]. In one of the study, curcumin and zinc ion eutectic metal-organic frameworks-based scaffolds were evaluated against DW in STZ induced male C57 diabetic mice. The results revealed that curcumin and zinc ion eutectic metal-organic frameworks-based scaffolds exhibited about 3.18-folds and 2.28-folds increase in blood vessels and wound closure as compared to control groups respectively [298]. In another study, Yao et al., compared the wound healing potential of SDF-1 scaffolds against DW in STZ induced male SD diabetic rats with that by diabetic control and commercial wound dressing (Comfeel). It was found that SDF-1 scaffolds accelerated DW healing within 14 days by promoting 1.06-folds and 1.23-folds increase in wound closure as compared to diabetic control and Comfeel treated groups respectively [299]. In a study on the effect of dual growth factors (bFGF and VEGF) carrying NPs-based scaffolds against DW in STZ induced Swiss Albino diabetic mice. The results demonstrated that dual growth factors carrying NPs-based scaffolds exhibited about 1.06-folds and 1.22-folds increase in percentage wound closure as compared to bFGF and VEGF alone and diabetic control-treated groups respectively [300].

Various scaffolds used to treat DW are depicted in Table 5.

5.2. Nanofibers

Nanofibers are considered as the novel class of biomaterial that help in tissue rejuvenation and possess tunable features such as hydrophilic surface, suitable biocompatibility, controllable biodegradability and porosity [311,312]. They can be fabricated by using natural and synthetic polymers. The natural polymers include silk fibroin, cellulose, collagen, keratin, polysaccharides, gelatin, chitosan and alginate. The synthetic polymers include polyurethane (PU), poly (lactic acid) (PLA), polycaprolactone (PCL), poly (lactic-co-glycolic acid) (PLGA), poly (ethylene-co-vinyl acetate) (PEVA), poly (3-and hydroxybutyrate-co-3-hydroxy valerate) (PHBV) [312]. Some of the studies wherein nanofibers have been explored to treat DW are discussed in the subsequent sections.

Choi et al., investigated the effect of rhEGF-nanofibers at the dose of 10% w/w against DW in STZ induced female C57BL/6 mice. Topical application of rhEGF nanofibers accelerated DW healing within 14 days by 1.6- folds, 1.9-folds and 1.18-folds as compared to rhEGF/nanofibers (10% w/w), nanofibers alone and normal control groups respectively [313]. Similarly, wound healing potential of curcumin-PCL nanofibers were evaluated against DW in STZ induced male C57/B6 diabetic mice. Wound healing study revealed that the curcumin-PCL nanofibers treated group exhibited about 1.33-folds increase in wound closure as compared to PCL nanofibers alone treated groups [314]. In a study on the effect of Poly-N-Acetyl glucosamine (sNAG) membranes against DW in genetically modified diabetic Lep/r-db/db mice, it was found to exhibit about 1.2-folds increase in wound contraction as compared to non-treated groups. The granulation study revealed that sNAG membranes treated groups exhibited about 3-folds and 3.75-folds increase in granulation tissue as compared to non-treated groups and cellulose treated groups respectively [315]. In one of the study, potential of multiple growth factors (bFGF and EGF) based nanofibers were tested against DW in C57BL/6 diabetic mice. The results demonstrated that multiple growth factors-based nanofibers exhibited about 1.2-folds, 1.3-folds and 1.8-folds increase in percentage wound contraction as compared to bFGF nanofibers, EGF nanofibers and blank nanofibers treated groups respectively [316]. In another study, activity of multiple growth factors (VEGF, PDGF, bFGF and EGF) composite nanofibers were evaluated against DW in STZ induced male SD diabetic rats, it was found to exhibit 1.3-folds and 1.13-folds increase in wound contraction and the number

Table 5List of scaffolds prepared till date to treat diabetic wounds.

Sr. No.	Drug	Method of preparation	Animal model	Research highlights	References
1.	Curcumin and Zinc ion	Electrospinning	Male C57 mice	† 3.18-folds wound healing rate in 15 days † collagen deposition, re-epithelization and angiogenesis	[301]
2.	Insulin and poly-D-1-lactide-glycolide	Coaxial electrospinning	SD rats	 3.16-folds decrease in wound area in 14 days † collagen deposition, epithelial cell proliferation and fibroblast migration 	[302]
3.	Polydopamine Biactive glass	Electrospinning	Diabetic mice	 1.35-folds decrease in wound area within 14 days † angiogenesis, re-epithelization and collagen deposition 	[303]
4.	Genipin and SDF-1	Freeze-drying	Male SD rats	1.06-folds increase in wound healing in 14 days † angiogenesis, collagen synthesis and neovascularization	[304]
5.	Bone marrow MSCs	Electrospinning	Male C57BL/6 mice	 2.5-folds increase in re-epithelization rate within 7 days † ECM deposition, angiogenesis, re-epithelization and antiinflammatory action 	[305]
6.	Collagen-based Bcl-2-modified adipose-derived stem cells	-	Female db/db mice	 1.92-folds increase in wound healing within 14 days † neovascularization, angiogenesis, cellularity and reepithelialization 	[306]
7.	Timolol	-	Db/db mice	1.59-folds increase in re-epithelization rate within 14 days ↓ MMP-9	[307]
8.	Bilayered fibrin/poly(ether) urethane scaffolds loaded with platelet lysate	Combined apparatus	Male db/db mice	 3.47-folds decrease in wound area with 14 days ↑ re-epithelization and collagen deposition 	[308]
9.	2,2,6,6-tetramethyl-1-piperidinyloxy)/ Sacchachitin	Mechanical disintegration	SD rats	 DW healing within 14 days † neovascularization, collagen deposition and fibroblast migration 	[309]
10.	Poly-3- hydroxybutyrate and gelatin	Electrospinning	Male Wistar rats	 7.5-folds decrease in wound area within 14 days ↑ collagen synthesis, cell attachment and proliferation 	[310]

of blood vessels at the site of injury within 6 weeks as compared to diabetic control groups [317]. Similarly, Antibacterial activity of silver-containing nanofibres were evaluated against DW in STZ induced male Swiss albino diabetic mice. The results of the study showed that silver-containing nanofibres demonstrated increase in percentage healing factor by 1.1-folds and 1.2-folds as compared to AgNPs and insulin-treated groups [318]. In one of the study, Mohammadi et al., compared the wound healing potential of curcumin-loaded poly(\varepsilon-caprolactone) (PCL)/gum tragacanth (GT) (PCL/GT/Cur) nanofibers against DW in STZ induced male SD diabetic rats where they observed 5-folds increase in wound closure as compared to diabetic control groups indicated wound healing potential of the formulation [319]. In another study, effect of heparin mimetic peptide (GAG-PA/KP) nanofibers were evaluated against DW in STZ induced male SD diabetic rats, it was found to exhibit about 1.33-folds increase in percentage wound closure as compared to control groups. Additionally, it was observed from the study that GAG-PA/KP treated groups showed about 1.28-folds increase in granulation tissue per unit area in comparison to diabetic control groups [320]. Similarly, wound healing potential of CuNPs-yeast-carbon nanofibers were tested against DW in female Wistar diabetic rats. The results revealed that CuNPs-yeast-carbon nanofibers exhibited about 1.01-folds, 1.09-folds and 1.28-folds increase in percentage wound contraction as compared to CuNPs-carbon nanofibers, carbon nanofibers and control groups respectively [243]. In another study, Grip et al., compared the wound healing potential of β-glucan-loaded nanofibers against DW in male diabetic db/db mice with that by fiber carrier. It was observed from the study that β -glucan-loaded nanofibers exhibited about 3.5-folds decrease in open wound area within 24 days in comparison to fiber carrier treated groups [321]. In a study the effect of neurotensin-loaded on lactide-polyglycolide/cellulose nanocrystals (PLGA/CNC) composite nanofiber membranes against DW in adult male Wistar rats. Topical application of neurotensin loaded PLGA/CNC composite nanofiber accelerated wound healing within 14 days by 5.5-folds in comparison to diabetic control groups [322]. In one of the study, activity of nanofiber-expanded human CD34⁺ cells were tested against DW in STZ induced NOD/SCID diabetic mice. The results revealed that nanofiber-expanded human CD34+ cells exhibited 1.3-folds, 2.1-folds and 2.8-folds increase in percentage wound contraction, vascular density and collagen density as compared to CD34⁺ cells therapy alone treated groups respectively [323]. In another study, effect of 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) co-loaded sacchachitin nanofibers (SCNF) were evaluated against DW in STZ induced SD diabetic rats, it was found to exhibit 5.5 days and 2.5 days faster-wound healing in comparison to gauze and hydrogel treated groups respectively [307]. Similarly, potential of gelatin (GEL)-bacterial cellulose (BC)-glibenclamide (Gb) nanofibers and GEL-BC-metformin (Met) nanofibers were evaluated against DW in male SD diabetic rats. The wound healing study revealed that GEL-BC-Gb nanofibers exhibited about 1-fold, 1.6-folds and 1.9-folds decrease in wound area as compared to GEL-BC-Met, and diabetic control respectively. In addition, anti-inflammatory study demonstrated that GEL-BC-Gb nanofibers treated groups showed about 2.1-folds, 1.6-folds and 1.1-folds decrease in TNF- α levels as compared to diabetic control, GEL-BC nanofibers and GEL-BC-Met nanofibers treated groups respectively [324]. In a study on the effect of sesamol-cellulose acetate (CA)/zein composite nanofibers (H) at a dose of 5% w/v against DW in STZ induced male C57BL/6J diabetic mice. It was observed from the study that H treated groups exhibited about 1.25-folds, 1.66-folds and 1.02-folds increase in percentage wound closure within 9 days as compared to placebo nanofibers (M), diabetic control groups (S) and sesamol-CA/zein nanofibers (2% w/v) (L) treated groups [325]. Similarly, Samadian et al., compared the collagen density of cellulose acetate/gelatin/berberine (CA/Gel/Beri) nanofibers against DW in STZ induced adult Wistar rat model with that by sterilized CA/Gel and negative control groups. The results revealed that CA/Gel/Beri nanofibers accelerated DW healing within 16 days by

showing 1.23-folds and 3.01-folds increase in collagen density as compared to sterilized CA/Gel and negative control groups respectively [326]. In a study on the effect of Malva sylvestris extract-based nanofibers against DW in STZ induced male Wistar diabetic rats. Topical application of Malva sylvestris extract-based nanofibers accelerated DW healing by promoting myofibroblasts contraction at the site of injury. Additionally, it was observed from the study that Malva sylvestris extract nanofibers exhibited about 2.9-folds increase in percentage wound closure as compared to gauze treated groups [327]. Activity of oxygen-generating polycaprolactone-sodium per carbonate (PCL-SPC) nanofibers were evaluated against DW in STZ induced male SD diabetic rats. On the 27th day, PCL-SPC nanofibers exhibited about 1.05-folds and 1.28-folds increase in wound closure as compared to control and PCL + control-treated groups respectively [328]. In another study, effect of desferrioxamine (DFO) nanofibers against DW in STZ induced diabetic rats, it was found to exhibited 1.4-folds increase in percentage wound closure as compared to diabetic control groups [329]. In another study, Razzaj et al., compared the wound healing potential of cephradine (Ceph)-loaded gelatin (GE)/polyvinyl alcohol (PVA) electrospun nanofibers against DW in male NcZ10 diabetic mice with that by Ceph treated groups. The results revealed that on the 11th day, Ceph-loaded GE/PVA) electrospun nanofibers exhibited about 1.87-folds decrease in percentage wound areas compared to Ceph treated groups [330].

5.3. Nanocomposites

Nanocomposite system is consisting of NPs encapsulated in a matrix such as a nanofiber, hydrogel, foam, nanocrystals and films. These are popularly used in wound dressings for enhanced wound healing. These nano-based dressings possess higher porosity (surface to volume ratio). In addition to this, their structure causes stimulation of the topographic appearance of ECM that allows attachment and spreading of fibroblast and keratinocytes. Thus, it facilitates collagen synthesis and reepithelization of the wounds [331,332]. The various nanocomposites used against DW are discussed in the subsequent sections.

Activity of viscose/silver/alginate/nicotinamide/calcium (Vis/ AgA167NPs/ALG/NIC/CaCl2) nanocomposites was evaluated against DW in STZ induced diabetic rabbits. It was observed from the study that Vis/Ag-167NPs/ALG/NIC/CaCl2 treated groups exhibited about 1.51folds and 1.29-folds increase in percentage wound closure as compared to Vis/Ag-NPs/ALG and Vis/Ag-NPs/ALG/NIC treated groups respectively [333]. In another study, effect of bioactive glass (BG)/patterned electrospun membrane (PEM) nanocomposite against DW in STZ induced female C57/BL6 diabetic mice. On the 13th day, BG/PEM nanocomposites exhibited about 1.40-folds and 1.42-folds increase in wound contraction as compared to PEM and control groups respectively [334]. In another study, wound healing activity of reduced graphene oxide (rGO)-isabgol (Isab) nanocomposites were evaluated against DW in STZ induced male Wistar rats, it was found to exhibit DW healing within 20 days by 1.49-folds as compared to diabetic control groups [335]. In a study on the effect of AuNPs-calreticulin nanocomposites against DW in STZ induced male BALB/C mice. Topical application of AuNPs-calreticulin nanocomposites accelerated DW healing within 20 days by promoting collagen deposition, re-epithelization, keratinocytes and fibroblasts migration at the site of injury. On the 20th day, AuNPs-calreticulin nanocomposites exhibited about 1.11-folds, 1.08-folds and 1.07-folds increase in wound closure as compared to PBS, insulin and AuNPs treated groups respectively [231]. Similarly, activity of edaravone loaded alginate-based nanocomposite hydrogel were evaluated against DW in STZ induced male C57BL/6J mice where they exhibited 1.58-folds increase in wound contraction as compared to diabetic control groups [173]. In another study, effect of iron (Fe)-copper (Cu) nanocomposite were evaluated against DW in STZ induced Wistar albino diabetic rats, it was found to exhibit about 1.1-folds and 2-folds decrease in wound area in comparison to antibiotics and iodine solution treated groups respectively. Additionally, it was observed from the study that Fe–Cu nanocomposites showed a 1.3-folds and 1.1-folds decrease in IL-1 β level as compared to iodine solution and antibiotic-treated groups respectively [336]. In one of the study, Li et al., compared the wound healing effect of polyethylene glycol diacrylate (P)/sodium alginate (A)/bioactive glass nanoparticles

containing copper (BC) scaffolds (PABC scaffolds) against DW healing in male ICR mice with that by PAB and PA treated groups. Topical application of PABC scaffolds accelerated DW healing within 21 days by promoting angiogenesis, collagen deposition and neovascularization at the wound site. On the 21st day, PABC scaffolds exhibited about

Table 6
Clinical trials carried out till date for various therapeutics used to treat diabetic wounds.

Sr. No.	Drug	Delivery system	No. of patients	Research outcome	References
1.	CT-102 activated platelet supernatant	-	13	1.28-folds decrease in ulcer area as compared to placebo-treated groups	[339]
2.	Repifermin	-	94	1.41-folds increase in percentage wound closure as compared to placebo-treated group	[340]
3.	Pomegranate	Extract	60	Promoted wound healing by showing antibacterial, antioxidant and neovascularization at the wound site	[341]
4.	Acellular biomaterial derived from pig small intestine submucosa (OASIS)	Matrix	73	1.49-folds increase in wound closure as compared to marketed formulation (Regranex gel).	
5.	Connexin43-based peptide	Hydrogel	92	1.26-folds increase in mean per cent wound closure within 12 weeks	[343]
6.	Autologous platelet-rich plasma	Gel	129	1.59-folds increase in wound closure as compared to control groups	
7.	Nitric oxide	Patch	90	-	[345]
3.	Protease-modulating matrix and autologous growth factors + collagen biomaterials	_	54	1.59-folds decrease in ulcer depth within 8 weeks as compared to collagen biomaterials	[346]
9.	Mucopolysaccharide + chitosan	Dressing	85	4.5-folds increase in wound closure as compared to gauze treated groups	[347]
10.	Bemiparin	-	70	1.54-folds increase in ulcer healing rate as compared to placebo- treated groups	[348]
11.	Microcyn Rx	Superoxidized solution	67	1.65-folds increase in wound closure as compared to oral levofloxacin $+$ saline-treated groups	[349]
12.	HYAFF	Scaffolds	180	1.16-folds increase in wound closure as compared to the control group	[350]
13.	Dehydrated human amniotic membrane allografts	-	25	11.5-folds increase in wound healing rate compared to SOC	[351]
14.	Granulocyte stimulating factor mobilized peripheral blood CD34 ⁺ cell	-	5	The maximum time taken by the patient to heal the wound was 26 weeks. All the patients were relieved from pain after the treatment.	[352]
15.	Platelet	Gel	117	1.26-folds increase in wound closure as compared to control group	[353]
6.	Clostridial collagenase	Ointment	55	Promoted wound healing by 1.11-folds as compared to placebo- treated groups.	
7.	Honey	Dressings	348	1.32-folds increase in DW healing within 18 days	[355]
8.	Human acellular dermal matrix	-	168	 1.16-folds increase in wound healing rate as compared to conventional care 	
9.	Flaxseed oil omega-3 fatty acids	-	60	1.8-folds decrease in ulcer width as compared to placebo-treated groups	
20.	Zinc supplementation	_	60	1.07-folds decrease in ulcer length within 12 weeks as compared to placebo-treated groups	
21.	Silver	Nanoparticles	2	Accelerated diabetic wound healing within 25 days due to re- epithelization and antimicrobial action	
22.	Copper	Nanoparticles	1	Healed DFU within 3 months by showing epithelization and antibacterial action	[241]
23.	Recombinant human epidermal growth factor	Spray	167	1.46-folds increase in healing rate as compared to placebo-treated groups	[359]
24.	Cryopreserved placental membrane	_	26	65.4% patients showed wound closure within 5 weeks	[360]
5.	Sodium Alginate	Hydrogel	7	In the 12th week patients showed a remarkable reduction in DFU	[361]
26.	Platelet-rich fibrinogen	3D printed scaffold	1	DW healing within 14 days due to re-epithelization and neogranulation	[362]
27.	Allogeneic Adipose-Derived Stem Cell	Hydrogel	59	Accelerated diabetic wound healing within 12 weeks and the formulation was biocompatible	[363]
28.	Phenytoin	Nanostructured lipid carriers	27	Promoted angiogenesis, neovascularization and collagen deposition	[207]
29.	Aloe vera/Plantago Major	Gel	40	Healed the wound within 4 weeks and gel showed no side effects	[364]
30.	Erythropoietin	Hydrogel	20	Promoted angiogenesis, re-epithelization, proliferation and differentiation of keratinocytes at the site of injury	[365]
31.	Cerium dioxide	Nanoparticles	1	Increased antimicrobial, anti-inflammatory and antioxidant effect	[250]
32.	Esmolol Hydrochloride	Gel	350	DW healed within a period of 12 weeks	[366]
3.	rhEGF	-	610	Healed DW within 12 weeks	[367]
54. -	Hypothermically stored amniotic membrane (HSAM)	Membrane	76	HSAM treated groups exhibited about 60% reduction in wound area as compared to standard of care (SOC)	[368]
35.	Quercus infectoria	Solution	56	1.20-folds increase in wound closure within 8 weeks as compared to the control group	
36.	Cu/TiO2–SiO2	Nanogel	1	DFU got healed completely within 10 months due to re- epithelization and anti-inflammatory action	
37.	Quercetin and oleic acid	Nanohydrogel	56	Increased wound contraction rate by promoting antioxidant and antimicrobial action	[174]
38.	Plerixafor	S.C injection	26	Only 38% of patients showed wound healing and the trial terminated	[371]
39.	Omega-3-rich fish skin grafts	-	49	2.09-folds increase in wound healing rate as compared to SOC	[372]

1.5-folds and 1.63-folds increase in wound closure as compared to PAB and PA treated groups respectively [337]. In a study on the effect of methacrylated kappa-carrageenan (KaMA)-polydopamine (PD) modified ZnO/L-glutamic acid nanocomposite spray dressings against DW in female Wistar diabetic rats. Wound healing study showed that KaMA-PD modified ZnO/L-glutamic acid nanocomposite spray dressings treated group showed increase in wound contraction by 1.25-folds in comparison to control groups [338].

Various clinical as well ongoing clinical trials that have conducted recently on DW are depicted in Table 6 and Table 7. A list of patents that have been implicated for DW healing are given in Table 8.

6. Conclusion

The treatment of DW is still a major challenge due to the inability of the current therapies to provide a favorable environment for tissue rejuvenation. However, advancement in recent years has provided new insights in the field of DW healing. Taking these things into consideration, the present manuscript emphasized the advancement in the conventional and novel therapeutic moieties using NDDS and nanomaterials. A series of NDDS that have been designed to treat DW

include liposomes, exosomes, niosomes, transfersomes, NE, nano hydrogel, SLNs, NLCs, Metallic NPs, Polymeric NPs, dendrimers, hydrogel nanotubes and wafers. These nanocarriers provide sustained release, protection of the drug from the enzymatic degradation, target specificity and enhancement in the solubility. Owing to these benefits, offered by NDDS, the therapeutic agents loaded in them provide enhanced neo-vascularization, re-epithelization, antibacterial activity, collagen deposition and angiogenesis. The study also entailed the role of nanomaterials alone and in combination with NDDS to effectively treat DW. The nanomaterials covered in this article include scaffolds, nanofibers and nanocomposites. The advantages of nanomaterials such as biocompatibility, biodegradability, renewability, retention property at the wound site and less toxicity make them unique materials to be used for DW healing. Despite these enormous potentials, these delivery systems are associated with some limitations such as tedious preparation procedure, lack of international standards and less availability of evaluation methods for toxicological studies. The major issue is related to achieving a good in vitro-in vivo correlation because the in vitro experiments are performed in a controlled environment. In contrast, there are many biological barriers that impediment the passage of formulations under in vivo condition. Further, the physicomechanical and

Table 7

Patents filed till date for various therapeutics used to treat diabetic wounds

Sr. No.	Therapeutic moiety	Delivery system	Patent no.	Key Claims/finding	Reference
1.	Platelet-rich plasma	Gel	ES2600793T3	Accelerated wound healing by promoting increase in granulation tissue number, collagen deposition and vascularization	[373]
2.	Ulmus macrocarpa	Extract	KR20080071385A	Accelerated wound healing by promoting immunomodulation, granulation tissue and anti-inflammatory action at the wound site	[374]
3.	Stem/progenitor cells	Scaffolds	CN101330935B	Complete healing of DW within 26 days	[375]
4.	EGF	Cream	US7517528B2	Exhibited wound healing within 3-weeks	[376]
5.	Adipsin	Solution	US7638484B2	Increased epidermal migration and epidermal closure	[377]
6.	Exogenous polypeptide (KGF-2)	Gel	US20180311414A1	Helped in wound healing by showing antibacterial action	[378]
7.	HIF-1 modulator	Gel	US10751304B2	Promoted wound healing by showing angiogenesis and neovascularization at the site of injury. Additionally, HIF-1 modulator treated groups showed 2 days faster wound healing as compared to control group	[379]
8.	Treprostinil	Solution	US8563614B2	In this study, 10 patients with peripheral arterial disease were considered. The results revealed that out of 10 patients, 5 patients showed no sign of pain, 2 patients cured fully and 2 patients partly cured.	
9.	Collagen	Biofabric	US7928280B2	Out of 45 patients, 15 patients showed complete wound healing	[381]
10.	Adipose derived stem cells	Injection	US8105580B2	Accelerated DW healing within 7 weeks and also promoted hair growth at the wound site	[382]
11.	Cellulose/nano-silver sodium alga acid composite membrane dressings	Composite	CN103041438B	Accelerated wound healing by showing antibacterial action and exudate absorbing property	[383]
12.	Ulcer recovering oil (Dragon's blood, bletilla striata, frankincense, myrrh and catechu)	Oil	CN103479883B	Promoted 75% of the wound healing within 14 days	[384]
13.	Elastomeric matrix element (A61F2/02)	Matrix	US9050176B2	Accelerated wound healing by promoting angiogenesis and collagen deposition at the wound site	[385]
14.	Poly-N-acetylglucosamine	Nanofibers	US10561677B2	Exhibited wound healing by showing anti-inflammatory and antibacterial action against S. aureus	[386]
15.	Alimentary Protein-Based Scaffolds	Scaffolds	US8790921B2	Accelerated wound healing by promoting angiogenesis, collagen deposition, hydration and antibacterial action at the wound site	[387]
16.	Dichlorine monoxide	Solution	US20170049813A1	Helped in wound healing by antibacterial action	[388]
17.	Borate glass-based particles	Fibers	US9486554B2	Helped in wound closure after 22 days by promoting angiogenesis and antibacterial action	[389]
18.	Silver	Hydrogel	US9327029B2	Accelerated wound healing by promoting hydration, antibacterial action and relieved pain	[390]
19.	Anti-connexin polynucleotide	Gel	US20170240905A1	Healed venous ulcer within one month by showing anti-inflammatory action	[391]
20.	Nanofibrillary polysaccharide	Hydrogel	DK2958599T3	Maintained hydration, epithelization, contraction and antibacterial action at the site of injury	[392]
21.	Vesicles comprising EGF	Liposomes	US20180015145A1	Promoted wound healing by showing epithelization	[393]
22.	CRM1 inhibitor	Hydrogel	EP2968278B1	Accelerated DW healing within 14 days	[394]
23.	miR-92 inhibitor	Gel	US10280422B2	Promoted wound healing by promoting angiogenesis, granulation and re-epithelization	[395]
24.	Carnosine	Injection	CN105189531B	Promoted DW healing by providing strength to ECM, potentiate collagen deposition and increase insulin like growth factor-1	[396]
25.	Bismuth-thiols	Nanosuspension	US10835510B2	Accelerated wound healing by promoting migration of keratinocytes and fibroblasts at the site of injury	[397]

Table 8
Ongoing clinical trials till date for various therapeutics used to treat diabetic wounds.

Phase	Intervention	Condition	No. of participants	Allocation	Study start date	NCT number	Reference
NA	Lipoaspirate Injection	Investigate the effect of Lipoaspirate injection in diabetic lower extremity wound	250	Randomized	February 2009	NCT00815217	[398]
Phase2	Isoniazide	Investigated the safety and efficacy of isoniazide on DFU	60	Randomized	July 2011	NCT01342497	[399]
Phase1	Esmolol hydrochloride gel	Investigated the efficacy of esmolol hydrochloride gel on DFU	50	Randomized	February 2014	NCT01113515	[400]
Phase4	DermaPure	Study the effect of DermaPure against DFU	50	Randomized	August 2014	NCT02081352	[401]
NA	Cold plasma therapy	Investigate the effect of cold plasma therapy against DFU	65	Randomized	August 17, 2016	NCT04205942	[402]
Phase1	Autologous ADSC stem cells in fibrin gel	Study safety and efficacy of autologous ADSC stem cells in fibrin gel against DFU	20	Non- Randomized	September 3, 2018	NCT03865394	[403]
-	miR-200b and miR-21	Study the effect of miR-200b and miR-21 in DW	124	No	August 12, 2019	NCT02581098	[404]
Phase3	Platelet-rich plasma-fibrin glue in combination with vitamin E and C	Reduce oxidative stress by co- administration of Platelet-rich plasma-fibrin glue with vitamin E and C	24	Randomized	August 28, 2019	NCT04315909	[405]
-	Platelets rich plasma	Investigated the efficacy of platelets rich plasma against DFU	60	-	July 30, 2019	NCT03890172	[406]
-	Negative pressure wound therapy	Investigate the effect of negative pressure wound therapy against DFU	40	-	November 2019	NCT04093635	[407]
NA	Omega 3 wound fish skin graft	To investigate the effect of omega3 wound fish skin graft against DFU	100	Randomized	July 31, 2019	NCT04133493	[408]
NA	Kerecis Omega3 wound dies	Investigated the efficacy of Kerecis Omega3 wound dies in DFU	229	Randomized	July 2, 2020	NCT04537520	[409]
-	Biofilm modified macrophage	Study the effect of biofilms on DW	28	Non-Probability Sample	February 1, 2020	NCT03271580	[410]
NA	Topical Oxygen Chamber	Investigate the effect of oxygen therapy on DW	40	Randomized	February 1, 2020	NCT02313428	[411]
Phase2	Donepezil tablets	Investigate the effect of Donepezil tablets on DW	20	Randomized	August 30, 2020	NCT04505670	[412]
Phase1	Treprostinil iontophoresis	Investigate the effect of treprostinil iontophoresis against DFU	60	Randomized	January 28, 2020	NCT03654989	[413]
NA	Kelulut honey	Investigate the effect of Kelulut honey vs conventional dressings on DW	76	Randomized	June 2021	NCT04849143	[414]
Phase2	DFO gel	Investigate the effect of DFO gel on DFU	174	Randomized	August 30, 2021	NCT03137966	[415]
Phase1	Nitric oxide foot bath	Investigated the safety and efficacy of nitric oxide solution delivered as a foot bath to treat DFU	40	Randomized	February 23, 2021	NCT04755647	[416]
Phase2	Folic acid	Investigate the efficacy and safety of folic acid against DFU	30	Randomized	March 1, 2021	NCT04723134	[417]
NA	Celliant socks	Investigate the effect of Celliant socks against DFU	254	Randomized	June 1, 2021	NCT04709419	[418]
NA	Hybrid scale fiber matrix	Investigate the effect of hybrid-scale fiber matrix against DFU	40	Randomized	May 5, 2021	NCT04918784	[419]
Phase3	Umbical cord blood mononuclear cell gel	To investigate the effect of umbical cord blood mononuclear cell gel against DFU	40	Randomized	March 30, 2021	NCT04689425	[420]

storage instability, toxicity, poor drug loading and yield, poor site specificity and expensive manufacturing procedures restrict the use of nanocarriers/nanomaterials clinically. However, it is an in-escape trend for the researcher to further harness the potential of these delivery systems. Outlooking at the technical problems thereby, NDDS provide tangible advantages to improve patient compliance. In future, one should look for overcoming the aforementioned barriers and regulatory issues related to NDDS so that their better translation in market should be achieved. The conduct of exploratory clinical trials could further help in resolving these issues. These delivery systems are bound to establish the most favorable and economic therapies to treat the DW in the future.

Declaration of competing interest

Declared none.

References

- M.N. Piero, J.M. Gm Njagi, Diabetes mellitus-a devastating metabolic disorder, Asian J. Biomed. Pharm. Sci. 5 (2015) 1–7.
- [2] P.I. Saeedi P, P. Salpea, B. Malanda, S. Karuranga, N. Unwin, S. Colagiuri, L. Guariguata, A.A. Motala, K. Ogurtsova, J.E. Shaw, Global and regional diabetes

- prevalence estimates for 2019 and projections for 2030 and 2045: results from the international diabetes federation diabetes Atlas, Diabetes Res. Clin. Pract. 157 (2019), 107843.
- [3] C. Faselis, A. Katsimardou, K. Imprialos, P. Deligkaris, M. Kallistratos, K. Dimitriadis, Microvascular complications of type 2 diabetes mellitus, Curr. Vasc. Pharmacol. 18 (2020) 117–124.
- [4] A. Pfannkuche, A. Alhajjar, A. Ming, I. Walter, C. Piehler, P. Mertens, Prevalence and risk factors of diabetic peripheral neuropathy in a diabetics cohort: register initiative "diabetes and nerves", Endocr. Metabol. Sci. 1 (2020), 100053.
- [5] X.X. Zhang, J. Kong, K. Yun, Prevalence of diabetic nephropathy among patients with type 2 diabetes mellitus in China: a meta-analysis of observational studies, J. Diabetes Res. 2020 (2020), 2315607.
- [6] B.R. Sabanayagam C, M.L. Chee, R. Lee, Y.X. Wang, G. Tan, J.B. Jonas, E. L. Lamoureux, C.Y. Cheng, B.E. Klein, P. Mitchell, Incidence and progression of diabetic retinopathy: a systematic review, Lancet Diabetes Endocrinol. 7 (2019) 140, 140
- [7] S.a. Guo, L.A. DiPietro, Factors affecting wound healing, J. Dent. Res. 89 (2010) 219–229.
- [8] W.M. Wang C, T. Xu, X. Zhang, C. Lin, W. Gao, H. Xu, B. Lei, C. Mao, Engineering bioactive self-healing antibacterial exosomes hydrogel for promoting chronic diabetic wound healing and complete skin regeneration, Theranostics 9 (2019) 65.
- [9] A. Awasthi, B. Kumar, M. Gulati, R. Kumar, S. Wadhwa, R. Khursheed, L. Corrie, Kr Arya, R. Kumar, P. Patni, J. Kaur, S. Vishwas, A. Yadav, Treatment strategies against diabetic foot ulcer: success so far and the road ahead, Curr. Diabetes Rev. 17 (2021) 421–436.

- [10] S.R. Alavi A, D. Mayer, L. Goodman, M. Botros, D.G. Armstrong, K. Woo, T. Boeni, E.A. Ayello, R.S. Kirsner, Diabetic foot ulcers: Part I. Pathophysiology and prevention, J. Am. Acad. Dermatol. 70 (2014), 1. e1-1. e18.
- [11] J. Xu, W. Wu, L. Zhang, W. Dorset-Martin, M.W. Morris, M.E. Mitchell, K. W. Liechty, The role of microRNA-146a in the pathogenesis of the diabetic wound-healing impairment: correction with mesenchymal stem cell treatment, Diabetes 61 (2012) 2906–2912.
- [12] D. Li, A. Wang, X. Liu, F. Meisgen, J. Grünler, I.R. Botusan, S. Narayanan, E. Erikci, X. Li, L. Blomqvist, L. Du, MicroRNA-132 enhances transition from inflammation to proliferation during wound healing, J. Clin. Invest. 125 (2015) 3008–3026.
- [13] J. Ye, Y. Kang, S. Xiaofang, Pengwen Ni, Minjie Wu, Shuliang Lu, MicroRNA-155 inhibition promoted wound healing in diabetic rats, Int. J. Low. Extrem. Wounds 16 (2017) 74–84.
- [14] S. Dangwal S Bernd, C. Bang, J.M. Lorenzen, R. Kumarswamy, J. Fiedler, C. S. Falk, C.J. Scholz, T. Thum, D. Tschoepe, Impairment of wound healing in patients with type 2 diabetes mellitus influences circulating microRNA patterns via inflammatory cytokines, Arterioscler. Thromb. Vasc. Biol. 35 (2015) 1480–1488.
- [15] D.F. Ozdemir, W. Mark, MicroRNAs in diabetic wound healing: pathophysiology and therapeutic opportunities, Trends Cardiovasc. Med. 29 (2019) 131–137.
- [16] J. Natarajan, B.K.R. Sanapalli, M. Bano, S.K. Singh, M. Gulati, V.V.S.R. Karri, Nanostructured lipid carriers of pioglitazone loaded collagen/chitosan composite scaffold for diabetic wound healing, Adv. Wound Care 8 (2019) 499–513.
- [17] W.D.D. Aumiller, Harry Anderson, Pathogenesis and management of diabetic foot ulcers, JAAPA 28 (2015) 28–34.
- [18] T.A. Wilgus, New mechanisms of ECM production during wound healing: a role for parathyroid hormone 2 receptor signaling, J. Invest. Dermatol. 137 (2017) 1617–1619
- [19] M. Chang, Restructuring of the extracellular matrix in diabetic wounds and healing: a perspective, Pharmacol. Res. 107 (2016) 243–248.
- [20] H.V. Ezhilarasu, D. Dheen, S.T. Bay, B.-H. Srinivasan, D. Kumar, Nanoparticle-based therapeutic approach for diabetic wound healing, Nanomaterials 10 (2020) 1234.
- [21] M.A. Zubair, Role of growth factors and cytokines in diabetic foot ulcer healing: a detailed review, Rev. Endocr. Metab. Disord. 20 (2019) 207–217.
- [22] S. Lin, Q. Zhang, S. Li, T. Zhang, L. Wang, X. Qin, M. Zhang, S. Shi, X. Cai, Antioxidative and angiogenesis-promoting effects of tetrahedral framework nucleic acids in diabetic wound healing with activation of the Akt/Nrf2/HO-1 pathway, ACS Appl. Mater. Interfaces 12 (2020) 11397–11408.
- [23] Y. Li, Y. Tang, S. Shi, S. Gao, Y. Wang, D. Xiao, T. Chen, Q. He, J. Zhang, Y. Lin, Tetrahedral framework nucleic acids ameliorate insulin resistance in type 2 diabetes mellitus via the PI3K/Akt pathway, ACS Appl. Mater. Interfaces 13 (2021) 40354–40364.
- [24] P.S. Randeria, M.A. Seeger, X.-Q. Wang, H. Wilson, D. Shipp, C.A. Mirkin, A. S. Paller, siRNA-based spherical nucleic acids reverse impaired wound healing in diabetic mice by ganglioside GM3 synthase knockdown, Proc. Natl. Acad. Sci. U. S. A. 112 (2015) 5573–5578.
- [25] K. Rapala, The effect of tumor necrosis factor-alpha on wound healing. An experimental study, Annales chirurgiae et gynaecologiae, Supplement 211 (1996) 1–53
- [26] P.S. Rabbani, A. Zhou, Z.M. Borab, J.A. Frezzo, N. Srivastava, H.T. More, W. J. Rifkin, J.A. David, S.J. Berens, R. Chen, Novel lipoproteoplex delivers Keap1 siRNA based gene therapy to accelerate diabetic wound healing, Biomater. Sci. 132 (2017) 1–15.
- [27] L.N. Kasiewicz, K.A. Whitehead, Ipid nanoparticles silence tumor necrosis factor α to improve wound healing in diabetic mice, Bioeng 4 (2019) 75–82.
- [28] P.S. Rabbani, M.A. Soares, S.G. Hameedi, R.L. Kadle, A. Mubasher, M. Kowzun, D. Ceradini, Dysregulation of Nrf2/Keap1 redox pathway in diabetes affects multipotency of stromal cells, Diabetes 68 (2019) 141–155.
- [29] A. Rebane, T. Runnel, A. Aab, J. Maslovskaja, B. Rückert, M. Zimmermann, M. Plaas, J. Kärner, A. Treis, M. Pihlap, MicroRNA-146a alleviates chronic skin inflammation in atopic dermatitis through suppression of innate immune responses in keratinocytes. Journal of allergy clinical immunology, J. Allergy Clin. Immunol. 134 (2014) 836–847, e811.
- [30] G.-J. Shi, G.-R. Shi, J.-Y. Zhou, W.-J. Zhang, C.-Y. Gao, Ya-P Jiang, Z.-G. Zi, H.-H. Zhao, Y. Yang, J.-Q. Yu, Involvement of growth factors in diabetes mellitus and its complications: a general review, Biomed. Pharmacother. 101 (2018) 510–527.
- [31] M.W. Tsang, W.K.R. Wong, C.S. Hung, K.-M. Lai, W. Tang, E.Y. Cheung, G. Kam, L. Leung, C.W. Chan, C.M. Chu, Human epidermal growth factor enhances healing of diabetic foot ulcers, Diabetes Care 26 (2003) 1856–1861.
- [32] H. Li, X. Fu, L. Zhang, Q. Huang, Z. Wu, T. Sun, Research of PDGF-BB gel on the wound healing of diabetic rats and its pharmacodynamics, J. Surg. Res. 145 (2008) 41–48.
- [33] B. Cheng, H.-W. Liu, X.-B. Fu, T.-Z. Sun, Z.-Y. Sheng, Recombinant human platelet-derived growth factor enhanced dermal wound healing by a pathway involving ERK and c-fos in diabetic rats, J. Dermatol. Sci. 45 (2007) 193–201.
- [34] F. Cianfarani, G. Zambruno, L. Brogelli, F. Sera, P.M. Lacal, M. Pesce, M. C. Capogrossi, C.M. Failla, M. Napolitano, T. Odorisio, Lacenta growth factor in diabetic wound healing: altered expression and therapeutic potential, Am. J. Pathol. 169 (2006) 1167–1182.
- [35] X. Q Liu, L.S. Duan, Y.Q. Chen, X. J Jin, N. N Zhu, X. Zhou, H.W. Wei, L. Yin, J. R. Guo, *IncRNA* MALAT1 accelerates wound healing of diabetic mice transfused with modified autologous blood via the HIF-1α signaling pathway, Mol. Ther. Nucleic Acids 17 (2019) 504–515.

- [36] T. Kawanabe, T. Kawakami, Y. Yatomi, S. Shimada, Y. Soma, Sphingosine 1-phosphate accelerates wound healing in diabetic mice, J. Dermatol. Sci. 48 (2007) 53–60.
- [37] S. Patel, S. Srivastava, M. Rawat Singh, D. Singh, Mechanistic insight into diabetic wounds: pathogenesis, molecular targets and treatment strategies to pace wound healing, Biomed. Pharmacother. 112 (2019), 108615.
- [38] J. Casqueiro, J. Casqueiro, C. Alves, Infections in patients with diabetes mellitus: a review of pathogenesis, Indian J. Endocrinol. Metab. 16 (2012) S27.
- [39] S. Heravi, F.M. Zakrzewski, K. Vickery, D. G Armstrong, H. Hu, Bacterial diversity of diabetic foot ulcers: current status and future prospectives, J. Clin. Med. 8 (2019) 1935.
- [40] E. Estelle, N. Mathioudakis, Update on management of diabetic foot ulcers, Ann. N. Y. Acad. Sci. 1411 (2018) 153–165.
- [41] https://emedicine.medscape.com/article/237378-medication (Accessed on 21/3/ 2022).
- [42] M. Edmonds, A. Foster, The use of antibiotics in the diabetic foot, Am. J. Surg. 187 (2004) S25–S28.
- [43] https://study.com/academy/lesson/vancomycin-mechanism-of-action-drug-int eractions-side-effects.html (Accesed on 21/3/2022).
- [44] F. Davani, M. Alishahi, M. Sabzi, M. Khorram, A. Arastehfar, K. Zomorodian, Dual drug delivery of vancomycin and imipenem/cilastatin by coaxial nanofibers for treatment of diabetic foot ulcer infections, Mater. Sci. Eng. C Mater. Biol. Appl. 123 (2021), 111975.
- [45] T. Huidi, P. Kangambega, L. Noubou, B. Becsangele, S. Fluieraru, L. Teot, Antibiotic therapy of diabetic foot infections: a systematic review of randomized controlled trials, Wound Repair Regen. 26 (2018) 381–391.
- [46] M. Safdari, E. Shakiba, S.H. Kiaie, A. Fattahi, Preparation and characterization of Ceftazidime loaded electrospun silk fibroin/gelatin mat for wound dressing, Fibers Polym. 17 (2016) 744–750.
- [47] T.F. Francisco, L. Jaime R, R. Marcelo, Pharmacological aspects and spectrum of action of ceftazidime-avibactam: a systematic review, Infection 46 (2018) 165-181
- [48] B. Lipsky, A. Berendt, H. Deery, J. Embil, W. Joseph, A. Karchmer, J. LeFrock, D. Lew, J. Mader, C. Norden, Diagnosis and treatment of diabetic foot infections, Plast. Reconstr. Surg. 117 (2006) 212S–238S.
- [49] https://go.drugbank.com/drugs/DB00319 (Accessed on 21/3/2022).
- [50] P.R. Sarah, M.C. Anthony, M.C. Alyssa, M. Claudio, C. Isache, A.J. Christopher, Comparison of acute kidney injury in patients prescribed vancomycin in combination with piperacillin–tazobactam or cefepime for diabetic foot infections, J. Invest. Med. 69 (2021) 893–896.
- [51] S. P. M. Eswarawaka, D. Anne, P. I Shah, N. Srivastava, Bacteriological profile of diabetic foot, Int. Surg. J. 8 (2021) 704–709.
- [52] A. Awasthi, S.K. Singh, B. Kumar, M. Gulati, R. Kumar, S. Wadhwa, R. Khursheed, L. Corrie, A. Kr, R. Kumar, Treatment strategies against diabetic foot ulcer: success so far and the road ahead. Curr. Diabetes Rev. 17 (2021) 421–436.
- [53] C. Liu, A. Bayer, S.E. Cosgrove, R.S. Daum, S.K. Fridkin, R.J. Gorwitz, S.L. Kaplan, A.W. Karchmer, D.P. Levine, B.E. Murray, Clinical practice guidelines by the infectious diseases society of America for the treatment of methicillin-resistant Staphylococcus aureus infections in adults and children: executive summary, Clin. Infect. Dis. 52 (2011) 285–292.
- [54] N.D. Barwell, M.C. Devers, B. Kennon, H.E. Hopkinson, C. McDougall, M. J. Young, H.M. Robertson, D. Stang, S.J. Dancer, A. Seaton, Diabetic foot infection: antibiotic therapy and good practice recommendations, Int. J. Clin. Pract. 71 (2017), e13006.
- [55] M. Abbas, I. Uçkay, B.A. Lipsky, In diabetic foot infections antibiotics are to treat infection, not to heal wounds, Expet Opin. Pharmacother. 16 (2015) 1–12.
- [56] B.A. Lipsky, A.R. Berendt, P.B. Cornia, J.C. Pile, E.J. Peters, D.G. Armstrong, H. G. Deery, J.M. Embil, W.S. Joseph, A.W. Karchmer, Infectious Diseases Society of America clinical practice guideline for the diagnosis and treatment of diabetic foot infections, Clin. Infect. Dis. 54 (2012) e132–e173, 2012.
- [57] C. Kalaivani, G. Kuppusamy, V.V.S.R. Karri, Simvastatin loaded polycaprolactone-collagen scaffolds for the treatment of diabetic foot ulcer, Res. J. Pharm. Technol. 12 (2019) 2637–2644.
- [58] Abd Elmohsen, A. Fouad, D. H Abdelaziz, A histological and immunohistchemical study on the effect of atorvastatin versus adipose derived stem cells on cutaneous wound healing in diabetic male albino rats, Egypt, Jew Hist. 44 (2021) 96–112
- [59] M. Bagheri, B.M. Jahromi, H. Mirkhani, Z. Solhjou, A. Noorafshan, A. Zamani, Z. Amirghofran, zelnidipine, a new calcium channel blocker, promotes skin wound healing in diabetic rats, J. Surg. Res. 169 (2011) e101–e107.
- [60] S.F. Spampinato, G.I. Caruso, R. De Pasquale, M.A. Sortino, S. Merlo, The treatment of impaired wound healing in diabetes: looking among old drugs, Pharmaceuticals 13 (2020) 60.
- [61] R. Bhikha, J. Glynn, The pharmacological action of common herbal remedies, Amn.J. Int. Med. 6 (2018) 99–107.
- [62] A. Saeedi-Boroujeni, Natural remedies effective for the healing of diabetic wounds, J. Pharmaceut. Sci. Res. 9 (2017) 53–57.
- [63] J.A. Perini, T. Angeli-Gamba, J. Alessandra-Perini, L.C. Ferreira, L.E. Nasciutti, D. E. Machado, Opical application of Acheflan on rat skin injury accelerates wound healing: a histopathological, immunohistochemical and biochemical study, BMC Compl. Alternative Med. 15 (2015) 1–8.
- [64] T. Chakraborty, S. Gupta, A. Nair, S. Chauhan, V. Saini, Wound healing potential of insulin-loaded nanoemulsion with Aloe vera gel in diabetic rats, J. Drug Deliv. Sci. Technol. 64 (2021), 102601.

- [65] H. Maurya, M. Semwal, S.K. Dubey, Pharmacological evaluation of Chrozophora tinctoria as wound healing potential in diabetic rat's model, BioMed Res. Int. 20 (2016) 2016.
- [66] S. Hamedi, T. Shomali, A. Haghighat, Essential oil has protective effect against testicular damage in diabetic rats, J. Diet. Suppl. 15 (2018) 311–317.
- [67] R.H. Tuhin, M.M. Begum, M.S. Rahman, R. Karim, T. Begum, S.U. Ahmed, R. Mostofa, A. Hossain, M. Abdel-Daim, R. Begum, Wound healing effect of Euphorbia hirta linn.(Euphorbiaceae) in alloxan induced diabetic rats, BMC Compl. Alternative Med. 17 (2017) 1–14.
- [68] K.-M. Lau, K.-K. Lai, C.-L. Liu, J.C.-W. Tam, M.-H. To, H.-F. Kwok, C.-P. Lau, C.-H. Ko, P.-C. Leung, K.-P. Fung, Synergistic interaction between Astragali Radix and Rehmanniae Radix in a Chinese herbal formula to promote diabetic wound healing, J. Ethnopharmacol. 141 (2012) 250–256.
- [69] Y. R Kong, Y. Jong, M. Balakrishnan, Z. Bok, J. Weng, K. Tay, B. Goh, Beneficial role of Carica papaya extracts and phytochemicals on oxidative stress and related diseases: a mini review, Biology 10 (2021) 287.
- [70] Y. Huan, K.-j. Peng, Q.-l. Wang, Z.-y. Gu, Y.-q. Lu, Z. Jun, X. Fang, Y.-l. Liu, T. Ying, F-m. Deng, Effect of pomegranate peel polyphenol gel on cutaneous wound healing in alloxan-induced diabetic rats, Chin. Med. J. 126 (2013) 1700–1706.
- [71] S.P.A.S. Naddishani, L.D.A.M. Arawwawala, M.W.S.J. Kumari, P. Galappatthy, Effect of Curcuma longa L. and curcumin on diabetes and it's complications: a review, J. Ayurv. Herbal Med. 7 (2021) 109–118.
- [72] R. Khursheed, S.K. Singh, S. Wadhwa, M. Gulati, A. Awasthi, Enhancing the potential preclinical and clinical benefits of quercetin through novel drug delivery systems, Drug Discov. 25 (2020) 209–222.
- [73] R. Abdennabi, S. Bardaa, M. Mehdi, M.E. Rateb, A. Raab, F.N. Alenezi, Z. Sahnoun, N. Gharsallah, L. Belbahri, Phoenix dactylifera L. sap enhances wound healing in Wistar rats: phytochemical and histological assessment, Int. J. Biol. Macromol. 88 (2016) 443–450.
- [74] A.D. Kandhare, P. Ghosh, S.L. Bodhankar, Naringin, a flavanone glycoside, promotes angiogenesis and inhibits endothelial apoptosis through modulation of inflammatory and growth factor expression in diabetic foot ulcer in rats, Chem. Biol. Interact. 219 (2014) 101–112.
- [75] W. Li, A.D. Kandhare, A.A. Mukherjee, S.L. Bodhankar, Hesperidin, a plant flavonoid accelerated the cutaneous wound healing in streptozotocin-induced diabetic rats: role of TGF-B/Smads and Ang-1/Tie-2 signaling pathways, EXCLI J 17 (2018) 399–414.
- [76] S. Lodhi, A.K. Singhai, Wound healing effect of flavonoid rich fraction and luteolin isolated from Martynia annua Linn. on streptozotocin induced diabetic rats, Asian Pac. J. Trop. Med. 6 (2013) 253–259.
- [77] T. Ponrasu, L. Suguna, Efficacy of Annona squamosa on wound healing in streptozotocin-induced diabetic rats, Int. Wound J. 9 (2012) 613–623.
- [78] V. Kant, A. Gopal, D. Kumar, N.N. Pathak, M. Ram, B.L. Jangir, S.K. Tandan, D. Kumar, Curcumin-induced angiogenesis hastens wound healing in diabetic rats, J. Surg. Res. 193 (2015) 978–988.
- [79] J. Fu, J. Huang, M. Lin, T. Xie, T. You, Quercetin promotes diabetic wound healing via switching macrophages from M1 to M2 polarization, J. Surg. Res. 246 (2020) 213–223
- [80] R.I. Yassien, DE El-Ghazouly, The role of hesperidin on healing an incised wound in an experimentally induced diabetic adult male albino rats, Egypt J. Histol. 44 (2021) 144–162.
- [81] A.M. El Sayed, S.M. Ezzat, M.M. El Naggar, S.S. El Hawary, n vivo diabetic wound healing effect and HPLC-DAD-ESI-MS/MS profiling of the methanol extracts of eight Aloe species, Rev. Bras. Farmacogn. 26 (2016) 352–362.
- [82] O. Karaaslan, Y. Kankaya, N. Sungur, U. Kocer, S.S. Cuzdart, B. Sahin, A. Uysal, Case series of topical and orally administered β-glucan for the treatment of diabetic wounds: clinical study, J. Cutan. Med. Surg. 16 (2012) 180–186.
- [83] R. Weller, M.J. Finnen, The effects of topical treatment with acidified nitrite on wound healing in normal and diabetic mice, Nitric Oxide 15 (2006) 395–399.
- [84] J. Asai, H. Takenaka, S. Hirakawa, J.-i. Sakabe, A. Hagura, S. Kishimoto, K. Maruyama, K. Kajiya, S. Kinoshita, Y. Tokura, Topical simvastatin accelerates wound healing in diabetes by enhancing angiogenesis and lymphangiogenesis, Am. J. Pathol. 181 (2012) 2217–2224.
- [85] T.S. Lin, A. Abd Latiff, N.A. Abd Hamid, M. Mazlan, Evaluation of topical tocopherol cream on cutaneous wound healing in streptozotocin-induced diabetic rats, Evid 2012 (2012), 491027.
- [86] G. Wang, W. Li, Q. Chen, Y. Jiang, X. Lu, X. Zhao, Hydrogen sulfide accelerates wound healing in diabetic rats. International journal of clinical experimental pathology, Int. J. Clin. Exp. Pathol. 8 (2015) 5097.
- [87] Z. Zheng, Y. Liu, Y. Yang, J. Tang, B. Cheng, Opical 1% propranolol cream promotes cutaneous wound healing in spontaneously diabetic mice, Wound Repair Regen. 25 (2017) 389–397.
- [88] P.J. McLaughlin, J.D. Cain, M.B. Titunick, J.W. Sassani, I.S. Zagon, Topical naltrexone is a safe and effective alternative to standard treatment of diabetic wounds, Adv. Wound Care 6 (2017) 279–288.
- [89] H. El Banna, H. El Zorba, A. Hossny, W.J.I. Kamel, Comparative efficacy of grotto cream with Fucidin cream on normal and diabetic wound models in rats, Indian J. Physiol. Pharmacol. 62 (2018) 80–86.
- [90] K.S. Bhat, B. Vishwesh, M. Sahu, V.K. Shukla, A clinical study on the efficacy of Panchavalkala cream in Vrana Shodhana wsr to its action on microbial load and wound infection, Ayu 35 (2014) 135–140.
- [91] J.A. Perini, T. Angeli-Gamba, J. Alessandra-Perini, L.C. Ferreira, L.E. Nasciutti, D. E. Machado, Topical application of Acheflan on rat skin injury accelerates wound healing: a histopathological, immunohistochemical and biochemical study, BMC Compl. Alternative Med. 15 (2015) 203.

- [92] M.S. Hotkar, A.M. Avachat, S.S. Bhosale, Y.M. Oswal, Preliminary investigation of topical nitroglycerin formulations containing natural wound healing agent in diabetes-induced foot ulcer, Int. Wound J. 12 (2015) 210–217.
- [93] S. Yadollah-Damavandi, M. Chavoshi-Nejad, E. Jangholi, N. Nekouyian, S. Hosseini, A. Seifaee, S. Rafiee, H. Karimi, S. Ashkani-Esfahani, Y. Parsa, Opical Hypericum perforatum improves tissue regeneration in full-thickness excisional wounds in diabetic rat model, Evid. Based Complement Alternat. Med. (2015), 245328, 2015.
- [94] H. Maurya, M. Semwal, S.K. Dubey, Pharmacological evaluation of Chrozophora tinctoria as wound healing potential in diabetic rat's model, BioMed Res. Int. (2016), 7475124, 2016.
- [95] E. Mansouri, A. Hardani, M.R. Afzalzadeh, Combined effects of retinoic acid and hydro-alcoholic extract of Rosa damascena mill on wound in diabetic rats, Iran. J. Pharm. Res. (IJPR) 15 (2016) 583–589.
- [96] R.H. Tuhin, M.M. Begum, M.S. Rahman, R. Karim, T. Begum, S.U. Ahmed, R. Mostofa, A. Hossain, M. Abdel-Daim, R. Begum, Wound healing effect of Euphorbia hirta linn.(Euphorbiaceae) in alloxan induced diabetic rats, BMC Compl. Alternative Med. 17 (2017) 423.
- [97] S. Dehghani, R. Dalirfardouei, M.H. Jafari Najaf Abadi, M. Ebrahimi Nik, M. R. Jaafari, E. Mahdipour, Topical application of curcumin regulates the angiogenesis in diabetic-impaired cutaneous wound, Cell Biochem. Funct. 38 (2020) 558–566.
- [98] V. Falanga, S. Iwamoto, M. Chartier, T. Yufit, J. Butmarc, N. Kouttab, D. Shrayer, P. Carson, Autologous bone marrow-derived cultured mesenchymal stem cells delivered in a fibrin spray accelerate healing in murine and human cutaneous wounds, Tissue Eng. 13 (2007) 1299–1312.
- [99] P. Velander, C. Theopold, O. Bleiziffer, J. Bergmann, H. Svensson, Y. Feng, E. Eriksson, Cell suspensions of autologous keratinocytes or autologous fibroblasts accelerate the healing of full thickness skin wounds in a diabetic porcine wound healing model, J. Surg. Res. 157 (2009) 14–20.
- [100] M.K. Maharlooei, M. Bagheri, Z. Solhjou, B.M. Jahromi, M. Akrami, L. Rohani, A. Monabati, A. Noorafshan, G.R. Omrani, Adipose tissue derived mesenchymal stem cell (AD-MSC) promotes skin wound healing in diabetic rats, Diabetes Res. Clin. Pract. 93 (2011) 228–234.
- [101] K.B. Lee, J. Choi, S.B. Cho, J.Y. Chung, E.S. Moon, N.S. Kim, H.J. Han, Topical embryonic stem cells enhance wound healing in diabetic rats, J. Orthop. Res. 29 (2011) 1554–1562.
- [102] C. Fong, K. Gauthaman, S. Cheyyatraivendran, H. Lin, A. Biswas, A. Bongso, Human umbilical cord Wharton's jelly stem cells and its conditioned medium support hematopoietic stem cell expansion ex vivo, J. Cell. Biochem. 113 (2012) 658–668.
- [103] S. Kazemi-Darabadi, F. Sarrafzadeh-Rezaei, A.-A. Farshid, B. Dalir-Naghadeh, Allogenous skin fibroblast transplantation enhances excisional wound healing following alloxan diabetes in sheep, a randomized controlled trial, Int. J. Surg. 12 (2014) 751–756.
- [104] J.E. Jackson, Z. Kopecki, A.J. Cowin, Nanotechnological advances in cutaneous medicine, J. Nanomater. (2013), 808234, 2013.
- [105] B. Mordorski, J. Rosen, A. Friedman, Nanotechnology as an innovative approach for accelerating wound healing in diabetes, Diabetes Manag. 5 (2015) 329–332.
- [106] H. Choudhury, M. Pandey, Y.Q. Lim, C.Y. Low, C.T. Lee, T.C.L. Marilyn, H.S. Loh, Y.P. Lim, C.F. Lee, S.K. Bhattamishra, Silver nanoparticles: advanced and promising technology in diabetic wound therapy, Mater. Sci. Eng. C 112 (2020), 110925.
- [107] Y. Zhang, P. Zhang, X. Gao, L. Chang, Z. Chen, X.J.M.S. Mei, Preparation of exosomes encapsulated nanohydrogel for accelerating wound healing of diabetic rats by promoting angiogenesis, Mater. Sci. Eng. C Mater. Biol. Appl. 120 (2021), 111671.
- [108] J.U. Choi, S.W. Lee, R. Pangeni, Y. Byun, I.-S. Yoon, J.W. Park, Preparation and in vivo evaluation of cationic elastic liposomes comprising highly skin-permeable growth factors combined with hyaluronic acid for enhanced diabetic woundhealing therapy, Acta Biomater. 57 (2017) 197–215.
- [109] A. Valizadeh, M. Shirzad, M.R. Pourmand, M. Farahmandfar, H. Sereshti, A.J.D. D. Amani, Levofloxacin nanoemulsion gel has a powerful healing effect on infected wound in streptozotocin-induced diabetic rats, Drug Deliv. Transl. Res. 11 (2021) 292–304.
- [110] A. Ahmed, G. Getti, J. Boateng, Ciprofloxacin-loaded calcium alginate wafers prepared by freeze-drying technique for potential healing of chronic diabetic foot ulcers, Drug Deliv. Transl. Res. 8 (2018) 1751–1768.
- [111] V.T. Arantes, A.A. Faraco, F.B. Ferreira, C.A. Oliveira, E. Martins-Santos, P. Cassini-Vieira, L.S. Barcelos, L.A. Ferreira, G.A. Goulart, Etinoic acid-loaded solid lipid nanoparticles surrounded by chitosan film support diabetic wound healing in in vivo study, Colloids Surf. B Biointerfaces 188 (2020), 110749.
- [112] G. Gainza, D.C. Bonafonte, B. Moreno, J.J. Aguirre, F.B. Gutierrez, S. Villullas, J. L. Pedraz, M. Igartua, R.M. Hernandez, The topical administration of rhEGF-loaded nanostructured lipid carriers (rhEGF-NLC) improves healing in a porcine full-thickness excisional wound model, J. Contr. Release 197 (2015) 41–47.
- [113] F. Liu, X. Li, L. Wang, X. Yan, D. Ma, Z. Liu, X. Liu, Sesamol incorporated cellulose acetate-zein composite nanofiber membrane: an efficient strategy to accelerate diabetic wound healing, Int. J. Biol. Macromol. 149 (2020) 627–638.
- [114] Y. Jiang, Y. Li, J. Li, Y. Han, P. Zhang, Z. Yi, Q. Ke, H. Xu, A mussel-inspired extracellular matrix-mimicking composite scaffold for diabetic wound healing, ACS Appl. Bio Mater. 3 (2020) 4052–4061.
- [115] N.K. Rajendran, S.S.D. Kumar, N.N. Houreld, H. Abrahamse, A review on nanoparticle based treatment for wound healing, J. Drug Deliv. Sci. Technol. 44 (2018) 421–430.

- [116] R. Goyal, L.K. Macri, H.M. Kaplan, J. Kohn, Nanoparticles and nanofibers for topical drug delivery, J. Contr. Release 240 (2016) 77–92.
- [117] S. Jain, V. Jain, S. Mahajan, Lipid based vesicular drug delivery systems, Adv. Pharm. 2014 (2014).
- [118] A. Gabizon, H. Shmeeda, A.T. Horowitz, S. Zalipsky, Tumor cell targeting of liposome-entrapped drugs with phospholipid-anchored folic acid-PEG conjugates, Adv. Drug Deliv. Rev. 56 (2004) 1177–1192.
- [119] Y. Dai, R. Zhou, L. Liu, Y. Lu, J. Qi, W. Wu, Liposomes containing bile salts as novel ocular delivery systems for tacrolimus (FK506): in vitro characterization and improved corneal permeation, Int. J. Nanomed. 8 (2013) 1921–1933.
- [120] M.A.P. Olekson, R. Faulknor, A. Bandekar, M. Sempkowski, H.C. Hsia, F. Berthiaume, DF-1 liposomes promote sustained cell proliferation in mouse diabetic wounds, Wound Repair Regen. 23 (2015) 711–723.
- [121] P.S. Rabbani, A. Zhou, Z.M. Borab, J.A. Frezzo, N. Srivastava, H.T. More, W. J. Rifkin, J.A. David, S.J. Berens, R. Chen, Novel lipoproteoplex delivers Keap1 siRNA based gene therapy to accelerate diabetic wound healing, Biomater 132 (2017) 1–15.
- [122] S. Chhibber, J. Kaur, S. Kaur, Liposome entrapment of bacteriophages improves wound healing in a diabetic mouse MRSA infection, Front. Microbiol. 9 (2018) 561
- [123] R.K. Thapa, K.L. Kiick, M.O. Sullivan, Encapsulation of collagen mimetic peptidetethered vancomycin liposomes in collagen-based scaffolds for infection control in wounds, Acta Biomater. 103 (2020) 115–128.
- [124] D.A. Bassma, H. Elwakil, Ahmed A. Hussein, Reem A. Harfoush, Yousry M. Gohar, Chitosan and liposomes nanoparticles encapsulated CinnamonExtract: antiproteolytic activity and wound healing efficiency of diabetic rats running head: chitosan vs liposomes nanoparticles as drug delivery carriers, CMU J. Nat. Sci. 19 (2020) 595–611.
- [125] J. Bhattacharyya, G. Mondal, K. Madhusudana, S.B. Agawane, S. Ramakrishna, S. R. Gangireddy, R.D. Madhavi, P.K. Reddy, V.R. Konda, S.R. Rao, Single subcutaneous administration of RGDK-lipopeptide: rhPDGF-B gene complex heals wounds in streptozotocin-induced diabetic rats, Mol. Pharm. 6 (2009) 918–927.
- [126] S. Rai, V. Pandey, G. Rai, Transfersomes as versatile and flexible nano-vesicular carriers in skin cancer therapy: the state of the art, Nano Rev. Exp. 8 (2017), 1325708.
- [127] A.K. Ramanunny, S. Wadhwa, M. Gulati, S.K. Singh, B. Kapoor, H. Dureja, D. K. Chellappan, K. Anand, K. Dua, R. Khursheed, Nanocarriers for treatment of dermatological diseases: principle, perspective and practices, Eur. J. Pharmacol. 890 (2021), 173691.
- [128] S.A. El-Gizawy, A. Nouh, S. Saber, A.Y. Kira, Deferoxamine-loaded transfersomes accelerates healing of pressure ulcers in streptozotocin-induced diabetic rat, J. Drug Deliv. Sci. Technol. 58 (2020), 101732.
- [129] M.S. El-Ridy, S.A. Yehia, I. Elsayed, M.M. Younis, R.F. Abdel-Rahman, M.A. El-Gamil, Metformin hydrochloride and wound healing: from nanoformulation to pharmacological evaluation, J. Liposome Res. 29 (2019) 343–356.
- [130] Y. Liang, L. Duan, J. Lu, J. Xia, Engineering exosomes for targeted drug delivery, Theranostics 11 (2021) 3183–3195.
- [131] P. Goodarzi, B. Larijani, S. Alavi-Moghadam, A. Tayanloo-Beik, F. Mohamadi-Jahani, N. Ranjbaran, M. Payab, K. Falahzadeh, M. Mousavi, B. Arjmand, Mesenchymal stem cells-derived exosomes for wound regeneration, Cell Biol. Transl. Med. 4 (2018) 119–131.
- [132] M. Yu, W. Liu, J. Li, J. Lu, H. Lu, W. Jia, F. Liu, Exosomes derived from atorvastatin-pretreated MSC accelerate diabetic wound repair by enhancing angiogenesis via AKT/eNOS pathway, Stem Cell Res. 11 (2020) 1–17.
- [133] N. Xu, L. Wang, J. Guan, C. Tang, N. He, W. Zhang, S. Fu, Wound healing effects of a Curcuma zedoaria polysaccharide with platelet-rich plasma exosomes assembled on chitosan/silk hydrogel sponge in a diabetic rat model, Int. J. Biol. Macromol. 117 (2018) 102–107.
- [134] Q. Lv, J. Deng, Y. Chen, Y. Wang, B. Liu, J. Liu, Engineered human adipose stemcell-derived exosomes loaded with miR-21-5p to promote diabetic cutaneous wound healing, Mol. Pharm. 17 (2020) 1723–1733.
- [135] N.K. Pandey, S.K. Singh, M. Gulati, B. Kumar, B. Kapoor, D. Ghosh, R. Kumar, R. Khursheed, A. Awasthi, G. Kuppusamy, Overcoming the dissolution rate, gastrointestinal permeability and oral bioavailability of glimepiride and simvastatin co-delivered in the form of nanosuspension and solid selfnanoemulsifying drug delivery system: a comparative study, J. Drug Deliv. Sci. Technol. 60 (2020), 102083.
- [136] J. Xu, S. Bai, Y. Cao, L. Liu, Y. Fang, J. Du, L. Luo, M. Chen, B. Shen, Q. Zhang, miRNA-221-3p in endothelial progenitor cell-derived exosomes accelerates skin wound healing in diabetic mice, Diabetas Metab. Synd. 13 (2020) 1259–1270.
- [137] T. Fukui, A.T. Kawaguchi, S. Takekoshi, M. Miyasaka, H. Sumiyoshi, R. Tanaka, Liposome-encapsulated hemoglobin accelerates skin wound healing in diabetic dB/dB mice, Artif. Organs 41 (2017) 319–326.
- [138] O.E. Kaymakcalan, A. Abadeer, J.W. Goldufsky, U. Galili, S.J. Karinja, X. Dong, J. L. Jin, A. Samadi, J.A. Spector, Topical α-gal nanoparticles accelerate diabetic wound healing, Exp. Dermatol. 29 (2020) 404–413.
- [139] J. Zhang, C. Chen, B. Hu, X. Niu, X. Liu, G. Zhang, C. Zhang, Q. Li, Y. Wang, Exosomes derived from human endothelial progenitor cells accelerate cutaneous wound healing by promoting angiogenesis through Erk1/2 signaling, Int. J. Biol. Sci. 12 (2016) 1472–1487.
- [140] R. Dalirfardouei, K. Jamialahmadi, A.H. Jafarian, E. Mahdipour, Promising effects of exosomes isolated from menstrual blood-derived mesenchymal stem cell on wound-healing process in diabetic mouse model. ournal of tissue engineering regenerative medicine, J. Tissue Eng. Regen. Med. 13 (2019) 555–568.

- [141] M. Li, T. Wang, H. Tian, G. Wei, L. Zhao, Y. Shi, Macrophage-derived exosomes accelerate wound healing through their anti-inflammation effects in a diabetic rat model, Artif. Cell Nanomed. Biotechnol. 47 (2019) 3793–3803.
- [142] C. Wang, M. Wang, T. Xu, X. Zhang, C. Lin, W. Gao, H. Xu, B. Lei, C. Mao, Engineering bioactive self-healing antibacterial exosomes hydrogel for promoting chronic diabetic wound healing and complete skin regeneration, Theranostics 9 (2019) 65–76.
- [143] J. Yang, Z. Chen, D. Pan, H. Li, J. Shen, Umbilical cord-derived mesenchymal stem cell-derived exosomes combined pluronic F127 hydrogel promote chronic diabetic wound healing and complete skin regeneration, Int. J. Nanomed. 15 (2020) 5911–5926.
- [144] J. Kuhn, A. Hassan, S. Sharma, J. Kwong, M. Rahman, S. Adam, A. Villarreal-Ponce, R.G. Pena, D.J. Ceradini, P.S. Rabbani, Human nrf2-active multipotent stromal cell exosomes reverse pathologic diabetic wound healing, Plast. Reconstr. Surg.-Glob. Open. 8 (2020) 111–112.
- [145] R. Shi, Y. Jin, W. Hu, W. Lian, C. Cao, S. Han, S. Zhao, H. Yuan, X. Yang, J. Shi, Exosomes derived from mmu_circ_0000250-modified adipose-derived mesenchymal stem cells promote wound healing in diabetic mice by inducing miR-128-3p/SIRT1-mediated autophagy, Am. J. Physiol. Cell Physiol. 318 (2020) C848-C856.
- [146] P. Wei, C. Zhong, X. Yang, F. Shu, S. Xiao, T. Gong, P. Luo, L. Li, Z. Chen, Y. Zheng, Exosomes derived from human amniotic epithelial cells accelerate diabetic wound healing via PI3K-AKT-mTOR-mediated promotion in angiogenesis and fibroblast function, Burns trauma 8 (2020), tkaa020.
- [147] Y. Xiong, L. Chen, T. Yu, C. Yan, W. Zhou, F. Cao, X. You, Y. Zhang, Y. Sun, J. Liu, Nhibition of circulating exosomal microRNA-15a-3p accelerates diabetic wound repair, Aging 12 (2020) 8968–8986.
- [148] Bs Li, S. Luan, J. Chen, Y. Zhou, T. Wang, Z. Li, Y. Fu, A. Zhai, C. Bi, The MSC-derived exosomal *IncRNA* H19 promotes wound healing in diabetic foot ulcers by upregulating PTEN via MicroRNA-152-3p, Mol. Ther. Nucleic Acids 19 (2020) 814–826.
- [149] W. Liu, M. Yu, D. Xie, L. Wang, C. Ye, Q. Zhu, F. Liu, L. Yang, Melatonin-stimulated MSC-derived exosomes improve diabetic wound healing through regulating macrophage M1 and M2 polarization by targeting the PTEN/AKT pathway, Stem Cell Res. Ther. 11 (2020) 1–15.
- [150] P.A. Shiekh, A. Singh, A. Kumar, Exosome laden oxygen releasing antioxidant and antibacterial cryogel wound dressing OxOBand alleviate diabetic and infectious wound healing, Biomaterials 249 (2020), 120020.
- [151] A. Kabalnov, Ostwald ripening and related phenomena, Technology 22 (2001) 1–12.
- [152] T. Tadros, P. Izquierdo, J. Esquena, C. Solans, Ormation and stability of nanoemulsions, Adv. Colloid Interface Sci. 108 (2004) 303–318.
- [153] T.G. Mason, J.N. Wilking, K. Meleson, C.B. Chang, S.M. Graves, Nanoemulsions: formation, structure, and physical properties, J. Phys. Condens. Matter 18 (2006) R635.
- [154] H.D. Silva, M.A. Cerqueira, A.A. Vicente, Influence of surfactant and processing conditions in the stability of oil-in-water nanoemulsions, J. Food Eng. 167 (2015) 89–98.
- [155] T. Kaur, D.N. Kapoor, Development and evaluation of sea buckthorn (Hippophae rhamnoides L.) seed oil nanoemulsion gel for wound healing, Pharmacom. Mag. 14 (2018) 647–658.
- [156] K. Shanmugapriya, H. Kim, H. W Kang, A new alternative insight of nanoemulsion conjugated with κ-carrageenan for wound healing study in diabetic mice: in vitro and in vivo evaluation, Eur. J. Pharmaceut. Sci. 133 (2019) 236–250.
- [157] J.-P. Jee, R. Pangeni, S.K. Jha, Y. Byun, J.W. Park, Preparation and in vivo evaluation of a topical hydrogel system incorporating highly skin-permeable growth factors, quercetin, and oxygen carriers for enhanced diabetic woundhealing therapy, Int. J. Nanomed. 14 (2019) 5449–5475.
- [158] G. Gundogdu, K.A. Nalci, A.B. Ugur Kaplan, K. Gundogdu, T. Demirci, F. Demirkaya Miloglu, A. Hacımuftuoglu, M. Cetin, He evaluation of the effects of nanoemulsion formulations containing boron and/or zinc on the wound healing in diabetic rats, Int. J. Low. Extrem. Wounds (2020), 1534734620961892.
- [159] S.-K. Chua, J.-Y. Fu, M.H. Zulfakar, M.-H. Ng, Z.A.A. Hasan, S.-F. Ng, Optimisation and biological evaluation of palm glyceryl monocaprylate antimicrobial nanoemulsion for combating S. aureus wound infection, J. Mater. Res. Technol. 9 (2020) 12804–12817.
- [160] A. Valizadeh, M. Shirzad, M.R. Pourmand, M. Farahmandfar, H. Sereshti, A. Amani, Levofloxacin nanoemulsion gel has a powerful healing effect on infected wound in streptozotocin-induced diabetic rats, Drug Deliv. Transl. Res. 11 (2021) 292–304.
- [161] N. Al-Manasir, A.L. Kjøniksen, B.o Nyström, Preparation and characterization of cross-linked polymeric nanoparticles for enhanced oil recovery applications, J. Appl. Polym. 113 (2009) 1916–1924.
- [162] C. Chang, B. Duan, J. Cai, L. Zhang, Superabsorbent hydrogels based on cellulose for smart swelling and controllable delivery, Eur. Polym. J. 46 (2010) 92–100.
- [163] H. Yadav, N. Al Halabi, G. Alsalloum, Nanogels as novel drug delivery systems-a review, J. Pharm. Pharmaceut. Res. 1 (2017) 1–8.
- [164] P. Thangavel, B. Ramachandran, S. Chakraborty, R. Kannan, S. Lonchin, V. Muthuvijayan, Accelerated healing of diabetic wounds treated with L-glutamic acid loaded hydrogels through enhanced collagen deposition and angiogenesis: an in vivo study, Sci. Rep. 7 (2017) 1–15.
- [165] K. Obara, M. Ishihara, T. Ishizuka, M. Fujita, Y. Ozeki, T. Maehara, Y. Saito, H. Yura, T. Matsui, H. Hattori, Photocrosslinkable chitosan hydrogel containing fibroblast growth factor-2 stimulates wound healing in healing-impaired db/db mice, Bioamter 24 (2003) 3437–3444.

- [166] P. Y Lee, Z. Li, L. Huang, Thermosensitive hydrogel as a Tgf-β1 gene delivery vehicle enhances diabetic wound healing, Pharm. Res. (N. Y.) 20 (2003) 1995–2000.
- [167] G. Lao, L. Yan, C. Yang, L. Zhang, S. Zhang, Y. Zhou, Controlled release of epidermal growth factor from hydrogels accelerates wound healing in diabetic rats, J. Am. Podiatr. Med. Assoc. 102 (2012) 89–98.
- [168] R.S. Pawar, S. Kumar, F.A. Toppo, P. Lakshmi, P. Suryavanshi, Sida cordifolia Linn. accelerates wound healing process in type 2 diabetic rats, J. Acute Med. 6 (2016) 82–89.
- [169] M. Hajimiri, S. Shahverdi, M.A. Esfandiari, B. Larijani, F. Atyabi, A. Rajabiani, A. R. Dehpour, M. Amini, R. Dinarvand, Preparation of hydrogel embedded polymergrowth factor conjugated nanoparticles as a diabetic wound dressing, Drug Dev. Ind. Pharm. 42 (2016) 707–719.
- [170] S. B Natarajan, S. K Das, S. P Chandran, A. M Oo, S. S Kanneppady, M. Entezarian, O. Lwin, Moringa Olifera leaf extract loaded hydrogel for diabetic wound healing, MJMR 2 (2018) 35–41.
- [171] H. Chen, R. Cheng, X. Zhao, Y. Zhang, A. Tam, Y. Yan, H. Shen, Y.S. Zhang, J. Qi, Y. Feng, An injectable self-healing coordinative hydrogel with antibacterial and angiogenic properties for diabetic skin wound repair, NPG Asia Mater. 11 (2019) 1, 12
- [172] P.S. Patil, S. Fathollahipour, A. Inmann, A. Pant, R. Amini, L.P. Shriver, N. Leipzig, Fluorinated methacrylamide chitosan hydrogel dressings improve regenerated wound tissue quality in diabetic wound healing, Adv. Wound Care 8 (2019) 374–385.
- [173] Y. Fan, W. Wu, Y. Lei, C. Gaucher, S. Pei, J. Zhang, X. Xia, Edaravone-loaded alginate-based nanocomposite hydrogel accelerated chronic wound healing in diabetic mice, Mar. Drugs 17 (2019) 285.
- [174] G. Gallelli, E. Cione, R. Serra, A. Leo, R. Citraro, P. Matricardi, C. Di Meo, F. Bisceglia, M.C. Caroleo, S. Basile, Nano-hydrogel embedded with quercetin and oleic acid as a new formulation in the treatment of diabetic foot ulcer: a pilot study, Int. Wound J. 17 (2020) 485–490.
- [175] T. Wang, Q. Liao, Y. Wu, X. Wang, C. Fu, F. Geng, Y. Qu, J. Zhang, A composite hydrogel loading natural polysaccharides derived from Periplaneta americana herbal residue for diabetic wound healing, Int. J. Biol. Macromol. 164 (2020) 3846–3857.
- [176] D.Y. Shin, J.-U. Park, M.-H. Choi, S. Kim, H.-E. Kim, S.-H. Jeong, Polydeoxyribonucleotide-delivering therapeutic hydrogel for diabetic wound healing, Sci. Rep. 10 (2020) 1–14.
- [177] K.S.B. Masters, S.J. Leibovich, P. Belem, J.L. West, L.A. Poole-Warren, Effects of nitric oxide releasing poly (vinyl alcohol) hydrogel dressings on dermal wound healing in diabetic mice, Wound Repair Regen. 10 (2002) 286–294.
- [178] K. Obara, M. Ishihara, M. Fujita, Y. Kanatani, H. Hattori, T. Matsui, B. Takase, Y. Ozeki, S. Nakamura, T. Ishizuka, Acceleration of wound healing in healing-impaired db/db mice with a photocrosslinkable chitosan hydrogel containing fibroblast growth factor-2, Wound Repair Regen. 13 (2005) 390–397.
- [179] T. Tokatlian, C. Cam, T. Segura, Orous hyaluronic acid hydrogels for localized nonviral DNA delivery in a diabetic wound healing model, Adv. Healthc. Mater. 4 (2015) 1084–1091.
- [180] D.S. Yoon, Y. Lee, H.A. Ryu, Y. Jang, K.-M. Lee, Y. Choi, W.J. Choi, M. Lee, K. M. Park, K. Park, J. Lee, Cell recruiting chemokine-loaded sprayable gelatin hydrogel dressings for diabetic wound healing, Acta Biomater. 38 (2016) 59–68.
- [181] Y. Xiao, L.A. Reis, N. Feric, E.J. Knee, J. Gu, S. Cao, C. Laschinger, C. Londono, J. Antolovich, A. McGuigan, M. Radisic, Diabetic wound regeneration using peptide-modified hydrogels to target re-epithelialization, Proc. Natl. Acad. Sci. U. S. A. 113 (2016) E5792–E5801
- [182] J.S. Choi, D.W. Kim, D.S. Kim, J.O. Kim, C.S. Yong, K.H. Cho, Y.S. Youn, S.G. Jin, H.-G. Choi, Novel neomycin sulfate-loaded hydrogel dressing with enhanced physical dressing properties and wound-curing effect, Drug Deliv. 23 (2016) 2806–2812.
- [183] G. Tronci, J. Yin, R.A. Holmes, H. Liang, S.J. Russell, D. J Wood, Protease-sensitive atelocollagen hydrogels promote healing in a diabetic wound model, J. Mater. Chem. B 4 (2016) 7249–7258.
- [184] L. Zhao, L. Niu, H. Liang, H. Tan, C. Liu, F. Zhu, pH and glucose dual-responsive injectable hydrogels with insulin and fibroblasts as bioactive dressings for diabetic wound healing, ACS Appl. Mater. Interfaces 9 (2017) 37563–37574.
- [185] N. Wathoni, A. Hasanah, A.F.A. Mohammed, E. Pratiwi, R. Mahmudah, Accelerated wound healing ability of sacran hydrogel film by keratinocyte growth factor in alloxan-induced diabetic mice, Int. J. Appl. Pharm. 10 (2018) 57–61.
- [186] Q. Xu, A. Sigen, Y. Gao, L. Guo, J. Creagh-Flynn, D. Zhou, U. Greiser, Y. Dong, F. Wang, H. Tai, W. Liu, W. Wang, W. Wang, A hybrid injectable hydrogel from hyperbranched PEG macromer as a stem cell delivery and retention platform for diabetic wound healing, Acta Biomater. 75 (2018) 63–74.
- [187] A. Cifuentes, V. Gómez-Gil, M.A. Ortega, Á. Asúnsolo, S. Coca, J. San Román, M. Álvarez-Mon, J. Buján, N. García-Honduvilla, Biomed. Pharmacother. 129 (1) (2020), 10498.
- [188] J. Zhu, G. Jiang, W. Hong, Y. Zhang, B. Xu, G. Song, T. Liu, C. Hong, L. Ruan, Rapid gelation of oxidized hyaluronic acid and succinyl chitosan for integration with insulin-loaded micelles and epidermal growth factor on diabetic wound healing, Mater. Sci. Eng. C Mater. Biol. Appl. 117 (2020), 111273.
- [189] C.-M. Hsieh, W. Wang, Y.-H. Chen, P.-S. Wei, Y.-H. Liu, M.-T. Sheu, H. Ho, A novel composite hydrogel composed of formic acid-decellularized pepsin-soluble extracellular matrix hydrogel and sacchachitin hydrogel as wound dressing to synergistically accelerate diabetic wound healing, Pharm. Times 12 (2020) 538.
- [190] P. Zhang, L. He, J. Zhang, X. Mei, Y. Zhang, H. Tian, Z. Chen, Preparation of novel berberine nano-colloids for improving wound healing of diabetic rats by acting Sirt1/NF-κB pathway, Colloids Surf. B Biointerfaces 187 (2020), 110647.

- [191] Y. Zhao, Z. Li, Q. Li, L. Yang, H. Liu, R. Yan, L. Xiao, H. Liu, J. Wang, B. Yang, Q. Lin, Transparent conductive Supramolecular hydrogels with Stimuliresponsive properties for on-demand dissolvable diabetic foot wound dressings, Macromol. Rapid Commun. 41 (2020), 2000441.
- [192] M. Yin, X. Wang, Z. Yu, Y. Wang, X. Wang, M. Deng, D. Zhao, S. Ji, N. Jia, W. Zhang, γ-PGA hydrogel loaded with cell-free fat extract promotes the healing of diabetic wounds, J. Mater. Chem. B 8 (2020) 8395–8404.
- [193] Y. Zhang, P. Zhang, X. Gao, L. Chang, Z. Chen, X. Mei, Preparation of exosomes encapsulated nanohydrogel for accelerating wound healing of diabetic rats by promoting angiogenesis, Mater. Sci. Eng. C Mater. Biol. Appl. 120 (2021), 111671.
- [194] L. Sheng, Z. Zhang, Y. Zhang, E. Wang, B. Ma, Q. Xu, L. Ma, M. Zhang, G. Pei, J. Chang, A novel "hot spring"-mimetic hydrogel with excellent angiogenic properties for chronic wound healing, Biomater 264 (2021), 120414.
- [195] S. Mukherjee, S. Ray, R.S. Thakur, Solid lipid nanoparticles: a modern formulation approach in drug delivery system, Indian J. Pharmaceut. Sci. 71 (2009) 349–358.
- [196] A. Y. Mu Hu, Recent advances in drug delivery applications of cubosomes, hexosomes, and solid lipid nanoparticles, Acta Pharm. Sin. B 11 (2021) 871–885.
- [197] V. Mishra, K.K. Bansal, A. Verma, N. Yadav, S. Thakur, K. Sudhakar, J. M. Rosenholm, Olid lipid nanoparticles: emerging colloidal nano drug delivery systems, Pharm. Times 10 (2018) 191.
- [198] M. Üner, G. Yener, Importance of solid lipid nanoparticles (SLN) in various administration routes and future perspectives, Int. J. Nanomed. 2 (2007) 200, 200
- [199] G. Gainza, M. Pastor, J.J. Aguirre, S. Villullas, J.L. Pedraz, R.M. Hernandez, M. Igartua, A novel strategy for the treatment of chronic wounds based on the topical administration of rhEGF-loaded lipid nanoparticles: in vitro bioactivity and in vivo effectiveness in healing-impaired db/db mice, J. Contr. Release 185 (2014) 51–61.
- [200] J. Rajesh, S.M. Lakshmi, K. Thamizhvanan, T. Viswasanthi, Formulation, characterization and evaluation of methanolic extract of Abutilon indicum loaded solid lipid nanoparticles against microorganisms causing diabetic foot and urinary tract infection, JGTPS 5 (2012) 2093–2102.
- [201] L.N. Kasiewicz, K.A. Whitehead, Ipid nanoparticles silence tumor necrosis factor α to improve wound healing in diabetic mice, Bioeng. Transl. Med. 4 (2019) 75–82.
- [202] V.T. Arantes, A.A. Faraco, F.B. Ferreira, C.A. Oliveira, E. Martins-Santos, P. Cassini-Vieira, L.S. Barcelos, L.A. Ferreira, G.A. Goulart, Etinoic acid-loaded solid lipid nanoparticles surrounded by chitosan film support diabetic wound healing in in vivo study, Colloids Surf. B Biointerfaces 188 (2020), 110749.
- [203] A. Beloqui, M.Á. Solinís, A. Rodríguez-Gascón, A.J. Almeida, V. Préat, Nanostructured lipid carriers: promising drug delivery systems for future clinics, Nano 12 (2016) 143–161.
- [204] S.U. Rawal, M.M. Patel, Lipid Nanoparticulate Systems: Modern Versatile Drug Carriers, Lipid Nanocarriers for Drug Targeting, Elsevier, 2018, pp. 49–138.
- [205] K. Karn-Orachai, S.M. Smith, S. Phunpee, A. Treethong, S. Puttipipatkhachorn, S. Pratontep, U.R. Ruktanonchai, The effect of surfactant composition on the chemical and structural properties of nanostructured lipid carriers, J. Microencapsul. 31 (2014) 609–618.
- [206] D. Orgul, H. Eroglu, S.J. Hekimoglu, Formulation and characterization of tissue scaffolds containing simvastatin loaded nanostructured lipid carriers for treatment of diabetic wounds, J. Drug Deliv. Sci. Technol. 41 (2017) 280–292.
- [207] A. Motawea, A.E.-G.H. Abd El, T. Borg, M. Motawea, M. Tarshoby, The impact of topical phenytoin loaded nanostructured lipid carriers in diabetic foot ulceration, Foot 40 (2019) 14–21.
- [208] J. Natarajan, B.K.R. Sanapalli, M. Bano, S.K. Singh, M. Gulati, V.V.S.R. Karri, Nanostructured lipid carriers of pioglitazone loaded collagen/chitosan composite scaffold for diabetic wound healing, Adv. Wound Care 8 (2019) 499–513.
- [209] M. Ghodrati, M.R. Farahpour, H. Hamishehkar, S.A. Physicochemical, E. Aspects, Encapsulation of Peppermint essential oil in nanostructured lipid carriers: in-vitro antibacterial activity and accelerative effect on infected wound healing, Colloids Surf. A Physicochem. Eng. Asp. 564 (2019) 161–169.
- [210] K. Khezri, M.R. Farahpour, S. Mounesi Rad, Accelerated infected wound healing by topical application of encapsulated Rosemary essential oil into nanostructured lipid carriers, Artif. Cell Nanomed. Biotechnol. 47 (2019) 980–988.
- [211] I. Garcia-Orue, G. Gainza, P. Garcia-Garcia, F.B. Gutierrez, J.J. Aguirre, R. M. Hernandez, A. Delgado, M. Igartua, Composite nanofibrous membranes of PLGA/Aloe vera containing lipid nanoparticles for wound dressing applications, Int. J. Pharm. 556 (2019) 320–329.
- [212] H.-J. Lee, M. Jeong, Y.-G. Na, S.-J. Kim, H.-K. Lee, C.-W. Cho, An EGF-and curcumin-Co-encapsulated nanostructured lipid carrier accelerates chronicwound healing in diabetic rats, Mol 25 (2020) 4610.
- [213] D. Sun, S.-y. Guo, L. Yang, Y.-r. Wang, X.-h. Wei, S. Song, Y.-w. Yang, Y. Gan, Z. Wang, Silicone elastomer gel impregnated with 20 (S)-protopanaxadiol-loaded nanostructured lipid carriers for ordered diabetic ulcer recovery, Acta Pharmacol. Sin. 41 (2020) 119–128.
- [214] K. Khezri, M.R. Farahpour, S. Rad, S.A. Physicochemical, E. Aspects, Efficacy of Mentha pulegium essential oil encapsulated into nanostructured lipid carriers as an in vitro antibacterial and infected wound healing agent, Colloids Surf. A Physicochem. Eng. Asp. 589 (2020), 124414.
- [215] S. Laurent, D. Forge, M. Port, A. Roch, C. Robic, L. Vander Elst, R.N. Muller, Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications, Chem. Rev. 108 (2008) 2064–2110.

- [216] J.N. Tiwari, R.N. Tiwari, K.S. Kim, Zero-dimensional, one-dimensional, two-dimensional and three-dimensional nanostructured materials for advanced electrochemical energy devices, Prog. Polym. Sci. 57 (2012) 724–803.
- [217] I. Khan, K. Saeed, I. Khan, Nanoparticles: properties, applications and toxicities, Arab, J. Chem. 12 (2019) 908–931.
- [218] N.K. Rajendran, S.S.D. Kumar, N.N. Houreld, H. Abrahamse, A review on nanoparticle based treatment for wound healing, J. Drug Delv. Sci. Technol. 44 (2018) 421–430.
- [219] S. Naraginti, P.L. Kumari, R.K. Das, A. Sivakumar, S.H. Patil, V.V. Andhalkar, Amelioration of excision wounds by topical application of green synthesized, formulated silver and gold nanoparticles in albino Wistar rats, Mater. Sci. Eng., C 62 (2016) 293–300.
- [220] S.A. Chen, H. M Chen, Y. D Yao, C.F. Hung, C.S. Tu, Y. J Liang, Topical treatment with anti-oxidants and Au nanoparticles promote healing of diabetic wound through receptor for advance glycation end-products, Eur. J. Pharmaceut. Sci. 47 (2012) 875–883.
- [221] J. Yu, X. Wang, Y. Li, X. Huang, X. Luo, X. He, Synthesis of nerolidol functionalized gold nanoparticles for wound regeneration in people with diabetic foot ulcers in nursing care management, Sci. Adv. Mater. 10 (2018) 1775–1781.
- [222] S. Wang, C. Yan, X. Zhang, D. Shi, L. Chi, G. Luo, J. Deng, Antimicrobial peptide modification enhances the gene delivery and bactericidal efficiency of gold nanoparticles for accelerating diabetic wound healing, Biomater. Sci. 6 (2018) 2757–2772.
- [223] M. Ponnanikajamideen, S. Rajeshkumar, M. Vanaja, G. Annadurai, n vivo type 2 diabetes and wound-healing effects of antioxidant gold nanoparticles synthesized using the insulin plant Chamaecostus cuspidatus in albino rats, Can. J. Diabetes 43 (2019) 82–89, e86.
- [224] S. C Wei, L. Chang, C.C. Huang, H. T Chang, Dual-functional gold nanoparticles with antimicrobial and proangiogenic activities improve the healing of multidrugresistant bacteria-infected wounds in diabetic mice, Biomater. Sci. 7 (2019) 4482–4490.
- [225] S. Li, Q. Tang, H. Xu, Q. Huang, Z. Wen, Y. Liu, C. Peng, Improved stability of KGF by conjugation with gold nanoparticles for diabetic wound therapy, Nanomed 14 (2019) 2909–2923.
- [226] S.P. Hernández Martínez, T.I. Rivera González, M.A. Franco Molina, J.J. Bollain y Goytia, J.J. Martínez Sanmiguel, D.G. Zárate Triviño, C. Rodríguez Padilla, A novel gold calreticulin nanocomposite based on chitosan for wound healing in a diabetic mice model, Nanomaterials 9 (2019) 75.
- [227] A.Y.H.N. Azlan, H. Katas, N.H. Habideen, M.F.M. Busra, Dual-action of thermoresponsive gels containing DsiRNA-loaded gold nanoparticles for diabetic wound therapy: characterization, in vitro safety and healing efficacy, SA Pharm. J. 28 (2020) 1420–1430.
- [228] M. Mohajeri-Tehrani, M. Aalaa, A. Aalaa, N. Rohipour, M. Sanjari, P. Khashayar, F. Shahcheraghi, M. Khashayarfard, R. Heshmat, S. Madani, Antimicrobial effects of silver nanoparticles coated leather on diabetic foot ulcer, Sci. Rep. 1 (2012) 1–3
- [229] S. Bhagavathy, S. Kancharla, Wound healing and angiogenesis of silver nanoparticle from Azadirachta indica in diabetes induced mice, Int. J. Herb. Med. 4 (2016) 24–29.
- [230] C. Almonaci Hernández, K. Juarez-Moreno, M. Castañeda-Juarez, H. Almanza-Reyes, A. Pestryakov, N. Bogdanchikova, Silver nanoparticles for the rapid healing of diabetic foot ulcers, Int. J. Med. Nano Res. 4 (2017) 2378–3664.
- [231] R. Singla, S. Soni, V. Patial, P.M. Kulurkar, A. Kumari, S. Mahesh, Y.S. Padwad, S. K. Yadav, In vivo diabetic wound healing potential of nanobiocomposites containing bamboo cellulose nanocrystals impregnated with silver nanoparticles, Int. J. Biol. Macromol. 105 (2017) 45–55.
- [232] G. Shi, W. Chen, Y. Zhang, X. Dai, X. Zhang, Z. Wu, An antifouling hydrogel containing silver nanoparticles for modulating the therapeutic immune response in chronic wound healing, Langmuir 35 (2018) 1837–1845.
- [233] N. Masood, R. Ahmed, M. Tariq, Z. Ahmed, M.S. Masoud, I. Ali, R. Asghar, A. Andleeb, A. Hasan, Silver nanoparticle impregnated chitosan-PEG hydrogel enhances wound healing in diabetes induced rabbits, Int. J. Biol. Macromol. 559 (2019) 23–36.
- [234] M. Kuwabara, Y. Sato, M. Ishihara, T. Takayama, S. Nakamura, K. Fukuda, K. Murakami, H. Yokoe, T. Kiyosawa, Healing of Pseudomonas aeruginosainfected wounds in diabetic db/db mice by weakly acidic hypochlorous acid cleansing and silver nanoparticle/chitin-nanofiber sheet covering, Wound Med. 28 (2020), 100183.
- [235] S.S.D. Kumar, N.N. Houreld, H. Abrahamse, Elective laser efficiency of green-synthesized silver nanoparticles by aloe arborescens and its wound healing activities in normal wounded and diabetic wounded fibroblast cells: in vitro studies, Int. J. Nanomed. 15 (2020) 6855–6870.
- [236] S.T. Appapalam, B. Paul, S. Arockiasamy, R. Panchamoorthy, Phytofabricated silver nanoparticles: discovery of antibacterial targets against diabetic foot ulcer derived resistant bacterial isolates, Mater. Sci. Eng. C Mater. Biol. Appl. 117 (2020), 111256.
- [237] Y. H Lee, Y. L Hong, T. L Wu, Novel silver and nanoparticle-encapsulated growth factor co-loaded chitosan composite hydrogel with sustained antimicrobility and promoted biological properties for diabetic wound healing, Mater. Sci. Eng. C Mater. Biol. Appl. 118 (2021), 111385.
- [238] M.B. Dreifke, A.A. Jayasuriya, A. Jayasuriya, Current wound healing procedures and potential care, Mater. Sci. Eng. C Mater. Biol. Appl. 48 (2015) 651–662.
- [239] Z. Zhang, L. Qiu, C. Lin, H. Yang, H. Fu, R. Li, Y.J. Kang, Copper-dependent andindependent hypoxia-inducible factor-1 regulation of gene expression, Metallomics 6 (2014) 1889–1893.

- [240] S. Alizadeh, B. Seyedalipour, S. Shafieyan, A. Kheime, P. Mohammadi, N. Aghdami, Copper nanoparticles promote rapid wound healing in acute full thickness defect via acceleration of skin cell migration, proliferation, and neovascularization, Biochem. Biophys. Res. Commun. 517 (2019) 684–690.
- [241] T.L. Goerne, A. Arévalo, P. Ramírez, P. Larraza, Copper nanoparticles as treatment of diabetic foot ulcers: a case report, Glob. Adv. Res. J. Med. Med. Sci. 6 (2017) 267–270.
- [242] J. Xiao, Y. Zhu, S. Huddleston, P. Li, B. Xiao, O.K. Farha, G.A. Ameer, Copper metal-organic framework nanoparticles stabilized with folic acid improve wound healing in diabetes, ACS Nano 12 (2018) 1023–1032.
- [243] P. Bhadauriya, H. Mamtani, M. Ashfaq, A. Raghav, A.K. Teotia, A. Kumar, N. Verma, Synthesis of yeast-immobilized and copper nanoparticle-dispersed carbon nanofiber-based diabetic wound dressing material: simultaneous control of glucose and bacterial infections, ACS Appl. Bio Mater. 1 (2018) 246–258.
- [244] K.P. Singh, D. Latha, A. Jayasree, V.M. Laxmi, T. Selvi, S. Vijayalakshmi, R. Jevanand, Study on efficacy of herbal and copper nanoparticle ointment against wound pathogen isolated from diabetic foot ulcer, J. Pharm. Res. 12 (2018) 516.
- [245] P. Zhang, Y. Li, Y. Tang, H. Shen, J. Li, Z. Yi, Q. Ke, H. Xu, Copper-based metal-organic framework as a controllable nitric oxide-releasing vehicle for enhanced diabetic wound healing, ACS Appl. Mater. Interfaces 12 (2020) 18319–18331
- [246] T. Liu, B. Xiao, F. Xiang, J. Tan, Z. Chen, X. Zhang, C. Wu, Z. Mao, G. Luo, X. Chen, Ultrasmall copper-based nanoparticles for reactive oxygen species scavenging and alleviation of inflammation related diseases, Nat. Commun. 11 (2020) 1–16
- [247] S. Chigurupati, M.R. Mughal, E. Okun, S. Das, A. Kumar, M. McCaffery, S. Seal, M. P. Mattson, Effects of cerium oxide nanoparticles on the growth of keratinocytes, fibroblasts and vascular endothelial cells in cutaneous wound healing, Biomater 34 (2013) 2194–2201.
- [248] C. Zgheib, S.A. Hilton, L.C. Dewberry, M.M. Hodges, S. Ghatak, J. Xu, S. Singh, S. Roy, C.K. Sen, S. Seal, Use of cerium oxide nanoparticles conjugated with microRNA-146a to correct the diabetic wound healing impairment, J. Am. Coll. Surg. 228 (2019) 107–115.
- [249] R. Augustine, A. Hasan, N.K. Patan, Y.B. Dalvi, R. Varghese, A. Antony, R.N. Unni, N. Sandhyarani, A.-E.A. Moustafa, Cerium oxide nanoparticle incorporated electrospun poly (3-hydroxybutyrate-co-3-hydroxyvalerate) membranes for diabetic wound healing applications, ACS Biomater. Sci. Eng. 6 (2019) 58–70.
- [250] N. Kobyliak, L. Abenavoli, L. Kononenko, D. Kyriienko, M. Spivak, Neuropathic diabetic foot ulcers treated with cerium dioxide nanoparticles: a case report, Diabetes Metabolic Syndrome, Clin. Res. Rev. 13 (2019) 228–234.
- [251] G. Sener, S.A. Hilton, M.J. Osmond, C. Zgheib, J.P. Newsom, L. Dewberry, S. Singh, T.S. Sakthivel, S. Seal, K.W. Liechty, Injectable, self-healable zwitterionic cryogels with sustained microRNA-cerium oxide nanoparticle release promote accelerated wound healing. Acta Biomater. 101 (2020) 262–272.
- [252] S.M. Niemiec, A.E. Louiselle, S.A. Hilton, L.C. Dewberry, L. Zhang, M. Azeltine, J. Xu, S. Singh, T.S. Sakthivel, S. Seal, K. Liechty, M. Krebs, Nanosilk increases the strength of diabetic skin and delivers CNP-miR146a to improve wound healing, Front. Immunol. 11 (2020) 262–272.
- [253] P.-H. Lin, M. Sermersheim, H. Li, P.H. Lee, S.M. Steinberg, J. Ma, Zinc in wound healing modulation, Nutrition 10 (2018) 16.
- [254] K. Steffy, G. Shanthi, A.S. Maroky, S. Selvakumar, Enhanced antibacterial effects of green synthesized ZnO NPs using Aristolochia indica against Multi-drug resistant bacterial pathogens from Diabetic Foot Ulcer, J. Infect. Public Health 11 (2018) 463–471.
- [255] M. Ezealisiji Kenneth, O. Chidiebere, X. Siwe-Noudou, Novel zinc oxide nanohydrogel and mucin Co-formulation for diabetic wound healing: a Synergistic approach, Eu. J. Pharm. Med. Res. 7 (2020) 93–100.
- [256] D. Liu, L. Liu, L. Yao, X. Peng, Y. Li, T. Jiang, H. Kuang, Ynthesis of ZnO nanoparticles using radish root extract for effective wound dressing agents for diabetic foot ulcers in nursing care, J. Drug Deliv. Sci. Technol. 55 (2020), 101364.
- [257] H. Hu, Y. Tang, L. Pang, C. Lin, W. Huang, D. Wang, W. Jia, Angiogenesis and full-thickness wound healing efficiency of a copper-doped borate bioactive glass/poly (lactic-co-glycolic acid) dressing loaded with vitamin E in vivo and in vitro, ACS Appl. Mater. Interfaces 10 (2018) 22939–22950.
- [258] P. Losi, E. Briganti, C. Errico, A. Lisella, E. Sanguinetti, F. Chiellini, G. Soldani, Fibrin-based scaffold incorporating VEGF-and bFGF-loaded nanoparticles stimulates wound healing in diabetic mice, Acta Biomater. 9 (2013) 7814–7821.
- [259] K.K. Chereddy, A. Lopes, S. Koussoroplis, V. Payen, C. Moia, H. Zhu, P. Sonveaux, P. Carmeliet, A. des Rieux, G. Vandermeulen, Combined effects of PLGA and vascular endothelial growth factor promote the healing of non-diabetic and diabetic wounds, Nanomed, Nanotechnol. Biol. Med. 11 (2015) 1975–1984.
- [260] U. Bairagi, P. Mittal, J. Singh, B. Mishra, Preparation, characterization, and in vivo evaluation of nano formulations of ferulic acid in diabetic wound healing, Drug Dev. Ind. Pharm. 44 (2018) 1783–1796.
- [261] D.H. Abdelkader, M.M. Tambuwala, C.A. Mitchell, M.A. Osman, S.A. El-Gizawy, A.M. Faheem, M. El-Tanani, P.D. McCarron, Enhanced cutaneous wound healing in rats following topical delivery of insulin-loaded nanoparticles embedded in poly (vinyl alcohol)-borate hydrogels, Drug Delv. Trans. Res. 8 (2018) 1053–1065.
- [262] N. Hasan, J. Cao, J. Lee, M. Naeem, S.P. Hlaing, J. Kim, Y. Jung, B.-L. Lee, J.-W. Yoo, PEI/NONOates-doped PLGA nanoparticles for eradicating methicillin-resistant Staphylococcus aureus biofilm in diabetic wounds via binding to the biofilm matrix, Mater. Sci. Eng. C Mater. Biol. Appl. 103 (2019), 109741.

- [263] K. Gourishetti, R. Keni, P.G. Nayak, S.R. Jitta, N.A. Bhaskaran, L. Kumar, N. Kumar, N. Krishnadas, R. Shenoy, Sesamol-loaded PLGA nanosuspension for accelerating wound healing in diabetic foot ulcer in rats, Int. J. Nanomed. 15 (2020) 9265–9282.
- [264] D.M. Malini, Y.P. Kuntana, M. Madihah, W. Furqon, W. Hermawan, Treatment of PLGA nanoparticles ointment-ethanol extract of Archidendron pauciflorum in the wound healing in diabetic mice, J. Biodjati. 5 (2020) 214–222.
- [265] M. Yu, J. Huang, T. Zhu, J. Lu, J. Liu, X. Li, X. Yan, F. Liu, Liraglutide-loaded PLGA/gelatin electrospun nanofibrous mats promote angiogenesis to accelerate diabetic wound healing via the modulation of miR-29b-3p, Biomater. Sci. 8 (2020) 4225–4238.
- [266] K. Matthews, H. Stevens, A. Auffret, M. Humphrey, G. Eccleston, Lyophilised wafers as a drug delivery system for wound healing containing methylcellulose as a viscosity modifier, Int. J. Pharm. 289 (2005) 51–62.
- [267] A. Ahmed, G. Getti, J. Boateng, Ciprofloxacin-loaded calcium alginate wafers prepared by freeze-drying technique for potential healing of chronic diabetic foot ulcers, J. Drug Delv. Trans. Res. 8 (2018) 1751–1768.
- [268] N.M. Atia, H.A. Hazzah, P.M. Gaafar, O.Y. Abdallah, Diosmin nanocrystal-loaded wafers for treatment of diabetic ulcer: in vitro and in vivo evaluation, J. Pharmacol. Sci. 108 (2019) 1857–1871.
- [269] C. Sandoval-Yañez, C. Castro Rodriguez, Dendrimers: amazing platforms for bioactive molecule delivery systems, Mater 13 (2020) 570.
- [270] A.A. Chis, C. Dobrea, C. Morgovan, A.M. Arseniu, L.L. Rus, A. Butuca, A. M. Juncan, M. Totan, A.L. Vonica-Tincu, G. Cormos, Applications and limitations of dendrimers in biomedicine, Mol 25 (2020) 3982.
- [271] M.J. Kwon, S. An, S. Choi, K. Nam, H.S. Jung, C.S. Yoon, J.H. Ko, H.J. Jun, T. K. Kim, S. Jung, Effective healing of diabetic skin wounds by using nonviral gene therapy based on minicircle vascular endothelial growth factor DNA and a cationic dendrimer, J. Gene Med. 14 (2012) 272–278.
- [272] X. Deng, X. Li, W. Chen, T. Zhao, W. Huang, H. Qian, Design, synthesis and biological evaluation of peptide dendrimers with wound healing promoting activity, Med. Chem. Res. 26 (2017) 580–586.
- [273] S. Hussain, Z. Ji, A.J. Taylor, L.M. DeGraff, M. George, C.J. Tucker, C.H. Chang, R. Li, J.C. Bonner, S. Garantziotis, Multiwalled carbon nanotube functionalization with high molecular weight hyaluronan significantly reduces pulmonary injury, ACS Nano 10 (2016) 7675–7688.
- [274] A. Singh, R. Bhattacharya, A. Shakeel, A.K. Sharma, S. Jeevanandham, A. Kumar, S. Chattopadhyay, H.B. Bohidar, S. Ghosh, S. Chakrabarti, Hydrogel nanotubes with ice helices as exotic nanostructures for diabetic wound healing, Mater. Horiz. 6 (2019) 274–284.
- [275] C. Chaudhary, T. Garg, Scaffolds: a novel carrier and potential wound healer, Crit. Rev. Ther. Drug Carrier Syst. 32 (2015) 277–321.
- [276] Y. Yang, A.C. Ritchie, N.M. Everitt, Comparison of glutaraldehyde and procyanidin cross-linked scaffolds for soft tissue engineering, Mater. Sci. Eng. C Mater. Biol. Appl. 80 (2017) 263–273.
- [277] F. Baino, G. Novajra, V. Brovarone, Bioeng. Biotechnol. 3 (2015) 202.
- [278] K.J. Rambhia, P.X. Ma, J. Contr. Release 219 (2015) 119–128.
- [279] L. Roseti, V. Parisi, M. Petretta, C. Cavallo, G. Desando, I. Bartolotti, B.J.M. S. Grigolo, E. C Mater. Sci. Eng. C Mater. Biol. Appl. 78 (2017) 1246–1262.
- [280] Q. Tan, B. Chen, X. Yan, Y. Lin, Z. Xiao, X. Hou, J. Dai, Bioceramics and scaffolds: a winning combination for tissue engineering, J. Tissue Eng. Regen. Med. 8 (2014) 195–201.
- [281] C. H Lee, S. H Chang, W. J Chen, K. C Hung, Y. H Lin, S. J Liu, M. J Hsieh, J.H. S Pang, J. H Juang, Augmentation of diabetic wound healing and enhancement of collagen content using nanofibrous glucophage-loaded collagen/PLGA scaffold membranes. J. Colloid Interface Sci. 439 (2015) 88–97.
- [282] J.R. Martin, C.E. Nelson, M.K. Gupta, F. Yu, S.M. Sarett, K.M. Hocking, A. C. Pollins, L.B. Nanney, J.M. Davidson, S. Guelcher, C. Duvall, Local delivery of PHD2 siRNA from ROS-degradable scaffolds to promote diabetic wound healing, Adv. Healthc. Mater. 5 (2016) 2751–2757.
- [283] M.S. Lord, A.L. Ellis, B.L. Farrugia, J.M. Whitelock, H. Grenett, C. Li, R. L. O'Grady, A.A. DeCarlo, Erlecan and vascular endothelial growth factor-encoding DNA-loaded chitosan scaffolds promote angiogenesis and wound healing, J. Contr. Release 250 (2017) 48–61.
- [284] C. Dwivedi, I. Pandey, H. Pandey, S. Patil, S.B. Mishra, A.C. Pandey, P. Zamboni, P.W. Ramteke, A.V. Singh, In vivo diabetic wound healing with nanofibrous scaffolds modified with gentamicin and recombinant human epidermal growth factor, J. Biomed. Mater. Res. 106 (2018) 641–651.
- [285] J. Ishihara, A. Ishihara, K. Fukunaga, K. Sasaki, M.J. White, P.S. Briquez, J. A. Hubbell, Laminin heparin-binding peptides bind to several growth factors and enhance diabetic wound healing, Nat. Commun. 9 (2018) 1–14.
- [286] P.K. Veerasubramanian, P. Thangavel, R. Kannan, S. Chakraborty, B. Ramachandran, L. Suguna, V. Muthuvijayan, An investigation of konjac glucomannan-keratin hydrogel scaffold loaded with Avena sativa extracts for diabetic wound healing, Colloids Surf. B Biointerfaces 165 (2018) 92–102.
- [287] T. Ponrasu, P.K. Veerasubramanian, R. Kannan, S. Gopika, L. Suguna, V. J. Muthuvijayan, Morin incorporated polysaccharide-protein (psyllium-keratin) hydrogel scaffolds accelerate diabetic wound healing in Wistar rats, RSC Adv. 8 (2018) 2305–2314.
- [288] J. Chu, P. Shi, W. Yan, J. Fu, Z. Yang, C. He, X. Deng, H. Liu, PEGylated graphene oxide-mediated quercetin-modified collagen hybrid scaffold for enhancement of MSCs differentiation potential and diabetic wound healing, Nanoscale 10 (2018) 9547–9560.
- [289] C.G. Elliott, J. Wang, J.T. Walker, S. Michelsons, J. Dunmore-Buyze, M. Drangova, A. Leask, D.W. Hamilton, Periostin and CCN2 scaffolds promote the

- wound healing response in the skin of diabetic mice, Tissue Eng. 25 (2019) 1326-1339.
- [290] R. Kamel, R. El-Batanony, A.J. Salama, Pioglitazone-loaded three-dimensional composite polymeric scaffolds: a proof of concept study in wounded diabetic rats, Int. J. Pharm. 570 (2019), 118667.
- [291] B.Y. Yang, C. H Hu, W.C. Huang, C.Y. Ho, C. H Yao, C. Huang, Effects of bilayer nanofibrous scaffolds containing curcumin/lithospermi radix extract on wound healing in streptozotocin-induced diabetic rats, Polym 11 (2019) 1745.
- [292] Y. Han, Y. Jiang, Y. Li, M. Wang, T. Fan, M. Liu, Q. Ke, H. Xu, Z. Yi, An aligned porous electrospun fibrous scaffold with embedded asiatic acid for accelerating diabetic wound healin, J. Mater. Chem. B 7 (2019) 6125–6138.
- [293] W. Wan, F. Cai, J. Huang, S. Chen, Q. Liao, A skin-inspired 3D bilayer scaffold enhances granulation tissue formation and anti-infection for diabetic wound healing, J. Mater. Chem. B 7 (2019) 2954–2961.
- [294] J. Fu, Y. Zhang, J. Chu, X. Wang, W. Yan, Q. Zhang, H. Liu, Reduced graphene oxide incorporated acellular dermal composite scaffold enables efficient local delivery of mesenchymal stem cells for accelerating diabetic wound healing, ACS Biomater. Sci. Eng. 5 (2019) 4054–4066.
- [295] Y. Jiang, Y. Han, J. Wang, F. Lv, Z. Yi, Q. Ke, H. Xu, Space-oriented nanofibrous scaffold with silicon-doped amorphous calcium phosphate nanocoating for diabetic wound healing, ACS Appl. Bio Mater. (2) (2019) 787–795.
- [296] G. Long, D. Liu, X. He, Y. Shen, Y. Zhao, X. Hou, B. Chen, W. OuYang, J. Dai, X. Li, A dual functional collagen scaffold coordinates angiogenesis and inflammation for diabetic wound healing, Biomater. Sci. 8 (2020) 6337–6349.
- [297] P. Khazaeli, M. Alaei, M. Khaksarihadad, M. Ranjbar, Preparation of PLA/ chitosan nanoscaffolds containing cod liver oil and experimental diabetic wound healing in male rats study, J. Nanobiotechnol. 18 (2020) 1–9.
- [298] Y. Wang, T. Ying, J. Li, Y. Xu, R. Wang, Q. Ke, S.G. Shen, H. Xu, K. Lin, Hierarchical micro/nanofibrous scaffolds incorporated with curcumin and zinc ion eutectic metal organic frameworks for enhanced diabetic wound healing via anti-oxidant and anti-inflammatory activities, Chem. Eng. J. 402 (2020), 126273.
- [299] C.-H. Yao, K.-Y. Chen, M.-H. Cheng, Y.-S. Chen, C.-H. Huang, Effect of genipin crosslinked chitosan scaffolds containing SDF-1 on wound healing in a rat model, Mater. Sci. Eng. C 109 (2020), 110368.
- [300] A. Vijayan, C. Nanditha, G. Kumar, ECM-mimicking nanofibrous scaffold enriched with dual growth factor carrying nanoparticles for diabetic wound healing, Nanoscale Adv. 3 (2021) 3085–3092.
- [301] C. H Lee, K. C Hung, M. J Hsieh, S. H Chang, J. H Juang, I. C Hsieh, M.S. Wen, S. J Liu, Core-shell insulin-loaded nanofibrous scaffolds for repairing diabetic wounds, Biology, Nanomed. Nanotechnol. Biol. Med. 24 (2020), 102123.
- [302] Y. Jiang, Y. Li, J. Li, Y. Han, P. Zhang, Z. Yi, Q. Ke, H. Xu, A mussel-inspired extracellular matrix-mimicking composite scaffold for diabetic wound healing, ACS Appl. Bio Mater. 3 (2020) 4052–4061.
- [303] S. Chen, H. Wang, Y. Su, J.V. John, A. McCarthy, S.L. Wong, J. Xie, Mesenchymal stem cell-laden, personalized 3D scaffolds with controlled structure and fiber alignment promote diabetic wound healing, Acta Biomater. 108 (2020) 153–167.
- [304] S. Ding, Y. Xu, X. Yan, Y. Lin, Q. Tan, Effect of collagen scaffold with Bcl-2-modified adipose-derived stem cells on diabetic mice wound healing, Int. J. Low. Extrem. Wounds 19 (2020) 139–147.
- [305] H.y. Yang, F. Fierro, M. So, D.J. Yoon, A.V. Nguyen, A. Gallegos, M.D. Bagood, T. Rojo-Castro, A. Alex, H. Stewart, Combination product of dermal matrix, human mesenchymal stem cells, and timolol promotes diabetic wound healing in mice. Stem Cells Transl. Med. 9 (2020) 1353–1364.
- [306] P. Losi, T. Al Kayal, M. Buscemi, I. Foffa, A. Cavallo, G. Soldani, Bilayered fibrin-based electrospun-Sprayed scaffold loaded with platelet lysate enhances wound healing in a diabetic mouse model, Nanomaterials 10 (2020) 2128.
- [307] F.-C. Chao, M.-H. Wu, L.-C. Chen, H.-L. Lin, D.-Z. Liu, H.-O. Ho, M.-T. Sheu, Preparation and characterization of chemically TEMPO-oxidized and mechanically disintegrated sacchachitin nanofibers (SCNF) for enhanced diabetic wound healing, Carbohydr. Polym. 229 (2020), 115507.
- [308] C. Sanhueza, J. Hermosilla, A. Bugallo-Casal, A. Da Silva-Candal, C. Taboada, R. Millán, A. Concheiro, C. Alvarez-Lorenzo, F.J.M.S. Acevedo, One-step electrospun scaffold of dual-sized gelatin/poly-3-hydroxybutyrate nano/microfibers for skin regeneration in diabetic wound, Mater. Sci. Eng. C Mater. Biol. Appl. 119 (2021), 111602.
- [309] S. Chen, B. Liu, M.A. Carlson, A.F. Gombart, D.A. Reilly, J. Xie, O, Recent advances in electrospun nanofibers for wound healing, Nanomed 12 (2017) 1335–1352.
- [310] J. Fang, H. Niu, T. Lin, X. Wang, Applications of electrospun nanofibers, Chin. Sci. Bull. 53 (2008) 2265–2286.
- [311] R. Vasita, D. Katti, Nanofibers and their applications in tissue engineering, Int. J. Nanomed. 1 (2006) 15–30.
- [312] R. Khajavi, M. Abbasipour, A. Bahador, Electrospun biodegradable nanofibers scaffolds for bone tissue engineering, J. Appl. Polym. Sci. 133 (2016), 42883.
- [313] J.S. Choi, K.W. Leong, H. Yoo, In vivo wound healing of diabetic ulcers using electrospun nanofibers immobilized with human epidermal growth factor (EGF), Biomater 29 (2008) 587–596.
- [314] G. Jonathan, W. Shaun, T. Lu, T. Cato, F. Alex, S. Lakshmi, Curcumin loaded poly (E-caprolactone) nanofibers: diabetic wound dressing with antioxidant and antiinflammatory properties, Clin. Exp. Pharmacol. Physiol. 36 (2009) 1149–1156.
- [315] S.S. Scherer, G. Pietramaggiori, J. Matthews, S. Perry, A. Assmann, A. Carothers, M. Demcheva, R.C. Muise-Helmericks, A. Seth, J. Vournakis, Poly-N-acetyl glucosamine nanofibers: a new bioactive material to enhance diabetic wound healing by cell migration and angiogenesis, Ann. Surg. 250 (2009) 322–330.

- [316] J.S. Choi, S.H. Choi, H.S. Yoo, Coaxial electrospun nanofibers for treatment of diabetic ulcers with binary release of multiple growth factors, J. Mater. Chem. 21 (2011) 5258–5267.
- [317] H.-J. Lai, C.-H. Kuan, H.-C. Wu, J.-C. Tsai, T.-M. Chen, D.-J. Hsieh, T.-W. Wang, Tailored design of electrospun composite nanofibers with staged release of multiple angiogenic growth factors for chronic wound healing, Acta Biomater. 10 (2014) 4156–4166.
- [318] T.I. Shalaby, N.M. Fekry, A.S.E. Sodfy, A.G.E. Sheredy, M. Moustafa, Preparation and characterisation of antibacterial silver-containing nanofibres for wound healing in diabetic mice, Int. J. Nanoparticles (IJNP) 8 (2015) 82–98.
- [319] M. Ranjbar-Mohammadi, S. Rabbani, S.H. Bahrami, M. Joghataei, F. Moayer, Antibacterial performance and in vivo diabetic wound healing of curcumin loaded gum tragacanth/poly (ε-caprolactone) electrospun nanofiber, Mater. Sci. Eng. C Mater. Biol. Appl. 69 (2016) 1183–1191.
- [320] B. Senturk, S. Mercan, T. Delibasi, M.O. Guler, A. Tekinay, Angiogenic peptide nanofibers improve wound healing in STZ-induced diabetic rats, ACS Biomater. Sci. Eng. 2 (2016) 1180–1189.
- [321] J. Grip, R.E. Engstad, I. Skjæveland, N. Škalko-Basnet, J. Isaksson, P. Basnet, A. Holsæter, Beta-glucan-loaded nanofiber dressing improves wound healing in diabetic mice, Eur. J. Pharmaceut. Sci. 121 (2018) 269–280.
- [322] Z. Zheng, Y. Liu, W. Huang, Y. Mo, Y. Lan, R. Guo, B. Cheng, Neurotensin-loaded PLGA/CNC composite nanofiber membranes accelerate diabetic wound healing, Artif. Cell Nanomed. Biotechnol. 46 (2018) 493–501.
- [323] S. Kanji, M. Das, M. Joseph, R. Aggarwal, S.M. Sharma, M. Ostrowski, V. J. Pompili, H.-Q. Mao, H. Das, Nanofiber-expanded human CD34+ cells heal cutaneous wounds in streptozotocin-induced diabetic mice, Sci. Rep. 9 (2019) 1–11
- [324] M.E. Cam, M. Crabbe-Mann, H. Alenezi, A.N. Hazar-Yavuz, B. Ertas, C. Ekentok, G.S. Ozcan, F. Topal, E. Guler, Y. Yazir, The comparision of glybenclamide and metformin-loaded bacterial cellulose/gelatin nanofibres produced by a portable electrohydrodynamic gun for diabetic wound healing, Eur. Polym. J. 134 (2020), 109844.
- [325] F. Liu, X. Li, L. Wang, X. Yan, D. Ma, Z. Liu, X. Liu, Sesamol incorporated cellulose acetate-zein composite nanofiber membrane: an efficient strategy to accelerate diabetic wound healing, Int. J. Biol. Macromol. 149 (2020) 627–638.
- [326] H. Samadian, S. Zamiri, A. Ehterami, S. Farzamfar, A. Vaez, H. Khastar, M. Alam, A. Ai, H. Derakhshankhah, Z. Allahyari, Electrospun cellulose acetate/gelatin nanofibrous wound dressing containing berberine for diabetic foot ulcer healing: in vitro and in vivo studies, Sci. Rep. 10 (2020) 1–12.
- [327] A. Almasian, F. Najafi, M. Eftekhari, M.R.S. Ardekani, M. Sharifzadeh, M. Khanavi, Polyurethane/carboxymethylcellulose nanofibers containing Malva sylvestris extract for healing diabetic wounds: preparation, characterization, in vitro and in vivo studies, Mater. Sci. Eng. C Mater. Biol. Appl. 114 (2020), 111039.
- [328] M. Zehra, W. Zubairi, A. Hasan, H. Butt, A. Ramzan, M. Azam, A. Mehmood, M. Falahati, A.A. Chaudhry, I. Rehman, Oxygen generating polymeric nano fibers that stimulate angiogenesis and show efficient wound healing in a diabetic wound model. Int. J. Nanomed. 15 (2020) 3511.
- [329] Z. Ding, Y. Zhang, P. Guo, T. Duan, W. Cheng, Y. Guo, X. Zheng, G. Lu, Q. Lu, D. Kaplan, Injectable desferrioxamine-laden silk nanofiber hydrogels for accelerating diabetic wound healing, ACS Biomater. Sci. Eng. 7 (2021) 1147–1158.
- [330] A. Razzaq, Z. Khan, A. Saeed, K. Shah, N. Khan, B. Menaa, H. Iqbal, F. Menaa, Development of cephradine-loaded gelatin/polyvinyl alcohol electrospun nanofibers for effective diabetic wound healing: in-vitro and in-vivo assessments, Pharm. Times 13 (2021) 349.
- [331] Y. Pu, P. Wang, Y. Rong, X. Tan, T. Shi, J. Ma, W. Xue, B. Chi, Bio-fabricated nanocomposite hydrogel with ROS scavenging and local oxygenation accelerates diabetic wounds healing, J. Mater. Chem. B 10 (21) (2022) 4083–4095, https:// doi.org/10.1039/d2tb00343k.
- [332] P.V.K. Kumari, Y.S. Rao, S. Akhila, Role of nanocomposites in drug delivery, Biol. Pharm. Sci. 8 (2019) 94–103.
- [333] A. Montaser, A. Abdel-Mohsen, M. Ramadan, A. Sleem, N. Sahffie, J. Jancar, A. Hebeish, Preparation and characterization of alginate/silver/nicotinamide nanocomposites for treating diabetic wounds, Int. J. Biol. Macromol. 92 (2016) 739–747.
- [334] J. Li, F. Lv, H. Xu, Y. Zhang, J. Wang, Z. Yi, J. Yin, J. Chang, C. Wu, A patterned nanocomposite membrane for high-efficiency healing of diabetic wound, J. Mater. Chem. B 5 (2017) 1926–1934.
- [335] P. Thangavel, R. Kannan, B. Ramachandran, G. Moorthy, L. Suguna, V. Muthuvijayan, Development of reduced graphene oxide (rGO)-isabgol nanocomposite dressings for enhanced vascularization and accelerated wound healing in normal and diabetic rats, J. Colloid Interface Sci. 517 (2018) 251–264.
- [336] M. Das, U. Goswami, R. Kandimalla, S. Kalita, S.S. Ghosh, A. Chattopadhyay, Iron-copper bimetallic nanocomposite reinforced dressing materials for infection control and healing of diabetic wound, ACS Appl. Bio Mater. 2 (2019) 5434–5445.
- [337] Y. Li, T. Xu, Z. Tu, W. Dai, Y. Xue, C. Tang, W. Gao, C. Mao, B. Lei, C. Lin, Bioactive antibacterial silica-based nanocomposites hydrogel scaffolds with high angiogenesis for promoting diabetic wound healing and skin repair, Theranostic 10 (2020) 4929–4943.
- [338] S. Tavakoli, H. Mokhtari, M. Kharaziha, A. Kermanpur, A. Talebi, J. Moshtaghian, A multifunctional nanocomposite spray dressing of Kappa-carrageenanpolydopamine modified ZnO/L-glutamic acid for diabetic wounds, Mater. Sci. Eng. C Mater. Biol. Appl. 111 (2020), 110837.

- [339] D.L. Steed, J.B. Goslen, G.A. Holloway, J.M. Malone, T. Bunt, M. Webster, Randomized prospective double-blind trial in healing chronic diabetic foot ulcers: CT-102 activated platelet supernatant, topical versus placebo, Diabetes Care 15 (1992) 1598–1604.
- [340] M.C. Robson, T.J. Phillips, V. Falanga, D.J. Odenheimer, L.C. Parish, J.L. Jensen, D. Steed, Randomized trial of topically applied repifermin (recombinant human keratinocyte growth factor-2) to accelerate wound healing in venous ulcers, Wound Repair Regen. 9 (2001) 347–352.
- [341] S. Eghbali, F.A. Sayyedeh, A. Razieh, S. Amirhossein, Therapeutic effects of Punica granatum (pomegranate): an updated review of clinical trials, J. Nutri. Metabol. (2021) 2021.
- [342] J.A. Niezgoda, C.C. Van Gils, R.G. Frykberg, J.P. Hodde, Randomized clinical trial comparing OASIS Wound Matrix to Regranex Gel for diabetic ulcers, Adv. Skin Wound Care 18 (2005) 258–266.
- [343] Y. Kusumanto, V. Van Weel, N. Mulder, A. Smit, J. Van Den Dungen, J. Hooymans, W. Sluiter, R. Tio, P. Quax, R. Gans, Treatment with intramuscular vascular endothelial growth factor gene compared with placebo for patients with diabetes mellitus and critical limb ischemia: a double-blind randomized trial, Hum. Gene Ther. 17 (2006) 683-691.
- [344] V.R. Driver, J. Hanft, C.P. Fylling, J. Beriou, A prospective, randomized, controlled trial of autologous platelet-rich plasma gel for the treatment of diabetic foot ulcers, Ostomy/Wound Manag. 52 (2006) 68–70.
- [345] S.Y. Silva, L.C. Rueda, G.A. Márquez, M. López, D.J. Smith, C.A. Calderón, J. C. Castillo, J. Matute, C.F. Rueda-Clausen, A. Orduz, Double blind, randomized, placebo controlled clinical trial for the treatment of diabetic foot ulcers, using a nitric oxide releasing patch:PATHON, Trials 8 (2007) 1–8.
- [346] D.D. Kakagia, K.J. Kazakos, K.C. Xarchas, M. Karanikas, G.S. Georgiadis, G. Tripsiannis, C. Manolas, Synergistic action of protease-modulating matrix and autologous growth factors in healing of diabetic foot ulcers. A prospective randomized trial, J. Diabet. Complicat. 21 (2007) 387–391.
- [347] S. Kordestani, M. Shahrezaee, M. Tahmasebi, H. Hajimahmodi, D.H. Ghasemali, M. Abyaneh, A randomised controlled trial on the effectiveness of an advanced wound dressing used in Iran, J. Wound Care 17 (2008) 323–327.
- [348] M. Rullan, L. Cerdà, G. Frontera, L. Masmiquel, J. Llobera, Treatment of chronic diabetic foot ulcers with bemiparin: a randomized, triple-blind, placebocontrolled, clinical trial, Diabet, Med. 25 (2008) 1090–1095.
- [349] A. Landsman, P.A. Blume, D.A. Jordan Jr., D. Vayser, A. Gutierrez, An open-label, three-arm pilot study of the safety and efficacy of topical Microcyn Rx wound care versus oral levofloxacin versus combined therapy for mild diabetic foot infections, J. Am. Podiatr. Med. Assoc. 101 (2011) 484–496.
- [350] L. Uccioli, L. Giurato, V. Ruotolo, A. Ciavarella, M.S. Grimaldi, A. Piaggesi, I. Teobaldi, L. Ricci, L. Scionti, C. Vermigli, Two-step autologous grafting using HYAFF scaffolds in treating difficult diabetic foot ulcers: results of a multicenter, randomized controlled clinical trial with long-term follow-up, Int. J. Low. Extrem. Wounds 10 (2011) 80–85.
- [351] C.M. Zelen, T.E. Serena, G. Denoziere, D.E. Fetterolf, A prospective randomised comparative parallel study of amniotic membrane wound graft in the management of diabetic foot ulcers, Int. Wound J. 10 (2013) 502–507.
- [352] R. Tanaka, H. Masuda, S. Kato, K. Imagawa, K. Kanabuchi, C. Nakashioya, F. Yoshiba, T. Fukui, R. Ito, M. Kobori, Autologous G-CSF-mobilized peripheral blood CD34+ cell therapy for diabetic patients with chronic nonhealing ulcer, Cell Transplant. 23 (2014) 167-179.
- [353] L. Li, D. Chen, C. Wang, N. Yuan, Y. Wang, L. He, Y. Yang, L. Chen, G. Liu, X. Li, Autologous platelet-rich gel for treatment of diabetic chronic refractory cutaneous ulcers: a prospective, randomized clinical trial, Wound Repair Regen. 23 (2015) 495–505.
- [354] T.A. Motley, A.M. Gilligan, D.L. Lange, C.R. Waycaster, J. Dickerson, Cost-effectiveness of clostridial collagenase ointment on wound closure in patients with diabetic foot ulcers: economic analysis of results from a multicenter, randomized, open-label trial, J. Foot Ankle Res. 8 (2015) 1–12.
- [355] M. Imran, M.B. Hussain, M. Baig, A randomized, controlled clinical trial of honeyimpregnated dressing for treating diabetic foot ulcer, J. Coll. Phy. Surg. Pak. 25 (2015) 721–725.
- [356] S. Cazzell, D. Vayser, H. Pham, J. Walters, A. Reyzelman, B. Samsell, K. Dorsch, M. Moore, A randomized clinical trial of a human acellular dermal matrix demonstrated superior healing rates for chronic diabetic foot ulcers over conventional care and an active acellular dermal matrix comparator, Wound Repair Regen. 25 (2017) 483–497.
- [357] Z. Soleimani, F. Hashemdokht, F. Bahmani, M. Taghizadeh, M.R. Memarzadeh, Z. Asemi, Clinical and metabolic response to flaxseed oil omega-3 fatty acids supplementation in patients with diabetic foot ulcer: a randomized, double-blind, placebo-controlled trial, J. Diabet. Complicat. 31 (2017) 1394–1400.
- [358] M. Heravi, E. Barahimi, R. Razzaghi, F. Bahmani, H.R. Gilasi, Z. Asemi, The effects of zinc supplementation on wound healing and metabolic status in patients with diabetic foot ulcer: a randomized, double-blind, placebo-controlled trial, Wound Repair Regen. 25 (2017) 512–520.
- [359] K.H. Park, S.H. Han, J.P. Hong, S.-K. Han, D.-H. Lee, B.S. Kim, J.H. Ahn, J.W. Lee, Topical epidermal growth factor spray for the treatment of chronic diabetic foot ulcers: a phase III multicenter, double-blind, randomized, placebo-controlled trial, Diabetes Res. Clin. Pract. 142 (2018) 335–344.
- [360] L. Lavery, J. Fulmer, K.A. Shebetka, M. Regulski, D. Vayser, D. Fried, H. Kashefsky, T.M. Owings, J. Nadarajah, Z. Hesp, Open-label extension phase of a chronic diabetic foot ulcer multicenter, controlled, randomized clinical trial using cryopreserved placental membrane, Wounds 30 (2018) 283–289.

- [361] M.A.G. Barbosa, A.O. Paggiaro, V.F. de Carvalho, C. Isaac, R. Gemperli, Effects of hydrogel with enriched sodium alginate in wounds of diabetic patients, Plast. Surg. Nurs. 38 (2018) 133–138.
- [362] H. Sun, H. Lv, F. Qiu, D. Sun, Y. Gao, N. Chen, Y. Zheng, K. Deng, Y. Yang, H. Zhang, Clinical application of a 3D-printed scaffold in chronic wound treatment: a case series, J. Wound Care 27 (2018) 262–271.
- [363] K.-C. Moon, H.-S. Suh, K.-B. Kim, S.-K. Han, K.-W. Young, J.-W. Lee, M.-H. Kim, Potential of allogeneic adipose-derived stem cell–hydrogel complex for treating diabetic foot ulcers, Diabetes 68 (2019) 837–846.
- [364] Y. Najafian, Z.M. Khorasani, M.N. Najafi, S.S. Hamedi, M. Mahjour, Z. Feyzabadi, Efficacy of Aloe vera/Plantago major gel in Diabetic Foot Ulcer: a randomized double-blind clinical trial, Curr. Drug Discov. Technol. 16 (2019) 223–231.
- [365] S. Hamed, M. Belokopytov, Y. Ullmann, M. Safadi, Y. Stark, A. Shoufani, S. Akita, P.Y. Liu, L. Teot, Nterim results of the remede d'Or study: a multicenter, singleblind, randomized, controlled trial to assess the safety and efficacy of an innovative topical formulation of erythropoietin for treating diabetic foot ulcers, Adv. Wound Care 8 (2019) 514–521.
- [366] https://www.clinicaltrials.gov/ct2/show/NCT03998436. Accessed on 11/11/ 2021.
- [367] Q. Yang, Y. Zhang, H. Yin, Y. Lu, Topical recombinant human epidermal growth factor for diabetic foot ulcers: a meta-analysis of randomized controlled clinical trials, Ann. Vasc. Surg. 62 (2020) 442–451.
- [368] T.E. Serena, R. Yaakov, S. Moore, W. Cole, S. Coe, R. Snyder, K. Patel, B. Doner, M.A. Kasper, R. Hamil, A randomized controlled clinical trial of a hypothermically stored amniotic membrane for use in diabetic foot ulcers, J. Comp. Eff. Res. 9 (2020) 23–34.
- [369] J. Chokpaisarn, S. Chusri, S. Voravuthikunchai, Clinical randomized trial of topical Quercus infectoria ethanolic extract for the treatment of chronic diabetic ulcers, J. Herb. Med. 21 (2020), 100301.
- [370] T. Goerne, P. Olivares, L. A Dávalos, J.A. Muñoz, J. González, Catalytic nanomedicine. Cu/TiO2–SiO2 nanoparticles as treatment of diabetic foot ulcer: a case report, Curr. Nanomed. 10 (2020) 290–295.
- [371] B.M. Bonora, R. Cappellari, M. Mazzucato, M. Rigato, M. Grasso, M. Menegolo, A. Bruttocao, A. Avogaro, G. Fadini, Stem cell mobilization with plerixafor and healing of diabetic ischemic wounds: a phase IIa, randomized, double-blind, placebo-controlled trial, Stem Cells Transl. Med. 9 (2020) 965–973.
- [372] E.J. Lullove, B. Liden, C. Winters, P. McEneaney, A. Raphael, J. Li, A multicenter, blinded, randomized controlled clinical trial evaluating the effect of omega-3-rich fish skin in the treatment of chronic, nonresponsive diabetic foot ulcers, Wounds 33 (2021) 169–171.
- [373] https://patents.google.com/patent/ES2600793T3/en. (Accessed 14 November 2021).
- [374] https://europepmc.org/article/PAT/KR20080071385. (Accessed 14 November 2021).
- [375] https://europepmc.org/article/PAT/US2008248005. (Accessed 15 November 2021).
- [376] https://patents.google.com/patent/US7517528B2/en. (Accessed 15 November 2021).
- [377] https://patents.google.com/patent/US20080159978A1/en. (Accessed 15 November 2021).
- [378] https://patents.google.com/patent/EP3100611A1/en. (Accessed 15 November 2021).
- [379] https://patents.justia.com/patent/11160775. (Accessed 16 November 2021).
- [380] https://patents.google.com/patent/CN101287457B/en. (Accessed 16 November 2021).
- [381] https://patents.google.com/patent/US7928280B2/en. (Accessed 16 November 2021).
- [382] https://patents.google.com/patent/US8105580B2/en. (Accessed 16 November 2021).
- [383] https://patents.google.com/patent/CN103041438B/en?oq=CN103041438B. (Accessed 16 November 2021).
- [384] https://patents.google.com/patent/CN103479883B/en?oq=CN103479883B. (Accessed 16 November 2021).
- [385] https://patents.google.com/patent/US9050176B2/en. (Accessed 12 November 2021).
- [386] https://patents.google.com/patent/US9198928. (Accessed 12 November 2021).
- [387] https://patentscope.wipo.int/search/en/detail.jsf?docId=US205824080&docAn=15656088. (Accessed 12 November 2021).

- [388] https://patents.google.com/patent/CN101932330B/en. (Accessed 12 November 2021).
- [389] https://patents.google.com/patent/US9486554B2/en. (Accessed 12 November 2021).
- [390] https://patents.google.com/patent/WO2012151438A1/en. (Accessed 12 November 2021).
- [391] https://patents.google.com/patent/US8063023B2/en. (Accessed 12 November 2021).
- [392] https://patentscope.wipo.int/search/en/detail.jsf?docId=W02014128354. (Accessed 12 November 2021).
- [393] https://patents.google.com/patent/US9717776B2/en. (Accessed 12 November 2021).
- [394] https://patents.google.com/patent/CN102480969A/en. (Accessed 12 November 2021).
- [395] https://patents.google.com/patent/SG10201906716QA/en. (Accessed 12 November 2021).
- [396] https://patents.google.com/patent/WO2014140890A3/el. (Accessed 12 November 2021).
- [397] https://patents.google.com/patent/WO2010091124A3. (Accessed 12 November 2021).
- [398] https://clinicaltrials.gov/ct2/show/NCT00815217. (Accessed 12 November 2021).
- [399] https://clinicaltrials.gov/ct2/show/NCT01342497. (Accessed 12 November 2021).
- [400] https://clinicaltrials.gov/ct2/show/NCT01113515. (Accessed 12 November 2021).
- [401] https://clinicaltrials.gov/ct2/show/NCT02081352. (Accessed 12 November 2021).
- [402] https://clinicaltrials.gov/ct2/show/NCT04205942. (Accessed 12 November 2021).
- [403] https://www.clinicaltrials.gov/ct2/show/NCT03865394. (Accessed 12 November 2021).
- [404] https://clinicaltrials.gov/ct2/show/NCT02581098. (Accessed 12 November 2021).
- [405] https://clinicaltrials.gov/ct2/show/NCT04315909. (Accessed 12 November 2021).
- [406] https://clinicaltrials.gov/ct2/show/NCT03890172. (Accessed 12 November 2021).
- [407] https://clinicaltrials.gov/ct2/show/NCT04093635. (Accessed 12 November 2021).
- [408] https://www.clinicaltrials.gov/ct2/show/NCT04133493. (Accessed 12 November 2021).
- [409] https://www.clinicaltrials.gov/ct2/show/NCT04537520. (Accessed 12 November 2021).
- [410] https://clinicaltrials.gov/ct2/show/NCT03271580. (Accessed 12 November 2021).
- [411] https://clinicaltrials.gov/ct2/show/NCT02313428. (Accessed 12 November 2021).
- [412] https://clinicaltrials.gov/ct2/show/NCT04505670. (Accessed 12 November 2021).
- [413] https://clinicaltrials.gov/ct2/show/NCT03654989. (Accessed 12 November 2021).
- [414] https://www.clinicaltrials.gov/ct2/show/NCT04849143. (Accessed 12 November 2021).
- [415] https://clinicaltrials.gov/ct2/show/NCT03137966. (Accessed 12 November 2021).
- [416] https://www.clinicaltrials.gov/ct2/show/NCT04755647. (Accessed 12 November 2021).
- [417] https://clinicaltrials.gov/ct2/show/NCT04723134. (Accessed 12 November 2021).
- [418] https://clinicaltrials.gov/ct2/show/NCT04709419. (Accessed 12 November 2021).
- [419] https://clinicaltrials.gov/ct2/show/NCT04918784. (Accessed 12 November 2021).
- [420] https://www.clinicaltrials.gov/ct2/show/NCT04689425. (Accessed 12 November 2021).